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Green Gold of the Ocean: Unlocking the Potential of *Caulerpa* in Global Seaweed Markets – A Review

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ABSTRACT

Seaweeds support food, health, and sustainability, yet global production is dominated by red and brown taxa. The green macroalga Caulerpa ("sea grapes"/"green caviar") remains underutilized despite notable nutritional and functional potential. This review synthesizes current knowledge on Caulerpa taxonomy, biology, distribution, cultivation, utilization, and sustainability. As siphonous, coenocytic algae, Caulerpa spp. are prevalent in tropical-subtropical waters, with some lineages invasive outside native ranges. Farming spans traditional lagoon culture in Southeast Asia to modern aquaculture and IMTA. Post-harvest advances (e.g., brining, UV-C, improved packaging) extend shelf life, though quality trade-offs remain. Nutritionally, Caulerpa provides proteins, polysaccharides, PUFAs, minerals, and vitamins; bioactives (e.g., sulfated polysaccharides, caulerpin, caulerpenyne) exhibit antioxidant, anti-inflammatory, and metabolic activities primarily in experimental models. Applications include foods, nutraceuticals, cosmetics, fertilizers/feeds, and bioenergy. Key constraints are short shelf life, variable cultivation standards, limited market acceptance beyond Asia, and ecological/biosecurity risks. Priorities include standardized cultivation and post-harvest protocols, selective breeding, toxicology and exposure assessments, and enabling harmonized policies. With sustainable management, Caulerpa could evolve from niche product to "green gold of the ocean," aligning ecological responsibility with economic opportunity.

INTRODUCTION

The global seaweed industry is experiencing robust growth. The market was valued at approximately USD 17 billion in 2023 and is projected to expand significantly in the







coming decade. According to UNCTAD, emerging seaweed markets, including textiles, biopackaging, and biofuels, represents a potential USD 11.8 billion in additional value by 2030. Academic analyses estimate global seaweed production reached 34–36 million tonnes in 2019 (Janke, 2024). Despite this growth, the sector is dominated by red and brown macroalgae, notably *Eucheuma, Kappaphycus, Gracilaria, Saccharina, Undaria*, and *Pyropia* (Hurtado *et al.*, 2019; Arias-Echeverri *et al.*, 2022; Khan *et al.*, 2024; Mantri *et al.*, 2024). In contrast, green macroalgae (Chlorophyta), including *Caulerpa*, remain underrepresented, constituting less than 1% of global seaweed biomass production (Moreira *et al.*, 2021).

Among green seaweeds, species such as Caulerpa (commonly known as "sea grapes" or "green caviar") stand out for their unique texture and growing regional popularity (Stuthmann et al., 2023). It has attracted attention for its unique sensorial qualities and rich nutritional profile, containing dietary fiber, proteins, polyunsaturated fatty acids, vitamins, and essential minerals such as calcium, magnesium, and iron (du Preez et al., 2020; Zhang et al., 2020a). Traditionally consumed in Southeast Asia and Oceania, Caulerpa is now attracting growing attention in global food markets. Within these regions, eight species have been reported, including C. lentillifera, C. peltata, C. racemosa, C. scalpelliformis, C. serrulata, C. sertularioides, C. taxifolia, and C. verticillata (Abdul Razak et al., 2020; Syakilla et al., 2022; Stuthmann et al., 2023). Recent studies confirm that *Caulerpa* is rich in polyunsaturated fatty acids (PUFAs), minerals, dietary fiber, and various bioactive compounds, including influential antioxidants that confer health-promoting properties (Tanna et al., 2018; Manoppo et al., 2022; Nurkholis et al., 2023), and several bioactive compounds such as sulfated polysaccharides, known for their antioxidant, antihyperglycemic, anticoagulant, and immunestimulatory effects (**Tesvichian** et al., 2024). These properties position it not only as a functional food but also as a promising candidate for nutraceutical formulations. Other species, such as C. racemosa and C. prolifera, are similarly consumed in coastal Asia and the Mediterranean, where they are integrated into traditional diets or valorized for their secondary metabolites, including caulerpenyne and caulerpin, with potential antimicrobial, anticancer, antioxidant, and antidiabetic (Belkacemi et al., 2020; Permatasari et al., 2022; Lau et al., 2024; Ouhabi et al., 2025). Moreover, nutritional profiling across different populations demonstrates notable protein, sugar, and ash content, with variations linked to geographical and environmental factors (Zhou et al., 2025).

Despite this growing evidence, *Caulerpa* remains a niche genus in global seaweed markets. Cultivation is restricted mainly to smallholder farms in Southeast Asia and Japan, often lacking standardized farming methods and robust supply chains. Critical challenges continue to constrain its commercialization, including short post-harvest shelf life, variable quality across regions, and limited consumer awareness outside Asia-Pacific markets (**Estrada** *et al.*, **2021**; **Pan-utai**, **2023**). Furthermore, while bioactive properties

have been identified, comprehensive toxicological assessments, yield and nutrient optimization breeding programs, and systematic post-harvest innovations remain limited. Given these gaps, a comprehensive review is needed to synthesize current understanding of *Caulerpa* across its taxonomy, biology, global distribution, cultivation techniques, nutritional and functional properties, and market prospects. This review highlights the opportunities and ecological trade-offs of expanding *Caulerpa* cultivation, evaluates its role in sustainability and ecosystem services, and identifies future research and policy priorities. By positioning *Caulerpa* as a potential "green gold of the ocean," this paper underscores its untapped role in diversifying global seaweed markets and contributing to climate-smart aquaculture, functional food development, and blue economy innovation.

MATERIALS AND METHODS

This review synthesizes recent knowledge on the taxonomy, biology, cultivation, nutritional and bioactive properties, market trends, and sustainability challenges of *Caulerpa* spp., with emphasis on *C. lentillifera* and *C. racemosa*. We searched Scopus and Web of Science (WoS) for English-language records published during 2015–2025 (last search: 30 September 2025). Searches targeted titles, abstracts, and keywords (Scopus: TITLE-ABS-KEY; WoS: Topic/TS). Core query: "*Caulerpa*" combined with AND/OR terms: "taxonomy," "cultivation," "nutritional composition," "bioactive compounds," "applications," "sustainability," and "market trends". Example: *Caulerpa* AND (taxonomy OR cultivation OR "nutritional composition" OR "bioactive compounds" OR sustainability OR "market trends").

After de-duplication, titles/abstracts were screened to remove (i) studies not centered on *Caulerpa*; (ii) items not indexed in Scopus or WoS; (iii) non-English items; and (iv) non-article/non-review formats (e.g., conference abstracts, editorials, theses). Records lacking experimental data or review-based synthesis relevant to our themes were excluded. Approximately ~100 eligible articles were retained for full-text analysis (Mengist *et al.*, 2020).

Data were extracted into thematic matrices aligned with the review structure and synthesized narratively. All figures (morphology schematics, flowcharts, value-chain and SWOT diagrams, roadmap) were created de novo by the authors from the reviewed sources to ensure clarity and originality.

RESULTS

$Taxonomy, biology, and \ global \ distribution$

Taxonomy and morphology

The genus Caulerpa (Caulerpaceae; Bryopsidales) is a diverse clade of siphonous, coenocytic green macroalgae, each thallus comprising a single multinucleate cell lacking

cross walls. Recent revisions recognize ~104 accepted species arranged across multiple subgenera and sections, reflecting substantial morphological and phylogenetic complexity (**Lagourgue** *et al.*, **2024**). These green algae belong to the order Bryopsidales, characterized by a siphonous, coenocytic structure, wherein each organism consists of a single multinucleate cell without cross walls, allowing them to attain large sizes while maintaining cytoplasmic continuity (**Ranjan** *et al.*, **2015**). The morphology of *Caulerpa* can be seen in Fig. (1).

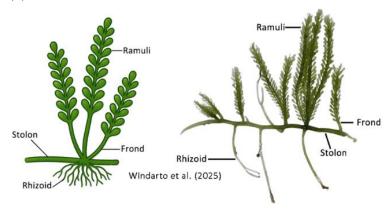


Fig. 1. Morphology of *Caulerpa*

Caulerpa species share a creeping stolon anchored by rhizoids, from which erect assimilatory fronds arise. Fronds typically bear a central axis (rachis) with lateral branchlets (ramuli) whose shapes (e.g., vesiculate, clavate, falcate) and arrangements (e.g., distichous, verticillate) vary among taxa (**Zubia** et al., 2020; **Lagourgue** et al., 2024). In C. lentillifera, the stolon–rhizoid–frond architecture underpins rapid clonal spread and structural integrity (**Arimoto** et al., 2019). Pronounced morphological plasticity, long recognized in the genus, has complicated species delimitation; many ecads (environmentally induced morphotypes) were historically described as distinct forms. Integrative taxonomy that couples morphology with molecular markers continues to refine species boundaries.

Global distribution

Caulerpa species occur throughout tropical to warm-temperate coasts worldwide, with high diversity and abundance across the Indo-Pacific (e.g., Southeast Asia, Pacific Islands, northern Australia) and notable presences in the Mediterranean and Caribbean—West Atlantic regions (Schembri et al., 2015; Lagourgue et al., 2024). C. prolifera is broadly distributed in the Mediterranean, eastern Atlantic, and western Atlantic, including parts of the Americas (Fraissinet et al., 2025).

