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Enhancing Growth and Feed Efficiency in the Nile Tilapia (*Oreochromis niloticus* L.) Using Black Soldier Larvae Meal (*Hermetia illucens* L.) as a Sustainable Aquafeed Source

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ABSTRACT

This study evaluated the effects of incorporating black soldier fly larvae meal (BSFLM) at 0, 15, 30, and 45% of total ration (D1-D4) on growth performance, feed utilization, body composition, and energy retention of the Nile tilapia (Oreochromis niloticus) fingerlings. A total of 120 fingerlings, with an initial average weight of 365 ± 0.685 g, were acclimated and randomly assigned to 12 aquariums (10 fish per aquarium). BSFLM analysis revealed 35.40% crude protein, 30.00% ether extract, 17.95% ash, and 5511 kcal/kg dry matter gross energy. All experimental diets were formulated to be iso-nitrogenous with minimal differences in caloric content. Results indicated that fish fed diets containing 15, 30, or 45% BSFLM exhibited significant improvements in final weight, total body weight gain, average daily gain, specific growth rate, relative growth rate, feed intake, feed conversion ratio, protein efficiency ratio, and protein productive value. The incorporation of BSFLM also enhanced energy retention, recorded at 16.27, 30.37, 21.36, and 18.75%, and PPV% values of 18.33, 41.25, 31.25, and 38.40% for D1-D4, respectively. Fish body composition was significantly influenced by BSFLM, showing increased CP% and decreased dry matter, EE, ash, and gross energy content, while organic matter remained unaffected. Moreover, inclusion of BSFLM reduced feed costs and improved net economic efficiency. These findings suggest that BSFLM is a promising alternative aquafeed source in the Nile tilapia diets, capable of enhancing growth performance, improving feed utilization, increasing energy retention and protein productivity, and reducing feeding costs. In conclusion, dietary inclusion of BSFLM at different levels positively influenced growth, body composition, energy utilization, and economic efficiency in the Nile tilapia fingerlings, highlighting its potential as a sustainable and cost-effective feed ingredient for aquaculture.







INTRODUCTION

Tilapia (*Oreochromis* spp.) has emerged as one of the most important aquaculture species worldwide, ranking third in global production with nearly 4.5 million tons annually (FAO, 2022). Its popularity is largely attributed to desirable traits such as tolerance to suboptimal water quality, resistance to stress, and relatively low susceptibility to diseases (Suresh & Lin, 1992; Prabu *et al.*, 2019; Sutthi & Van Doan, 2020; Wardani *et al.*, 2021), rapid growth rates (El-Sayed, 1999; Lim, 2006; Abdel-Tawwab *et al.*, 2006), wide acceptance of alternative protein sources (El-Sayed, 1999; Meric *et al.*, 2011; Ng & Romano, 2013), and efficient feed conversion (El-Sayed, 2016; Mengistu *et al.*, 2020). Nutritionally, tilapia require diets containing 35–45% protein during juvenile stages, which gradually decline to 20–30% in sub-adults (Siddiqui *et al.*, 1988; El-Sayed, 2006). Broodstock, however, demand higher protein levels (35–45%) to maintain optimal reproductive performance (Gunasekera *et al.*, 1996a, b; El-Sayed *et al.*, 2003). Lipid requirements are variable, but 10–15% has been suggested for optimum growth, although farmers frequently apply lower levels of 6–8% (El-Sayed, 2006).

In the search for alternative and sustainable feed ingredients, insect meals have gained growing attention. They are nutrient-dense, rich in essential amino acids, and contain beneficial trace elements (Kroeckel et al., 2012; Barroso et al., 2014). Additionally, insect farming offers substantial environmental benefits, including reduced greenhouse gas emissions, lower water demand, and the ability to recycle organic waste into valuable protein biomass (Smetana et al., 2016; Wang & Shelomi, 2017). Among insect species, the black soldier fly (Hermetia illucens; BSF) stands out as particularly promising. It can be reared on agricultural by-products, thereby addressing two major challenges simultaneously: the need for cost-effective feed ingredients and the management of food waste streams (Diener et al., 2009; Nguyen et al., 2015). Furthermore, BSF production is highly scalable and can be automated to support industrial-level feed production (Vo, 2019; Zhang et al., 2020).

Nutritionally, BSF larvae provide 42–58% crude protein and 33–39% lipids (Newton et al., 2005; Kroeckel et al., 2012), making them suitable for incorporation into livestock and aquaculture diets. Previous studies have confirmed their potential as a feed ingredient in chickens (Elwert et al., 2010; Elangovan et al., 2021), several fish species, including the African catfish (Clarias gariepinus), barramundi (Lates calcarifer), rainbow trout (Oncorhynchus mykiss), salmon (Salmo salar), sea bream (Sparus aurata), striped catfish (Pangasianodon hypophthalmus), turbot (Scophthalmus maximus), tench (Tinca tinca), and the Nile tilapia (O. niloticus) (Dietz & Liebert, 2018; Bruni et al., 2020; Fabrikov et al., 2021), pigs (Yu et al., 2019), and ruminants (Fukuda et al., 2022). In tilapia, BSF meal has successfully replaced more than half of dietary protein

without compromising performance when compared with soybean meal-based diets (Dietz & Liebert, 2018).