Several species are established outside their native ranges. *C. taxifolia* spread widely in the Mediterranean and was also detected and subsequently eradicated in coastal California, USA; ongoing management is required in other localities. *C. racemosa* sensu lato, now treated as *C. cylindracea* in the Mediterranean, has expanded extensively in the

basin, illustrating strong dispersal and ecological plasticity (Mannino *et al.*, 2019). Originally Indo-Pacific, *C. brachypus* has been recorded as invasive in Florida (USA), the Caribbean, and New Zealand (Wells & Bieler, 2020). The global distribution of *Caulerpa* can be seen in Fig. (2).



Fig. 2. Global distribution hotspots of *Caulerpa* spp.

Worldwide, there are hotspots of *Caulerpa* distribution and invasiveness, including the Indo-Pacific (Indonesia, Philippines, Japan), Mediterranean Sea (site of *C. taxifolia* invasion), Caribbean Sea, southern Australia, and California. Native ranges highlight *Caulerpa*'s prevalence in tropical and subtropical ecosystems, while invasion records underscore its ecological risks.

Ecological significance

Caulerpa plays dual ecological roles, both beneficial and potentially detrimental. The thriving benthic macroalgae contribute to habitat complexity, substrate stabilization, and nutrient cycling, supporting biodiverse marine communities. Their rapid vegetative propagation through fragmentation enables them to recolonize disturbed habitats efficiently (Herawati et al., 2021; Parreira et al., 2021; Fraissinet et al., 2025).

Conversely, *C. taxifolia* and *C. cylindracea* (*C. racemosa*) exhibit invasive behavior when introduced into non-native ecosystems with minimal predator pressure. Their invasiveness can lead to ecological disruptions, including the displacement of native seagrasses and alterations of benthic habitats, especially in the Mediterranean, where habitat and environmental balance are particularly sensitive (Montefalcone *et al.*, 2015; Mannino *et al.*, 2019).

Cultivation and processing Traditional farming techniques

In the Indo-Pacific region, especially in the Philippines and Indonesia, *Caulerpa* species have been traditionally harvested from the wild. Still, over time, shallow pond and lagoon farming have been adopted to support local communities. Techniques such as bottom-planting, off-bottom culture, floating long lines, and land-based raceways (Fig. 3) are used to cultivate *C. lentillifera* ("sea grapes") for fresh consumption, capitalizing on its high demand for local and export markets (**Islam** *et al.*, **2023**).

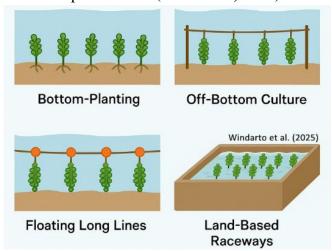


Fig. 3. Farming techniques of *Caulerpa* cultivation

Modern techniques

Efforts to integrate *C. lentillifera* into modern integrated multi-trophic aquaculture (IMTA) systems, where seaweeds serve as extractive species to utilize nutrients from fed organisms, have shown mixed success. In one trial in Southern Cebu, baskets in openwater IMTA failed to support *C. lentillifera* effectively, highlighting challenges in adapting the species to such systems (Largo *et al.*, 2016). Meanwhile, broader developments in IMTA models, particularly in land-based pond systems, demonstrate high nutrient removal efficiencies, reinforcing the potential to include *Caulerpa* as a biofilter in these eco-friendly aquaculture models (Yang *et al.*, 2025).

Beyond biofiltration, *C. lentillifera* can act as a productive substrate that upgrades feed quality. In co-culture, the 60 g m⁻² substrate level yielded the best outcomes for L. vannamei fed *P. pacifica* grown on that substrate: RGR 34.98%, length gain 1.97 cm, biomass 0.0104 g (per individual), survival 96%, grazing rate 23.034 ind. day⁻¹; the feed contained 62.15% protein, 8.04% fat, EPA 7.95%, and lysine 11.95% (**Herawati** *et al.*, **2021**). These results position *C. lentillifera* as both a controllable nutrient scrubber in land-based IMTA and a substrate that enriches intermediate trophic culture for shrimp grow-out.

Recent trials indicate that low-cost, plant-derived PGRs can enhance *C. racemosa* growth and quality under culture. Combinations of coconut water (*Cocos nucifera*) and

shallot crude extract (Allium cepa L.) have been reported to stimulate stolon extension, ramuli density, and overall biomass (Ghani et al., 2023). Additional studies using natural PGRs similarly show gains in specific growth rate and selected nutritional attributes (e.g. proximate composition) for C. racemosa (Windarto et al., 2023; Windarto et al., 2024). In practice, these extracts are applied via short immersion or periodic dosing in tanks/raceways at empirically optimized concentrations and frequencies; outcomes are monitored via SGR, thallus/ramuli metrics, chlorophyll-a, typically proximate/nutrient profiles. For deployment at scale, including basic safeguards, standardizing extract preparation, verifying the absence of contaminants, tracking residues where relevant, and documenting batch conditions are important to support reproducibility, food safety, and exporter documentation.

Regional practices and production hotspots

C. lentillifera is prominently cultivated in regions such as Okinawa (Japan) and Mindanao (Philippines), where it holds cultural and economic value. Although cross-regional productivity statistics remain limited, its distinctive texture and niche demand position it differently from staple carrageenophytes such as *Kappaphycus* or *Gracilaria* (**Stuthmann** *et al.*, **2023**).

In the Philippines, *C. racemosa*, locally "ar-arusip", is commonly harvested alongside *C. lentillifera*. A multi-region socio-economic survey identified Mactan (Cebu) as a top producer, with peak dry-season harvests of ~42 tonnes per month and estimated annual incomes near PHP 2.26 million (≈USD 47,700) (Estrada *et al.*, 2021). These findings underscore the genus's economic role and cultural integration in Southeast Asia and the Pacific, while highlighting persistent constraints: reliance on wild harvests, limited standardized cultivation, short post-harvest shelf life, and uncertain site carrying capacity.

Priority needs include: (i) development and dissemination of sustainable farming protocols tailored to local conditions; (ii) post-harvest innovations to extend shelf life and stabilize quality; and (iii) community-based value-chain strengthening to improve market access and resilience (Estrada et al., 2021; Stuthmann et al., 2023).

Post-harvest and preservation

A primary constraint to commercialization of *C. lentillifera* is its short shelf life, typically ~3–5 days at ambient temperature, driven by high water content and rapid microbial spoilage (**Toyen** *et al.*, **2025**). Preservation efforts have focused on brining: *C. lentillifera* can retain acceptable nutritional quality for up to ~12 weeks, though with perceptible color and firmness changes; maintaining salt concentrations below ~30% (w/v) helps preserve sensory attributes (**Pan-utai** *et al.*, **2023**). Recent work also explores UV-C irradiation to suppress microbial growth and extend freshness during short-term

storage (**Toyen** *et al.*, **2025**). The flowchart of post-harvest and preservation of *Caulerpa* can be seen in Fig. (4).

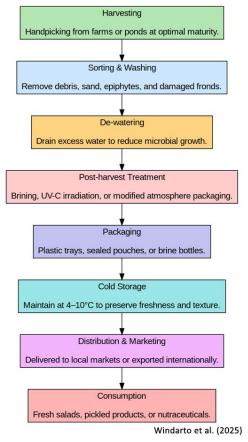


Fig. 4. Post-harvest and preservation of *Caulerpa*

C. racemosa presents similar challenges owing to delicate tissues and high moisture, leading to rapid quality loss post-harvest. Although species-specific studies remain limited, broader analyses of green seaweeds indicate that brining, controlled dehydration, and temperature management can stabilize texture and extend shelf life (**Zhu** et al., 2021). Metabolite profiling of C. racemosa under varied handling suggests several secondary metabolites remain stable under mild processing, supporting the feasibility of gentle preservation strategies (**Taslim** et al., 2024).

Nutritional, functional, and industrial applications Nutritional composition

C. lentillifera is a nutrient-rich green macroalga. Dry-weight analysis reveals it contains approximately 7% protein, 14% lipid, 17.5% dietary fiber, and 44% carbohydrates, indicating a robust macronutrient profile supportive of human nutrition (de Preez et al., 2020). C. racemosa, or in Indonesia locally known as "Latoh", contains 6% of ash, 0.5-0.7% of fat, 1-1.5% of fiber, and 6-8% of protein (Windarto et al., 2023;

Windarto et al., 2024). Furthermore, Caulerpa showed a high level of polyunsaturated fatty acids (PUFAs) (Brix da Costa et al., 2025).

Comparative studies across regions such as Hainan, Shandong, Sabah, and Thailand show high levels of polysaccharides (up to ~59%) and proteins (up to ~19%), along with notably high contents of calcium (3,300– 3,700mg/ 100g), magnesium (6,700– 8,100mg/ 100g), iron (500– 2,000mg/ 100g), selenium, and zinc (**Zhang** *et al.*, **2020a**). *C. racemosa* from Jepara, Indonesia, showed a high Na (2.740 mg/100g), Si (1.630mg/ 100g), Cl (4.5mg/ 100g), and Ca (1.960mg/ 100g) (**Windarto** *et al.*, **2025a**). These figures underscore *Caulerpa*'s potential as a vibrant source of essential minerals and macronutrients.

Bioactive compounds

Beyond its impressive nutritional profile, *C. lentillifera* also harbors significant bioactive compounds. Research highlights the presence of sulfated polysaccharides exhibiting immune-enhancing and anti-inflammatory properties (**Tesvichian** *et al.*, 2024). Experimental extracts of *C. lentillifera* have demonstrated antioxidant, anti-hyperglycemic, and anti-hypercholesterolemic effects *in vivo*, likely via modulation of oxidative stress and metabolic pathways (**Manoppo** *et al.*, 2022). Furthermore, treatment of inflammatory macrophages (RAW 264.7 cells) with *C. lentillifera* extract led to decreased production of pro-inflammatory cytokines such as IL-6, IL-1β, and TNF-α, indicating strong anti-inflammatory activity (**Yoojam** *et al.*, 2021). *C. racemosa* contains sulfated polysaccharides, alkaloids, flavonoids, and terpenoids with diverse bioactivities. Sulfated polysaccharides (SPCr) enhance AMPK and SIRT1 expression while reducing mTOR signaling, supporting anti-aging and metabolic regulation (**Nurkolis** *et al.*, 2023).

Food and nutraceutical applications

Across Southeast Asia, *C. lentillifera* ("sea grapes"/"green caviar") is commonly eaten fresh (salads) or lightly salted, valued for its crisp texture and briny flavor (**Stuthmann** *et al.*, **2023**). Beyond fresh use, compositional profiles and bioactives support development of functional foods and dietary supplements.

For *C. racemosa*, techno-functional measurements indicate water-holding capacity ≈ 2.16 g/g, swelling capacity ≈ 6.56 mL/g, and oil-holding capacity ≈ 1.15 g/g, suggesting applicability as a texture/moisture modifier and emulsion stabilizer (**Windarto** *et al.*, **2025a**). Fermented products (e.g., *C. racemosa* kombucha) have been explored for "antiageing" positioning, though substantiation remains preliminary (**Permatasari** *et al.*, **2021**).

Sulfated polysaccharides isolated from *C. racemosa* (SPCr) show pancreatic-lipase inhibitory and anti-inflammatory activities in experimental models, indicating anti-obesity potential (**Mayulu** *et al.*, **2023**; **Inoubli** *et al.*, **2025**). Metabolomic and bioassay studies report antioxidant and cytotoxic effects, including growth inhibition of breast-

cancer cell lines (**Taslim** *et al.*, **2024**). Broader screenings identify terpenoids, alkaloids, sterols, and other secondary metabolites with reported antihypertensive, antihyperlipidemic, antimicrobial, antidiabetic/antihyperglycemic, anti-inflammatory, and antioxidant activities, primarily demonstrated *in vitro* or in animal models (**Iveša** *et al.*, **2024**). Translation to human health outcomes requires controlled clinical studies and robust safety/toxicology assessments. The summary of bioactive compounds and nutraceutical functions of *Caulerpa* can be seen in Table (1).

Table 1. Caulerpa species and their nutraceutical functions

Table 1. Camerpa species and then nutraceutical functions				
Species	Bioactive Compounds	Applications	References	
C. lentilifera	Sulfated polysaccharides	Anti-inflammatory,	Sun et al., (2018);	
		immunomodulatory,	Tian et al.,	
		antioxidant, anti-	(2019);	
		obesity, anti-viral,	Khairuddin et	
		anti-diabetic	al., (2020); Sun et	
			al., (2020); Le et	
			al., (2022);	
			Manoppo et al.,	
			(2022); You et al.,	
			(2022);	
			Tesvichian et al.,	
			(2024);	
C. lentilifera	Polyphenols, carotenoids, flavonoids, peptides, PUFAs	Antioxidant, antihyperglycemic, antihypertensive, antiinflammatory, antidiabetic, antiobesity	Yoojam et al., (2021); Syakilla et al., (2022); Lin et al., (2022); Nurkolis et al., (2022); Sommer et al., (2022); Koodkaew et al., (2024); Montolalu et al., (2024); Kurniawan et al.,	
C. lentilifera	13-	Antioxidant, anti-	(2025) Manmuan et al.,	
C. ichingera	sophorosyloxydocosanoic	cancer	(2025)	
	acid, jubanine, cysteinyl-	Carico	(2023)	
	tyrosine, polyphenols			
C. lentilifera	dl-2-phenyltryptophane	reduce hepatic lipid	Sangpairoj et al.,	
	or 2 phonyrayprophane	reduce hepatic lipid	- Samspanioj et ali,	

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	and	accumulation	(2024)
	benzoic acid		
C. sertularioides	Caulerpin, difucol, quercetin, phenol, flavonoids, carotenoids, sulfated polysaccharides	Antioxidant, anti- cancer	Anjali et al., (2022); Agena et al., (2023)
Caulerpa spp	Caulerpin, monomethyl caulerpinate, and caulersin	Anti-cancer (in silico)	Olgun <i>et al.</i> , (2022)
C. racemosa	Caulerpin, bioactive peptides, squalene, carotenoids, phenolic content, fatty acids	Antifungal, anticancer, antioxidant, anti-inflammatory	Shanura Fernando et al., (2018); Permatasari et al., (2022); Palaniyappan et al., (2023); Nursidika et al., (2024); Permatasari et al., (2024); Taslim et al., (2024)
C. racemosa	Bioactive peptides, ADM, flavonoids, phenolic content	Antioxidant, anti- hypertensive, anti- viral (in silico), antimicrobial	Abdul-Razak et al., (2020); Pangestuti et al., (2021); Tassakka et al., (2023); Purwati et al., (2024); Windarto et al., (2025b)
C. taxifolia	Heptadecane, Hexadecanoic acid, ester derivatives, phenolic, Ag NPs, caulerpenyne	Antimicrobial, antioxidant, anti- cancer, antitumor	Akbar et al., (2023); Zhang et al., (2020b); Sfecci et al., (2017)
C. prolifera	Vanillin, p-coumaric acid, sinapic acid, 7,3',4'-flavon-3-ol, and kaempferol, di-(2-ethylhexyl) phthalate, β-sitosterol, erucic acid, nervonic acid	Antioxidant, antidiabetic, antiaging	Ouhabi et al., (2025); Rosa et al., (2025)

C. peltata	Heptadecane, Hexadecanoic	Antimicrobial,	Hao et al., (2019);
	acid, ester derivatives,	antioxidant, anti-	Akbar et al.,
	phenolic, alkaloid, terpenes,	inflammatory, and	(2023); Nayaka et
	caulerpin, manomethyl	immunostimulatory	al., (2020)
	caulerpinate	activity	

Cosmetics and industrial uses

Although specific cosmetic applications of *Caulerpa* remain exploratory, related green seaweeds are known for their antioxidant and hydrating properties, making them desirable in anti-aging and skin-care formulations (**Peñalver** *et al.*, **2020**). Additionally, the genus *Caulerpa* has shown promise in bioremediation and biofilter, while certain extracts are under investigation for uses in animal feed and bioplastic or bioenergy technologies (**Margono** *et al.*, **2021**; **Landi** *et al.*, **2022**; **Natsir** *et al.*, **2022**; **Borazjani** *et al.*, **2025**). Moreover, *C. racemosa* has also been found to be a promising antimicrobial edible film for active packaging of food (**Rusli** *et al.*, **2024**).

Market trends

Although precise market data for *Caulerpa* remain limited, the Asia-Pacific region, particularly the Philippines, Japan, and parts of Southeast Asia, dominates its production and consumption, with established value chains for fresh and lightly processed forms (**Zhang** *et al.*, **2020**; **Stuthmann** *et al.*, **2023**). The growing popularity of *C. lentillifera* globally signals increasing potential for export, functional food, and bioproduct innovation.

Sustainability, environmental impact, and challenges Sustainability in aquaculture

C. lentillifera has significant potential in climate-smart aquaculture, notably through its integration into Integrated Multi-Trophic Aquaculture (IMTA) systems. While studies specific to Caulerpa are limited, broader IMTA approaches, where seaweeds act as nutrient sinks, are recognized for their environmental benefits. These approaches enable nutrient recycling, eutrophication control, and diverse co-culture productivity (Loayza-Aguilar et al., 2023; Alam et al., 2024).

In an experimental setting, *C. lentillifera* demonstrated high nutrient uptake capacities (nitrogen and phosphorus) when cultured with fish effluent, suggesting its utility as a biofilter in sustainable aquaculture (**Bambaranda** *et al.*, **2019a**, **b**).

Ecosystem services

Caulerpa enhances carbon sequestration by fixing CO₂ during photosynthesis, contributing to "blue carbon" storage and helping offset greenhouse gas emissions (Farghali et al., 2023; Pessarrodona et al., 2024). Moreover, C. lentillifera has proven

effective in water purification, notably in the removal of nitrogenous waste (e.g., nitrate, phosphate), crucial for ameliorating aquaculture effluent impacts (**Bambaranda** *et al.*, **2019a**, **b**).

Invasiveness and trade-offs

Some *Caulerpa* species pose ecological risks in non-native habitats. For example, *Caulerpa cylindracea* (formerly *C. racemosa* var. cylindracea) has aggressively invaded the Mediterranean, outcompeting native benthic species and altering local ecosystems (**Brewton & Lapointe, 2023**). Balancing *Caulerpa*'s utilization and ecological caution requires stringent cultivation protocols and targeted deployment to minimize accidental spread.