Given the rising global demand for fish and the escalating cost of conventional protein sources such as fishmeal (Hardy, 2010; Munguti et al., 2021; Tran et al., 2022), the development of cost-effective and sustainable alternatives is urgent. Therefore, this study was conducted to evaluate the effects of incorporating different levels of black soldier fly larvae meal (BSFLM) in the Nile tilapia fingerling diets on growth performance, feed utilization, body composition, nutrient retention, and economic efficiency.

MATERIALS AND METHODS

The present study was conducted to evaluate the effects of incorporating black soldier fly larvae meal (BSFLM) at different dietary levels on the growth performance, feed utilization, body composition, and economic efficiency of the Nile tilapia (*Oreochromis niloticus*) fingerlings. The experimental work was carried out at the Fish Experimental Laboratory, Animal Production Department, Biological Agriculture Research Institute, National Research Centre (NRC), 33 El-Bohouth Street, P.O. Box 12622, Dokki, Cairo, Egypt, in collaboration with Department of Cell Biology, Biotechnology Research Institute, National Research Centre, Giza, Dokki, Giza, 12622, Egypt.

Experimental unit

A total of 120 the Nile tilapia (*Oreochromis niloticus*) fingerlings were acclimated and then randomly allocated into 12 glass aquaria, each stocked with 10 fish. The aquaria $(80 \times 40 \times 30 \text{ cm})$ were filled with 60L of dechlorinated tap water. Fish had an initial mean weight of $36.5 \pm 0.69 \text{g}$ per aquarium. The stock was obtained from the Abbassa Fish Hatchery, Sharkia Governorate, Egypt. Prior to the feeding trial, fish were adapted to laboratory conditions and fed a control diet for two weeks.

Experimental designed and diets

Experimental Nile tilapia (*Oreochromis niloticus*) fingerlings were randomly divided into four dietary treatment groups as follows:

D1 (Control): fed a basal diet without black soldier fly larvae meal (BSFLM).

D2: fed a diet containing 15% BSFLM of the total formulation.

D3: fed a diet containing 30% BSFLM of the total formulation.

D4: fed a diet containing 45% BSFLM of the total formulation.

The feeding trial was conducted over a period of 56 days, from the beginning of May to the end of June 2024.

The composition of the different experimental diets is presented in Table (1).

Item		diets			
	Zero	15%	30%	45%	Price of
	BSFLM	BSFLM	BSFLM	BSFLM	tone LE
	\mathbf{D}_1	\mathbf{D}_2	\mathbf{D}_3	$\mathbf{D_4}$	
Composition of tested diets					
black soldier fly larvae meal	0	15	30	45	25000
Soybean meal (44%)	40	30	9	3	30000
Protein concentration (56%)	17	14	18	12	19000
Ground Yellow corn (8%)	28	26	28	24	11500
Wheat bran (13%)	10	10	10	11	9100
Vegetable oil	3	3	3	3	50000
Vitamin and Minerals mixture*	2	2	2	2	50000
Price of ton fed (LE)	21860	21810	20250	20691	
Price of kg fed (LE)	21.860	21.810	20.250	20.691	

Table 1. Composition of the different experimental diets

*Vitamin and Minerals mixture: contained Vit. A (E672) (IU) 876.19, Vit. D3 (IU) 1141.39, Vit. E 114.30, Vit. K3 7.55, Vit. B1 13.71, Vit. B2 11.44, Vit. B6 15.33, Vit. B12 0.03, Niacin 60.96, Calpan 30.48, Folic Acid 3.04, Biotin 0.37, Vit. C 11.44, Selenium 0.27, Manganese 19.04, Iron 9.15, Iodine 0.77, Zinc 76.19, Copper 3.04, Cobalt 0.37, Choline Chloride 457.14, and Antioxidant 95.23 (Vit. vitamin; IU international unit), Price of tone LE according to 2024, BSFLM: black soldier fly larvae meal

Parameters of growth performance (NRC, 2011)

Body weight gain (BWG) = Final weight - Initial weight.

Survival rate (SR %) = Number of fish at final / Number of fish at start x100.

Specific growth rate (SGR) =

[In final weight (g) - In initial weight (g)] / Experimental days *100

Calculation of feed conversion ratio (FCR)

FCR = total dry matter intake, (TDMI), g / total body weight gain (TBWG), g.