Cultivation and market challenges

Commercial cultivation of *C. lentillifera* faces significant hurdles, particularly post-harvest perishability and shelf-life constraints. Fresh "sea grapes" are highly perishable due to their succulent, high-moisture tissues, which rapidly degrade when exposed to ambient conditions or oxidative stress. In a seasonal quality study of Taiwanese seagrapes, storage without seawater led to substantial 20–40% water loss, 70–90% lipid peroxidation (MDA production), and drastic reductions in total phenolics and chlorophyll within just a few days, underscoring the difficulty in preserving freshness under typical storage conditions. However, treatment with an alternating current electric field (ACEF) extended shelf-life by up to 12 days by mitigating physicochemical deterioration (e.g., maintaining chlorophyll and phenolic content) in spring-harvested samples (**Sulaimana** *et al.*, **2021**).

Another challenge lies in market development, particularly in Western regions. Consumer acceptance of seaweed products depends heavily on awareness and nutritional positioning. A 2024 consumer psychology study found that sharing information about nutritional benefits significantly improves Western consumers' acceptance and sensory perception of seaweed-based foods (**Moss** *et al.*, **2024**). The roadmap for *Caulerpa* development is illustrated in Fig. (5).

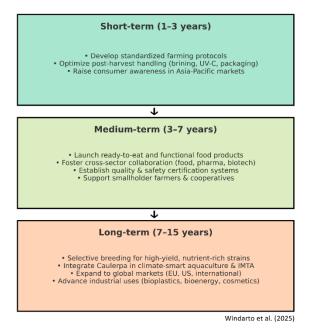


Fig. 5. Roadmap for Caulerpa development

Future outlook and recommendations

Caulerpa's market growth will depend on (i) product innovation, especially ready-toeat formats and standardized functional extracts (e.g., sulfated polysaccharides; alkaloids such as caulerpin), and (ii) cross-sector collaboration among aquaculture, food, pharma, and biotech to translate experimental bioactivities into regulated products with validated safety and quality profiles. Structured reviews for *C. lentillifera* already document workable post-harvest solutions (e.g., brining ranges) and premium fresh positioning that can extend to shelf-stable functional foods and nutraceuticals (**Stuthmann** *et al.*, **2023**).

At the policy/regulatory level, harmonized cultivation, food-safety, and export standards (contaminants, allergens, labeling, traceability) are needed for seaweeds; proactive risk assessment for offshore/novel farming should be embedded into permitting and guidance, elements that *Caulerpa* value chains can adopt to access higher-value markets while protecting consumers (**Banach** *et al.*, **2020**).

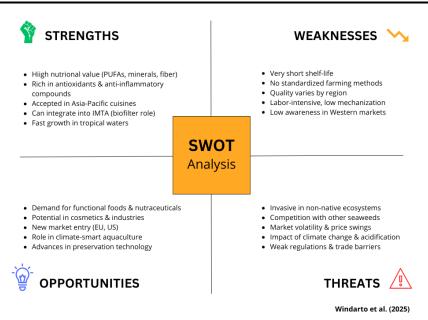


Fig. 6. SWOT analysis of Caulerpa

To translate *Caulerpa*'s promise into reliable products and supply, the next phase needs targeted R&D that closes known technical gaps while generating decision-grade safety and quality data. The following priorities are the most actionable near-term levers:

- 1. Breeding & domestication: Select high-yield, nutrient-dense, and morphologyoptimized strains suited to mechanization, following macroalgal selective-breeding frameworks (Charrier *et al.*, 2015).
- 2. Post-harvest technologies: Validate brine concentration—time—temperature windows; assess emerging physical treatments (e.g., UV-C) for quality and safety; develop packaging that limits oxidation and texture loss (Pan-utai et al., 2023; Stuthmann et al., 2024).
- 3. Safety/toxicology: Establish dose–response and exposure data for key metabolites (e.g., caulerpenyne, caulerpin) across intended uses (food, cosmetic, pharma); emerging in vivo data suggest low acute toxicity for caulerpin at dietary-relevant levels, supporting further development under GMP (Russo et al., 2024; dos Santos, 2025).

Commercial traction will come from matching product formats to the most ready markets while building credibility through standards and documentation. The steps below stage investments from existing hubs toward new regions and channels:

1. Near term (Asia–Pacific): Consolidate existing hubs with best-practice cultivation and post-harvest SOPs; deploy cooperative/certification schemes to stabilize quality and pricing.

- 2. New regions: Build evidence-based narratives with standardized quality metrics (moisture, ash, contaminants, bioactives), validated processes, and importer-aligned documentation.
- 3. Platforms: Position *Caulerpa* as (a) premium fresh produce, (b) functional ingredients (fiber/texture modifiers, extracts), and (c) co-products in IMTA/blue-economy initiatives (**Sultana** *et al.*, **2023**; **Khan** *et al.*, **2024**).

CONCLUSION

Caulerpa, particularly C. lentillifera and C. racemosa, is an underutilized but promising genus for diversified seaweed value chains. Its nutrient density, bioactive-rich profile, and suitability for fresh, functional-food, nutraceutical, and cosmetic applications position it as a strategic resource for climate-smart aquaculture and blue-economy development. Current barriers, short shelf life, limited and non-standardized farming, low consumer awareness outside Asia, and ecological/biosecurity risks, constrain scale.

Realizing this potential requires coordinated action: (i) validated post-harvest technologies (e.g., optimized brining, emerging physical treatments, fit-for-purpose packaging) to extend shelf life and stabilize quality; (ii) selective breeding and domestication targeting yield, texture, and nutrient traits; (iii) harmonized standards for cultivation, safety, and trade (contaminants, labeling, traceability); and (iv) cross-sector partnerships to translate experimental bioactivities into regulated, market-ready ingredients. With these steps, *Caulerpa* can credibly progress from regional "sea grapes" to globally recognized "green gold of the ocean," contributing meaningfully to future seaweed markets.

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REFERENCES

Abdul Razak, S.; Palaniveloo, K.; Nagappan, T.; Wee Jin, G.; Chellappan, D. K.; Chellian, J. and Kunnath, A. P. (2020). Bioprospecting cultivated tropical green algae, *Caulerpa racemosa* (Forsskal) J. Agardh: A perspective on nutritional properties, antioxidative capacity and anti-diabetic potential. *Foods*, *9*(9), 1313. https://doi.org/10.3390/foods9091313

Agena, R.; Jesús, A. D.; Marat, L.; Pablo, O.; García, V. H. and Jaramillo Flores, M. E. (2022). Pro-apoptotic activity and cell cycle arrest of *Caulerpa sertularioides*

- against SKLU-1 cancer cell in 2D and 3D cultures. *Molecules*, *28*(11), 4361. https://doi.org/10.3390/molecules28114361
- **Akbar, S.A.; Hasan, M.; Afriani, S. and Nuzlia, C.** (2023). Evaluation of phytochemical composition and metabolite profiling of macroalgae *Caulerpa taxifolia* and *C. peltata* from the Banda Aceh coast, Indonesia. *Biodiversitas*, *24*, 5283–5292. https://doi.org/10.13057/biodiv/d241009
- Alam, M.M.; Jørgensen, N.O.G.; Bass, D.; Santi, M.; Nielsen, M.; Rahman, M.A.; Hasan, N.A.; Bablee, A.L.; Bashar, A.; Hossain, M.I.; Hansen, L.H. and Haque, M.M. (2024). Potential of integrated multitrophic aquaculture to make prawn farming sustainable in Bangladesh. *Frontiers in Sustainable Food Systems*, *8*, 1412919. https://doi.org/10.3389/fsufs.2024.1412919
- Anjali, R.; Palanisamy, S.; Vinosha, M.; Selvi, A. M.; Manikandakrishnan, M.; Sathiyaraj, G.; Marudhupandi, T.; You, S. and Prabhu, N. M. (2022). Sulfated polysaccharides from *Caulerpa sertularioides*: Extraction and evaluation of antioxidant, antibacterial, and immunological properties. *Industrial Crops and Products*, *188*, 115671. https://doi.org/10.1016/j.indcrop.2022.115671
- **Arias-Echeverri, J. P.; Zapata-Ramirez, P. A.; Ramirez-Carmona, M.; Rendon-Castrillon, L. and Ocampo-Lopez, C.** (2022). Present and future of seaweed cultivation and its applications in Colombia. *Journal of Marine Science and Engineering*, *10*(2), 243. https://doi.org/10.3390/jmse10020243
- **Arimoto, A.; Nishitsuji, K.; Narisoko, H.; Shoguchi, E. and Satoh, N.** (2019). Differential gene expression in fronds and stolons of the siphonous macroalga, *Caulerpa lentillifera. Development, Growth & Differentiation*, *61*(9), 475–484. https://doi.org/10.1111/dgd.12634
- Bambaranda, B. V.; Sasaki, N.; Chirapart, A.; Salin, K. R. and Tsusaka, T. W. (2019a). Optimization of macroalgal density and salinity for nutrient removal by *Caulerpa lentillifera* from aquaculture effluent. *Processes*, *7*(5), 303. https://doi.org/10.3390/pr7050303
- **Bambaranda, B. V.; Tsusaka, T. W.; Chirapart, A.; Salin, K. R. and Sasaki, N.** (2019b). Capacity of *Caulerpa lentillifera* in the removal of fish culture effluent in a recirculating aquaculture system. *Processes*, *7*(7), 440. https://doi.org/10.3390/pr7070440
- Banach, J.; Van den Burg, S. and Van der Fels-Klerx, H. (2020). Food safety during seaweed cultivation at offshore wind farms: An exploratory study in the North Sea. *Marine Policy*, *120*, 104082. https://doi.org/10.1016/j.marpol.2020.104082
- **Belkacemi, L.; Mahmoud, B.; Ali, C. D. and Youcef, B.** (2020). Antioxidant and antibacterial activities and identification of bioactive compounds of various extracts of *Caulerpa racemosa* from Algerian coast. *Asian Pacific Journal of Tropical Biomedicine*, *10*(2), 87–94. https://doi.org/10.4103/2221-1691.275423

- Borazjani, Z.; Azin, R.; Osfouri, S.; Karami, R.; Kennedy, E. and Stockenhuber, M. (2025). Hydrothermal liquefaction of *Caulerpa sertularioides*: Optimized biocrude production and characterization with pretreatment techniques. *Biomass and Bioenergy*, *194*, 107635. https://doi.org/10.1016/j.biombioe.2025.107635
- **Brewton, R. A. and Lapointe, B. E.** (2023). The green macroalga *Caulerpa prolifera* replaces seagrass in a nitrogen-enriched, phosphorus-limited, urbanized estuary. *Ecological Indicators*, *156*, 111035. https://doi.org/10.1016/j.ecolind.2023.111035
- **Brix da Costa, B.; Kunzmann, A. and Springer, K.** (2025). Comparative analysis of the nutritional profiles of five edible macroalgae as sustainable food sources. *Discover Food*, *5*, 287. https://doi.org/10.1007/s44187-025-00603-3
- **Charrier, B.; Rolland, E.; Gupta, V. and Reddy, C. R.** (2015). Production of genetically and developmentally modified seaweeds: Exploiting the potential of artificial selection techniques. *Frontiers in Plant Science*, *6*, 127. https://doi.org/10.3389/fpls.2015.00127
- Dos Santos, J. S. P.; Silva, D. K. C.; da Silva Oliveira, V.; Junior, S. S. S.; Dos Santos Rodrigues, E.; de Souza, C. V. C.; Martinez, S. T.; Santos-Filho, O. A.; Meira, C. S. and Soares, M. B. P. (2025). The alkaloid caulerpin exhibits potent and selective anti-inflammatory activity through interaction with the glucocorticoid receptor. *Marine Drugs*, *23*(6), 232. https://doi.org/10.3390/md23060232
- du Preez, R.; Majzoub, M. E.; Thomas, T.; Panchal, S. K. and Brown, L. (2020). *Caulerpa lentillifera* (sea grapes) improves cardiovascular and metabolic health of rats with diet-induced metabolic syndrome. *Metabolites*, *10*(12), 500. https://doi.org/10.3390/metabo10120500
- **Estrada, J. L.; Arboleda, M. D. M. and Dionisio-Sese, M. L.** (2021). Current status of sea grapes (*Caulerpa* spp.) farming and wild harvesting in the Philippines. *Journal of Applied Phycology*, *33*, 3215–3223. https://doi.org/10.1007/s10811-021-02533-w
- **Farghali, M.; Mohamed, I. M. A.; Osman, A. I. and Rooney, D. W.** (2023). Seaweed for climate mitigation, wastewater treatment, bioenergy, bioplastic, biochar, food, pharmaceuticals, and cosmetics: a review. *Environmental Chemistry Letters*, *21*, 97–152. https://doi.org/10.1007/s10311-022-01520-y
- **Fraissinet, S.; Blanco, J.; Aguilar, R. and Rossi, S.** (2025). First record of *Caulerpa prolifera* (Bryopsidales, Chlorophyta) beyond 60 m depth in the SW Tenerife Island, Spain. *Regional Studies in Marine Science*, *90*, 104466. https://doi.org/10.1016/j.rsma.2025.104466
- Ghani, P. K.; Harwanto, D.; Amalia, R.; Windarto, S.; Haditomo, A. H.; Nurhayati, D. and Susilowati, T. (2023). The Effect of Combination of Coconut Water (*Cocos nucifera*) and Shallot Crude Extract (*Allium cepa* L.) on the Growth of *Caulerpa*

- racemosa. IOP Conference Series: Earth and Environmental Science, *1224*, 012001. https://doi.org/10.1088/1755-1315/1224/1/012001
- Hao, H.; Fu, M.; Yan, R.; He, B.; Li, M.; Liu, C.; Cai, Y.; Zhang, X. and Huang, R. (2019). Chemical composition and immunostimulatory properties of green alga *Caulerpa racemosa* var. *peltata*. *Food and Agricultural Immunology*, *30*(1), 937–954. https://doi.org/10.1080/09540105.2019.1646216
- Herawati, V.; Pinandoyo; Ariyati, R. W.; Rismaningsih, N.; Windarto, S.; Prayitno, S. B.; Darmanto, Y. S. and Radjasa, O. K. (2021). Effects of *Caulerpa lentillifera* added into culture media on the growth and nutritional values of *Phronima pacifica*, a natural fish-feed crustacean. *Biodiversitas*, *22*(1), 267–276. https://doi.org/10.13057/biodiv/d220152
- Herawati, V. E.; Windarto, S.; Prasetyo, D. Y. B.; Prayitno, S. B. and Karna Radjasa, O. (2021). The effect of mass cultured *Phronima pacifica* feed using *Caulerpa lentillifera* substrate on growth performance and nutritional quality of pacific white shrimp (*Litopenaeus vannamei*). *Journal of Applied Aquaculture*, *35*(3), 703–721. https://doi.org/10.1080/10454438.2021.2018378
- **Hurtado, A. Q.; Neish, I. C. and Critchley, A. T.** (2019). Phyconomy: The extensive cultivation of seaweeds, their sustainability and economic value, with particular reference to important lessons to be learned and transferred from the practice of eucheumatoid farming. *Phycologia*, *58*(5), 472–483. https://doi.org/10.1080/00318884.2019.1625632
- **Inoubli, S.; López-Álvarez, M.; Shili, A.; González, P.; Torres, M.; Ksouri, R. and Domínguez, H.** (2025). Sulphated polysaccharide extracted from the Mediterranean edible seaweed *Caulerpa racemosa*: Structural, rheological and biological features. *Sustainable Chemistry and Pharmacy*, *45*, 102034. https://doi.org/10.1016/j.scp.2025.102034
- Islam, M. S.; Senaha, I.; Nagamatsu, K.; Yoda, Y. and Saha, B. B. (2023). Green algae *Caulerpa lentillifera* cultivation technique utilising CO₂ and its social impacts. *International Journal of Innovation and Sustainable Development*, *18*(1–2), 146–167. https://doi.org/10.1504/IJISD.2024.135261
- Iveša, N.; Burić, P.; Buršić, M.; Kovačić, I.; Pustijanac, E.; Šegulja, S.; Modrušan, A.; Bilić, J. and Millotti, G. (2024). A review on nutrients, phytochemicals, health benefits and applications of the green seaweed *Caulerpa racemosa* (Forsskål) J. Agardh. *Journal of Applied Phycology*, *36*, 3451–3473. https://doi.org/10.1007/s10811-024-03341-8
- **Janke, L.** (2024). Mapping the global mass flow of seaweed: Cultivation to industry application. *Journal of Industrial Ecology*, *28*(5), 1256–1269. https://doi.org/10.1111/jiec.13539
- Khairuddin, K.; Sudirman, S.; Huang, L. and Kong, Z. (2019). Caulerpa lentillifera polysaccharides-rich extract reduces oxidative stress and proinflammatory cytokine

- levels associated with male reproductive functions in diabetic mice. *Applied Sciences*, *10*(24), 8768. https://doi.org/10.3390/app10248768
- **Khan, N.; Sudhakar, K. and Mamat, R.** (2024). Macroalgae farming for sustainable future: Navigating opportunities and driving innovation. *Heliyon*, *10*(7), e28208. https://doi.org/10.1016/j.heliyon.2024.e28208
- **Koodkaew, I.; Pitakwongsaporn, S.; Jarussophon, N. and Wichachucherd, B.** (2024). Chemical profile, antioxidant activity and α-glucosidase inhibition of sea grape *Caulerpa lentillifera* collected from different sites in Thailand. *Trends in Sciences*, *21*(4), 7520. https://doi.org/10.48048/tis.2024.7520
- Kurniawan, R.; Nurkolis, F.; Bukhari, A.; Syauki, A. Y.; Bahar, B.; Aman, A. M. and Taslim, N. A. (2025). Carotenoid and peptide supplementation from *Caulerpa* sp. (sea grapes) extract mitigate metabolic syndrome in cholesterol-enriched diet rats via modulation of gut microbiota. *Diabetology & Metabolic Syndrome*, *17*, 333. https://doi.org/10.1186/s13098-025-01869-4
- **Lagourgue, L.; Sauvage, T.; Zubia, M.; Draisma, S. G.; Vieira, C.; Engelen, A. and Payri, C. E.** (2024). Taxonomic insights into *Caulerpa* (Bryopsidales, Chlorophyta) species in French Polynesia: Confirmation of 13 species and reinstatement of *C. pickeringii* Harvey & Bailey. *Diversity*, *16*(4), 243. https://doi.org/10.3390/d16040243
- Landi, S.; Santini, G.; Vitale, E.; Natale, G. D.; Maisto, G.; Arena, C. and Esposito, S. (2022). Photosynthetic, molecular and ultrastructural characterization of toxic effects of zinc in *Caulerpa racemosa* indicate promising bioremediation potentiality. *Plants*, *11*(21), 2868. https://doi.org/10.3390/plants11212868
- **Largo, D. B.; Diola, A. G. and Marababol, M. S.** (2016). Developing an integrated multi-trophic aquaculture (IMTA) system for tropical marine species in southern Cebu, Central Philippines. *Aquaculture Reports*, *3*, 67–76. https://doi.org/10.1016/j.agrep.2015.12.006
- Lau, V.; Nurkolis, F.; Park, M. N.; Heriyanto, D. S.; Taslim, N. A.; Tallei, T. E.; Permatasari, H. K.; Tjandrawinata, R. R.; Moon, S. and Kim, B. (2024). Green seaweed *Caulerpa racemosa* as a novel non-small cell lung cancer inhibitor in overcoming tyrosine kinase inhibitor resistance: An analysis employing network pharmacology, molecular docking, and in vitro research. *Marine Drugs*, *22*(6), 272. https://doi.org/10.3390/md22060272
- **Le, B.; Do, D. T.; Nguyen, H. M.; Do, B. H. and Le, H. T.** (2022). Preparation, characterization, and anti-adhesive activity of sulfate polysaccharide from *Caulerpa lentillifera* against *Helicobacter pylori*. *Polymers*, *14*(22), 4993. https://doi.org/10.3390/polym14224993
- Lin, K.; Yang, H.; Yang, S.; Chen, Y.; Watanabe, Y. and Chen, J. (2022). *Caulerpa lentillifera* improves ethanol-induced liver injury and modulates the gut microbiota