Calculation of crude protein efficiency ratio (CPER)

(PER) = total body weight gain (TBWG), g / total crude protein intake (TCPI), g.

Feed efficiency

Feed efficiency (FE %) = [weight gain (g) / feed intake (g)]

Protein productive value (PPV %) = $[PR_1 - PR_0 / PI] 100$.

Where: PR_1 = is the total fish body protein at the end of the experiment.

 PR_0 = is the total fish body protein at the start of the experiment. PI = Protein intake.

Energy retention percentages (ER %)

The energy retention percentage was calculated according the following equation:

Energy retention (ER %) = $E-E_0 / E_F X 100$

Where: E= the energy in fish carcass (kcal) at the end of the experiment.

 E_0 = the energy in fish carcass (kcal) at the start of the experiment.

 E_F = the energy (kcal) in feed intake

Energy retention was estamited according to Blaxter (1968) and MacRae and Lobley (2003).

The proximate composition of the experimental diets and fish whole-body samples was determined following the standard procedures of the Association of Official Analytical Chemists (AOAC, 2016).

Body composition

At the beginning of the trial, 10 fish were sampled for initial body composition analysis, while at the end of the experiment, 7 fish from each treatment group were randomly selected to determine whole-body composition.

Economic efficiency of diet

The economic efficiency of the experimental diets was evaluated using input—output analysis. Calculations were performed as follows:

Feed cost per kg weight gain = Feed conversion ratio \times Cost of 1 kg diet

Profit per kg weight gain = Sale price per kg gain – Feeding cost per kg gain

Economic efficiency (%) = (Profit per kg gain ÷ Feeding cost per kg gain) × 100

Calculated data

The gross energy (GE, kcal/kg DM) of the experimental diets and fish body composition was estimated according to **Blaxter** (1968) and **MacRae and Lobley** (2003), using the following caloric values: crude protein (CP) = 5.65 kcal/g, ether extract (EE) = 9.40 kcal/g, and crude fiber (CF) and nitrogen-free extract (NFE) = 4.15 kcal/g. Metabolizable energy (ME) was calculated following **NRC** (2011), applying values of 4.50, 8.15, and 3.49 kcal/g for protein, fat, and carbohydrate, respectively. The protein-to-energy ratio (mg CP/kcal ME) was also determined according to **NRC** (2011).

Statistical analysis

All collected data were analyzed using one-way analysis of variance (ANOVA) with SPSS software (Version 26.0, IBM, 2020). Significant differences among treatment means were identified using Duncan's Multiple Range Test (**Duncan, 1955**).

RESULTS

Chemical analysis of experimental diets

As shown in Table (2), black soldier fly larvae meal (BSFLM) contained 35.40% crude protein, 30.00% ether extract, 17.95% ash, 5511 kcal/kg DM gross energy, 432.91 kcal/kg DM metabolizable energy, and a protein-to-energy ratio of 81.77mg CP/kcal ME which was calculated according to **NRC** (2011).

Table 2. Chemical analysis of different experimental diets

Item		Experime		erimental di	ental diets	
	BSFLM	Zero	15%	30%	45%	
		BSFLM	BSFLM	BSFLM	BSFLM	
		\mathbf{D}_1	\mathbf{D}_2	D ₃	\mathbf{D}_4	
Moisture	3.66	6.24	5.96	5.35	5.33	
Dry matter (DM)	96.34	93.76	94.04	94.65	94.67	
Chemical analysis on DM basis						
Organic matter (OM)	82.05	90.89	88.39	85.18	83.20	
Crude protein (CP)	35.40	29.26	29.50	29.88	29.43	
Crude fiber (CF)	8.31	4.57	4.56	4.45	4.71	
Ether extract (EE)	30.00	3.35	5.69	8.13	10.80	
Nitrogen free extract (NFE)	8.34	53.71	48.64	42.72	38.26	
Ash	17.95	9.11	11.61	14.82	16.80	
Gross energy kcal/ kg DM	5511	4387	4409	4410	4461	
Gross energy cal/ g DM	5.511	4.387	4.409	4.410	4.461	
Metabolizable energy kcal/ kg DM	432.91	346.42	348.88	349.81	353.98	
Protein energy ratio (mg CP/ Kcal ME)	81.77	84.46	84.56	85.42	83.14	

Gross energy (kcal/ kg DM) was calculated according to (Blaxter1968; MacRae & Lobley 2003).

Where, each g CP = 5.65 Kcal, g EE = 9.40 kcal and g CF and NFE = 4.15 Kcal.

Metabolizable energy (ME): Calculated according to **NRC (2011)** using values of 4.50, 8.15 and 3.49 Kcal for protein, fat and carbohydrate, respectively.

BSFLM: black soldier fly larvae meal.