- in rats. *Current Research in Food Science*, *7*, 100546. https://doi.org/10.1016/j.crfs.2023.100546
- **Loayza-Aguilar, R. E.; Huamancondor-Paz, Y. P.; Saldaña-Rojas, G. B. and Olivos-Ramirez, G. E.** (2023). Integrated multi-trophic aquaculture (IMTA): Strategic model for sustainable mariculture in Samanco Bay, Peru. *Frontiers in Marine Science*, *10*, 1151810. https://doi.org/10.3389/fmars.2023.1151810
- Manmuan, S.; Sirirak, T.; Tubtimsri, S.; Petchsomrit, A. and Chuenbarn, T. (2025). Phytochemical analysis, antioxidant activity, and cytotoxic effects of *Caulerpa lentillifera* extracts inducing cell apoptosis and sub-G/G0-G1 cell cycle arrest in KON oral cancer cells. *BMC Complementary Medicine and Therapies*, *25*, 101. https://doi.org/10.1186/s12906-025-04835-9
- Mannino, A. M.; Cicero, F.; Toccaceli, M.; Pinna, M. and Balistreri, P. (2019). Distribution of *Caulerpa taxifolia* var. *distichophylla* (Sonder) Verlaque, Huisman & Procaccini in the Mediterranean Sea. *Nature Conservation*, *37*, 17–29. https://doi.org/10.3897/natureconservation.37.33079
- Manoppo, J. I.; Nurkolis, F.; Pramono, A.; Ardiaria, M.; Murbawani, E. A.; Yusuf, M.; Qhabibi, F. R.; Yusuf, V. M.; Amar, N.; Karim, M. R.; Subali, A. D.; Natanael, H.; Rompies, R.; Halim, R. F.; Bolang, A. S.; Joey, G.; Novianto, C. A. and Permatasari, H. K. (2022). Amelioration of obesity-related metabolic disorders via supplementation of *Caulerpa lentillifera* in rats fed with a high-fat and high-cholesterol diet. *Frontiers in Nutrition*, *9*, 1010867. https://doi.org/10.3389/fnut.2022.1010867
- Mantri, V. A.; Munisamy, S. and Kambey, C. S. (2024). Biosecurity aspects in commercial *Kappaphycus alvarezii* farming industry: An India case study. *Aquaculture Reports*, *35*, 101930. https://doi.org/10.1016/j.aqrep.2024.101930
- Margono; Anggadiredja, J. T. and Nurhudah, M. (2021). Effectiveness of seaweed (*Caulerpa lentillifera*) as biofilter in vanamei shrimp (*Litopenaeus vannamei*) culture. *AACL Bioflux*, *14*(3), 1734–1746.
- Mayulu, N.; Gunawan, W. B.; Park, M. N.; Chung, S.; Suh, J. Y.; Song, H.; Kusuma, R. J.; Taslim, N. A.; Kurniawan, R.; Kartawidjajaputra, F.; Nurkolis, F. and Kim, B. (2023). Sulfated polysaccharide from *Caulerpa racemosa* attenuates the obesity-induced cardiometabolic syndrome via regulating the PRMT1-DDAH-ADMA with mTOR-SIRT1-AMPK pathways and gut microbiota modulation. *Antioxidants*, *12*(8), 1555. https://doi.org/10.3390/antiox12081555
- **Mengist, W.; Soromessa, T. and Legese, G.** (2019). Method for conducting systematic literature review and meta-analysis for environmental science research. *MethodsX*, *7*, 100777. https://doi.org/10.1016/j.mex.2019.100777
- Montefalcone, M.; Morri, C.; Parravicini, V. and Bianchi, C. N. (2015). A tale of two invaders: Divergent spreading kinetics of the alien green algae *Caulerpa taxifolia*

- and *Caulerpa cylindracea*. *Biological Invasions*, *17*, 2717–2728. https://doi.org/10.1007/s10530-015-0908-1
- Montolalu, R. I.; Dotulong, V.; Mentang, F.; Taher, N. and Makapedua, D. M. (2024). A comparative analysis on impact of drying methods on antioxidants, antidiabetes and antiobesity activities in green algae *Caulerpa lentillifera*: In vitro study. *Algal Research*, *84*, 103768. https://doi.org/10.1016/j.algal.2024.103768
- Moreira, A.; Cruz, S.; Marques, R. and Cartaxana, P. (2021). The underexplored potential of green macroalgae in aquaculture. *Reviews in Aquaculture*, *14*(1), 5–26. https://doi.org/10.1111/raq.12580
- Moss, R.; Stright, A.; Nicolle, L.; Richelle, E.; Baxter, L.; Frampton, K.; Dabas, T.; Gorman, M. and McSweeney, M. B. (2024). Impact of information about nutritional benefits, sustainability and consumption on consumer acceptance and emotional response to smoothies containing brown seaweed, *Ascophyllum nodosum*. Food and Humanity, *3*, 100373. https://doi.org/10.1016/j.foohum.2024.100373
- Natsir, S.; Tahya, A. M.; Nilawati, J. and Ismail, S. N. (2022). Utilization of *Caulerpa* sp. as a feed ingredient for growth and survival of whiteleg shrimp and *Chanos chanos* in polyculture. *Egyptian Journal of Aquatic Research*, *48*(2), 175–180. https://doi.org/10.1016/j.ejar.2022.01.005
- Nayaka, S. R.; Anand, J. V. S.; Shareef, S. M. and Usha, N. S. (2020). Gas chromatography—mass spectroscopy (GC–MS) analysis and phytochemical screening for bioactive compounds in *Caulerpa peltata* (green alga). *Biomedical and Pharmacology Journal*, *13*(4), 1627–1635. https://dx.doi.org/10.13005/bpj/2069
- Nurkolis, F.; Taslim, N. A.; Subali, D.; Kurniawan, R.; Hardinsyah, H.; Gunawan, W. B.; Kusuma, R. J.; Yusuf, V. M.; Pramono, A.; Kang, S.; Mayulu, N.; Syauki, A. Y.; Tallei, T. E.; Tsopmo, A. and Kim, B. (2022). Dietary supplementation of *Caulerpa racemosa* ameliorates cardiometabolic syndrome via regulation of PRMT-1/DDAH/ADMA pathway and gut microbiome in mice. *Nutrients*, *15*(4), 909. https://doi.org/10.3390/nu15040909
- Nurkolis, F.; Taslim, N. A.; Qhabibi, F. R.; Kang, S.; Moon, M.; Choi, J.; Choi, M.; Park, M. N.; Mayulu, N. and Kim, B. (2023). Ulvophyte green algae *Caulerpa lentillifera*: Metabolites profile and antioxidant, anticancer, anti-obesity, and in vitro cytotoxicity properties. *Molecules*, *28*(3), 1365. https://doi.org/10.3390/molecules28031365
- **Nursidika, P.; Julianti, E. and Kurniati, N. F.** (2024). Fungistatic activity and mechanism of *Caulerpa racemosa*, *Caulerpa lentillifera* fractions and caulerpin metabolite against pathogenic fungi. *Narra J*, *5*(1), e1714. https://doi.org/10.52225/narra.v5i1.1714

- **Olgun, N.; Basbinar, Y.; Cavas, L. and Ellidokuz, H.** (2022). In-silico molecular interactions among the secondary metabolites of *Caulerpa* spp. and colorectal cancer targets. *Frontiers in Chemistry*, *10*, 1046313. https://doi.org/10.3389/fchem.2022.1046313
- Ouahabi, S.; Daoudi, N. E.; Chebaibi, M.; Mssillou, I.; Rahhou, I.; Bnouham, M.; Hammouti, B.; Fauconnier, L.; Gotor, A. A.; Rhazi, L. and Ramdani, M. (2025). A comparative study of the phytochemical composition, antioxidant properties, and in vitro anti-diabetic efficacy of different extracts of *Caulerpa prolifera*. *Marine Drugs*, *23*(7), 259. https://doi.org/10.3390/md23070259
- Palaniyappan, S.; Sridhar, A.; Kari, Z. A.; Téllez-Isaías, G. and Ramasamy, T. (2023). Evaluation of phytochemical screening, pigment content, in vitro antioxidant, antibacterial potential and GC-MS metabolite profiling of green seaweed *Caulerpa racemosa*. *Marine Drugs*, *21*(5), 278. https://doi.org/10.3390/md21050278
- **Pangestuti, R.; Haq, M.; Rahmadi, P. and Chun, B.** (2021). Nutritional value and biofunctionalities of two edible green seaweeds (*Ulva lactuca* and *Caulerpa racemosa*) from Indonesia by subcritical water hydrolysis. *Marine Drugs*, *19*(10), 578. https://doi.org/10.3390/md19100578
- **Pan-utai, W.; Satmalee, P.; Saah, S.; Paopun, Y. and Tamtin, M.** (2023). Brine-processed *Caulerpa lentillifera* macroalgal stability: Physicochemical, nutritional and microbiological properties. *Life*, *13*(11), 2112. https://doi.org/10.3390/life13112112
- Parreira, F.; Martínez-Crego, B.; Lourenço Afonso, C. M.; Machado, M.; Oliveira, F.; Manuel dos Santos Gonçalves, J. and Santos, R. (2021). Biodiversity consequences of *Caulerpa prolifera* takeover of a coastal lagoon. *Estuarine, Coastal and Shelf Science*, *255*, 107344. https://doi.org/10.1016/j.ecss.2021.107344
- Peñalver, R.; Lorenzo, J. M.; Ros, G.; Amarowicz, R.; Pateiro, M. and Nieto, G. (2020). Seaweeds as a functional ingredient for a healthy diet. *Marine Drugs*, *18*(6), 301. https://doi.org/10.3390/md18060301
- Permatasari, H. K.; Nurkolis, F.; Augusta, P. S.; Mayulu, N.; Kuswari, M.; Taslim, N. A.; Wewengkang, D. S.; Batubara, S. C. and Ben Gunawan, W. (2021). Kombucha tea from seagrapes (*Caulerpa racemosa*) potential as a functional antiageing food: In vitro and in vivo study. *Heliyon*, *7*(9), e07944. https://doi.org/10.1016/j.heliyon.2021.e07944
- Permatasari, H. K.; Wewengkang, D. S.; Tertiana, N. I.; Muslim, F. Z.; Yusuf, M.; Baliulina, S. O.; Daud, V. P. A.; Setiawan, A. A. and Nurkolis, F. (2022). Anticancer properties of *Caulerpa racemosa* by altering expression of Bcl-2, BAX, cleaved caspase-3 and apoptosis in HeLa cancer cell culture. *Frontiers in Oncology*, *12*, 964816. https://doi.org/10.3389/fonc.2022.964816

- Permatasari, H. K.; Amar, N.; Qhabibi, F. R.; Tertiana, N. I.; Subali, A. D.; Yusuf, V. M.; Hakim, D. I.; Nugroho, S. A.; Riawan, W. and Prijadi, B. (2024). Anticancer potential of sea grape (*Caulerpa racemosa*) extract by altering epithelial—mesenchymal transition and pro-apoptosis proteins expression in MCF-7 breast cancer cells. *Journal of Applied Pharmaceutical Science*, *14*(05), 176–184. https://doi.org/10.7324/JAPS.2024.169691
- **Pessarrodona, A.; Howard, J.; Pidgeon, E.; Wernberg, T. and Filbee-Dexter, K.** (2024). Carbon removal and climate change mitigation by seaweed farming: A state of knowledge review. *Science of The Total Environment*, *918*, 170525. https://doi.org/10.1016/j.scitotenv.2024.170525
- Purwati, E.; Amalia, R.; Windarto, S.; Rejeki, S. and Saputra, D. (2024). Antibacterial activity of endophytic fungi *Caulerpa racemosa* against *Vibrio parahaemolyticus* in Vaname Shrimp (*Litopenaeus vannamei*). *Asia Pacific Journal of Molecular Biology and Biotechnology*, *32*(4), 97-107. https://doi.org/10.35118/apjmbb.2024.032.4.11
- Ranjan, A.; Townsley, B. T.; Ichihashi, Y.; Sinha, N. R. and Chitwood, D. H. (2015). An intracellular transcriptomic atlas of the giant coenocyte *Caulerpa taxifolia*. *PLOS Genetics*, *11*(1), e1004900. https://doi.org/10.1371/journal.pgen.1004900
- Rosa, G. P.; Barreto, M. C.; Seca, A. M. L. and Pinto, D. C. G. A. (2025). Antiaging potential of lipophilic extracts of *Caulerpa prolifera*. *Marine Drugs*, *23*(2), 83. https://doi.org/10.3390/md23020083
- Russo, T.; Coppola, F.; Paris, D.; De Marchi, L.; Meucci, V.; Motta, A.; Carbone, M.; Di Cosmo, A.; Soares, A. M.; Pretti, C.; Mollo, E.; Freitas, R. and Polese, G. (2024). Exploring toxicological interactions in a changing sea: The case of the alkaloids caffeine and caulerpin. *Science of The Total Environment*, *912*, 169190. https://doi.org/10.1016/j.scitotenv.2023.169190
- Rusli, A.; Santi, A.; Malle, S.; Arfini, F.; Arsyad, M. A.; Syukroni, I. and Inthe, M. G. (2024). Application of antimicrobial edible film incorporated with *Caulerpa racemosa* crude extract as active packaging of seaweed dodol. *Applied Food Research*, *4*(2), 100625. https://doi.org/10.1016/j.afres.2024.100625
- Sangpairoj, K.; Pranweerapaiboon, K.; Saengkhae, C.; Meemon, K.; Niamnont, N.; Tamtin, M.; Sobhon, P.; Yisarakun, W. and Siangcham, T. (2024). Extracts of tropical green seaweed *Caulerpa lentillifera* reduce hepatic lipid accumulation by modulating lipid metabolism molecules in HepG2 cells. *Heliyon*, *10*(6), e27635. https://doi.org/10.1016/j.heliyon.2024.e27635
- Schembri, P. J.; Barbara, J.; Deidun, A.; Lanfranco, E. and Lanfranco, S. (2015). It was only a matter of time: Occurrence of *Caulerpa taxifolia* (Vahl) C. Agardh var. *distichophylla* (Sonder) Verlaque, Huisman and Procaccini in the Maltese Islands (Chlorophyta, Ulvophyceae, Caulerpaceae). *BioInvasions Records*, *4*(1), 9–16. http://dx.doi.org/10.3391/bir.2015.4.1.02

- Shanura Fernando, I. P.; Asanka Sanjeewa, K. K.; Samarakoon, K. W.; Lee, W. W.; Kim, S. and Jeon, J. (2018). Squalene isolated from marine macroalgae *Caulerpa racemosa* and its potent antioxidant and anti-inflammatory activities. *Journal of Food Biochemistry*, *42*(5), e12628. https://doi.org/10.1111/jfbc.12628
- Sfecci, E.; Le Quemener, C.; Lacour, T.; Massi, L.; Amade, P.; Audo, G. and Mehiri, M. (2017). Caulerpenyne from *Caulerpa taxifolia*: A comparative study between CPC and classical chromatographic techniques. *Phytochemistry Letters*, *20*, 406–409. https://doi.org/10.1016/j.phytol.2017.01.014
- **Sommer, J.; Kunzmann, A.; Stuthmann, L. E. and Springer, K.** (2022). The antioxidative potential of sea grapes (*Caulerpa lentillifera*, Chlorophyta) can be triggered by light to reach comparable values of pomegranate and other highly nutritious fruits. *Plant Physiology Reports*, *27*, 186–191. https://doi.org/10.1007/s40502-021-00637-6
- **Stuthmann, L. E.; Brix da Costa, B.; Springer, K. and Kunzmann, A.** (2023). Sea grapes (*Caulerpa lentillifera* J. Agardh, Chlorophyta) for human use: Structured review on recent research in cultivation, nutritional value, and post-harvest management. *Journal of Applied Phycology*, *35*, 2957–2983. https://doi.org/10.1007/s10811-023-03031-x
- **Stuthmann, L. E.; Du, H. T.; Brix da Costa, B.; Kunzmann, A. and Springer, K.** (2024). Sea grape (*Caulerpa lentillifera*) aquaculture in Van Phong Bay, Viet Nam: Evaluation of the post-harvest quality. *Journal of Applied Phycology*, *36*, 567–578. https://doi.org/10.1007/s10811-023-03030-y
- Sulaimana, A. S.; Chang, C.; Hou, C.; Yudhistira, B.; Punthi, F.; Lung, C.; Cheng, K.; Santoso, S. P. and Hsieh, C. (2021). Effect of oxidative stress on physicochemical quality of Taiwanese seagrape (*Caulerpa lentillifera*) with the application of alternating current electric field (ACEF) during post-harvest storage. *Processes*, *9*(6), 1011. https://doi.org/10.3390/pr9061011
- Sultana, F.; Wahab, M. A.; Nahiduzzaman, M.; Mohiuddin, M.; Iqbal, M. Z.; Shakil, A.; Mamun, A.; Khan, M. S. R.; Wong, L. and Asaduzzaman, M. (2023). A review of seaweed farming for food and nutritional security, climate change mitigation and adaptation, and women empowerment. *Aquaculture and Fisheries*, *8*(5), 463–480. https://doi.org/10.1016/j.aaf.2022.09.001
- Sun, Y.; Gong, G.; Guo, Y.; Wang, Z.; Song, S.; Zhu, B.; Zhao, L. and Jiang, J. (2018). Purification, structural features and immunostimulatory activity of novel polysaccharides from *Caulerpa lentillifera*. *International Journal of Biological Macromolecules*, *108*, 314–323. https://doi.org/10.1016/j.ijbiomac.2017.12.016
- Sun, Y.; Liu, Z.; Song, S.; Zhu, B.; Zhao, L.; Jiang, J.; Liu, N.; Wang, J. and Chen,
 X. (2020). Anti-inflammatory activity and structural identification of a sulfated polysaccharide CLGP4 from Caulerpa lentillifera. International Journal of

- Biological Macromolecules, *146*, 931–938. https://doi.org/10.1016/j.ijbiomac.2019.09.216
- Syakilla, N.; George, R.; Chye, F. Y.; Pindi, W.; Mantihal, S.; Wahab, N. A.; Fadzwi, F. M.; Gu, P. H. and Matanjun, P. (2022). A review on nutrients, phytochemicals, and health benefits of green seaweed, *Caulerpa lentillifera*. Foods, *11*(18), 2832. https://doi.org/10.3390/foods11182832
- **Tanna, B.; Choudhary, B. and Mishra, A.** (2018). Metabolite profiling, antioxidant, scavenging and anti-proliferative activities of selected tropical green seaweeds reveal the nutraceutical potential of *Caulerpa* spp. *Algal Research*, *36*, 96–105. https://doi.org/10.1016/j.algal.2018.10.019
- Taslim, N. A.; Hardinsyah, H.; Radu, S.; Mayulu, N.; Tsopmo, A.; Kurniawan, R.; Tallei, T. E.; Herlina, T.; Maksum, I. P. and Nurkolis, F. (2024). Functional food candidate from Indonesian green algae *Caulerpa racemosa* (Försskal) J. Agardh by two extraction methods: Metabolite profile, antioxidant activity, and cytotoxic properties. *Journal of Agriculture and Food Research*, *18*, 101513. https://doi.org/10.1016/j.jafr.2024.101513
- Tassakka, A. C.; Iskandar, I. W.; Juniyazaki, A. B.; Zaenab, S.; Alam, J. F.; Rasyid, H.; Kasmiati, K.; Sinurat, E.; Dwiany, F. M.; Martien, R. and Moore, A. M. (2023). Green algae *Caulerpa racemosa* compounds as antiviral candidates for SARS-CoV-2: In silico study. *Narra J*, *3*(2), e179. https://doi.org/10.52225/narra.v3i2.179
- Tesvichian, S.; Sangtanoo, P.; Srimongkol, P.; Saisavoey, T.; Buakeaw, A.; Puthong, S.; Thitiprasert, S.; Mekboonsonglarp, W.; Liangsakul, J.; Sopon, A.; Prawatborisut, M.; Reamtong, O. and Karnchanatat, A. (2024). Sulfated polysaccharides from *Caulerpa lentillifera*: Optimizing the extraction process, structural characteristics, antioxidant capabilities, and anti-glycation properties. *Heliyon*, *10*(2), e24444. https://doi.org/10.1016/j.heliyon.2024.e24444
- **Tian, H.; Liu, H.; Song, W.; Zhu, L. and Yin, X.** (2019). Polysaccharide from *Caulerpa lentillifera*: Extraction optimization with response surface methodology, structure and antioxidant activities. *Natural Product Research*, *35*(20), 3417–3425. https://doi.org/10.1080/14786419.2019.1700507
- Toyen, D.; Suppakul, P.; Siwayaprahm, P.; Thanomchat, P.; Sukatta, U.; Kosawatpat, P.; Banleng, T. and Saenboonruang, K. (2025). Optimizing UV-C treatment for postharvest quality preservation of sea grapes (*Caulerpa lentillifera*). *Postharvest Biology and Technology*, *227*, 113602. https://doi.org/10.1016/j.postharvbio.2025.113602
- **Wells, F. E. and Bieler, R.** (2020). A low number of introduced marine species at low latitudes: A case study from southern Florida with a special focus on Mollusca. *Management of Biological Invasions*, *11*(3), 372–398. https://doi.org/10.3391/mbi.2020.11.3.02

- Windarto, S.; Kamaludin, D. M.; Bella, W.; Sarjito; Pinandoyo; Susilowati, T.; Haditomo, A. H. C. and Harwanto, D. (2023). Effect of coconut (*Cocos nucifera*) water, and aqueous extract of mung bean (*Vigna radiata*) sprouts and moringa (*Moringa oleifera*) leaf on the growth and nutrition of *Caulerpa racemosa*. *International Journal of Aquatic Biology*, *11*(6), 513–522. https://doi.org/10.22034/ijab.v11i6.2044
- **Windarto, S.; Susilowati, T.; Haditomo, A. H. C. and Harwanto, D.** (2024). Effect of exogenous natural plant growth regulators (PGRs) on the morphology, growth, and nutrient of sea grapes (*Caulerpa racemosa*). *Aquaculture International*, *32*, 3545–3562. https://doi.org/10.1007/s10499-023-01337-8
- Windarto, S.; Herawati, V. E.; Wijaya, Y. J.; Indriati, D. A.; Rachmasari, Y. and Elfitasari, T. (2025a). Exploring the physicochemical properties and nutritional value of abundant seaweed species along the Jepara Coast, Indonesia. *Thalassas:* An International Journal of Marine Sciences, *41*, 82. https://doi.org/10.1007/s41208-025-00846-y
- Windarto, S.; Herawati, V.; Wijaya, Y. J. and Indriati, D. A. (2025b). Exploring the bioactive potential of peptides derived from the RuBisCO protein in *Caulerpa racemosa*: An in silico approach. *Biotech Studies*, *34*(2), 71–85. https://doi.org/10.38042/biotechstudies.1756936
- Yang, H.; Tang, B.; Zhou, H.; Zhong, P. and Zhao, L. (2024). Research on the construction of an integrated multi-trophic aquaculture (IMTA) model in seawater ponds and its impact on the aquatic environment. *Water*, *17*(6), 887. https://doi.org/10.3390/w17060887
- Yap, W.; Tay, V.; Tan, S.; Yow, Y. and Chew, J. (2019). Decoding antioxidant and antibacterial potentials of Malaysian green seaweeds: *Caulerpa racemosa* and *Caulerpa lentillifera*. *Antibiotics*, *8*(3), 152. https://doi.org/10.3390/antibiotics8030152
- Yoojam, S.; Ontawong, A.; Lailerd, N.; Mengamphan, K. and Amornlerdpison, D. (2020). The enhancing immune response and anti-inflammatory effects of *Caulerpa lentillifera* extract in RAW 264.7 cells. *Molecules*, *26*(19), 5734. https://doi.org/10.3390/molecules26195734
- You, Y.; Song, H.; Wang, L.; Peng, H.; Sun, Y.; Ai, C.; Wen, C.; Zhu, B. and Song, S. (2022). Structural characterization and SARS-CoV-2 inhibitory activity of a sulfated polysaccharide from *Caulerpa lentillifera*. *Carbohydrate Polymers*, *280*, 119006. https://doi.org/10.1016/j.carbpol.2021.119006
- Zhang, M.; Ma, Y.; Che, X.; Huang, Z.; Chen, P.; Xia, G. and Zhao, M. (2020a). Comparative analysis of nutrient composition of *Caulerpa lentillifera* from different regions. *Journal of Ocean University of China*, *19*(2), 439–445. https://doi.org/10.1007/s11802-020-4222-x

- Zhang, D.; Ramachandran, G.; Mothana, R. A.; Siddiqui, N. A.; Ullah, R.; Almarfadi, O. M.; Rajivgandhi, G. and Manoharan, N. (2020b). Biosynthesized silver nanoparticles using *Caulerpa taxifolia* against A549 lung cancer cell line through cytotoxicity effect/morphological damage. *Saudi Journal of Biological Sciences*, *27*(12), 3421–3427. https://doi.org/10.1016/j.sjbs.2020.09.017
- **Zhou, W.; Wang, Y.; Xu, R.; Tian, J.; Li, T. and Chen, S.** (2025). Comparative analysis of the nutrient composition of *Caulerpa lentillifera* from various cultivation sites. *Foods*, *14*(3), 474. https://doi.org/10.3390/foods14030474
- **Zhu, X.; Healy, L.; Zhang, Z.; Maguire, J.; Sun, W. and Tiwari, B. K.** (2021). Novel postharvest processing strategies for value-added applications of marine algae. *Journal of the Science of Food and Agriculture*, *101*(11), 4444–4455. https://doi.org/10.1002/jsfa.11166
- **Zubia, M.; Draisma, S. G. A.; Morrissey, K. L.; Varela-Alvarez, E. and De Clerck, O.** (2020). Concise review of the genus *Caulerpa* J. V. Lamouroux. *Journal of Applied Phycology*, *32*, 23–39. https://doi.org/10.1007/s10811-019-01868-9