The proximate composition of the experimental diets revealed crude protein (CP) contents of 29.26, 29.50, 29.88, and 29.43% for D1, D2, D3, and D4, respectively. Crude fiber (CF) levels were similar across treatments (4.56–4.71%). Ether extract (EE) increased progressively with BSFLM inclusion, ranging from 3.35% in D1 to 10.80% in D4. Ash content rose from 9.11% in D1 to 16.80% in D4, while nitrogen-free extract (NFE) decreased from 53.71% to 38.26% with increasing BSFLM levels. Gross energy ranged from 4387 to 4461 kcal/kg DM, metabolizable energy from 346.42 to 353.98 kcal/kg DM, and protein-to-energy ratios from 83.14 to 85.42 mg CP/kcal ME. Overall, BSFLM proved to be a rich source of protein and energy, and the formulated diets were isonitrogenous (29.26–29.88% CP) with only minor differences in caloric values, all of which were within the nutritional requirements of the Nile tilapia fingerlings.

Growth performance and survival

As presented in Table (3), the incorporation of BSFLM at different dietary levels (15, 30, and 45%) significantly (P < 0.05) improved final weight (FW), total body weight

gain (TBWG), average daily gain (ADG), specific growth rate (SGR), and relative growth rate (RGR) compared with the control group. The highest growth performance values were observed in fish fed the diet containing 15% BSFLM (D2), followed by D3 and D4, while the control group (D1) showed the lowest results. Survival rate was 100% across all treatments, with no recorded mortality in any of the experimental groups.

Table 3. Growth performance, specific growth rate and survival ratio of the Nile tilapia fingerlings fed diets containing different levels of black soldier fly larvae meal

Item	Experimental diets					
	Zero	15%	30%	45%		
	BSFLM	BSFLM	BSFLM	BSFLM		Sign.
	G_1	G_2	G ₃	G ₄	SEM	P<0.05
Number of fish	30	30	30	30	-	-
IW, g	363	367	366	364	0.685	NS
FW, g	472°	615 ^a	520 ^b	516 ^b	15.780	*
TBWG, g	109 ^c	248 ^a	154 ^b	152 ^b	15.384	*
Duration trial	56 days				-	-
ADG, g	1.95°	4.43 ^a	2.75 ^b	2.71 ^b	0.275	*
SGR	0.21°	0.40 ^a	0.27 ^b	0.27 ^b	0.021	*
RGR	0.30°	0.68a	0.42 ^b	0.42 ^b	0.042	*
Starter number	30	30	30	30	-	-
End number of	30	30	30	30	-	-
SR %	100	100	100	100	-	-
Dead number	Zero	Zero	Zero	Zero	-	-
Mortality rate %	Zero	Zero	Zero	Zero	-	-

a, b and c: Means in the same row having different superscripts differ significantly (P<0.05).

SEM: Standard error of the mean NS: Not significant. *: Significant at (P<0.05). IW: Initial weight, g.

FW: Final weight, g. TBWG: Total body weight gain, g.

ADG: Average daily gain, g. SGR: Specific growth rate. RGR: Relative growth rate.

SR: Survival ratio. BSFLM: black soldier fly larvae meal.

Feed utilization of the different experimental groups

As shown in Table (4), feed utilization parameters were positively influenced by the inclusion of BSFLM in the Nile tilapia diets. Feed intake (FI) and crude protein intake (CPI) were significantly higher (P<0.05) in groups fed diets containing BSFLM (15%, 30%, and 45%) compared with the control group (D1, 0% BSFLM). The highest FI (1133 g) and CPI (334.24 g) were recorded in fish fed the 15% BSFLM diet (D2).

Feed conversion ratio (FCR) also improved significantly (P < 0.05) in groups D2, D3, and D4 compared to the control, with the most efficient conversion observed in D2 (4.569), followed by D3 (6.838) and D4 (6.875), whereas the control group exhibited the poorest FCR (9.229).

Similarly, protein efficiency ratio (PER) showed significant (P < 0.05) improvement in all BSFLM-supplemented groups (D2, D3, and D4), with the highest value in D2.

Item						
	Zero BSFLM	15% BSFLM	30% BSFLM	45% BSFLM		Sign.
	G_1	G_2	G ₃	G ₄	SEM	P<0.05
TBWG, g	109°	248a	154 ^b	152 ^b	15.384	*
FI, g	1006°	1133a	1053 ^b	1045 ^b	13.961	*
FCR	9.229°	4.569a	6.838 ^b	6.875 ^b	0.508	*
FCP%	29.26	29.50	29.88	29.43	-	-
CPI, g	294.36 ^d	334.24a	314.64 ^b	307.54 ^c	4.359	*
PER	0.3703°	0.7420a	0.4894 ^b	0.4942 ^b	0.041	*

Table 4. Feed utilization of the Nile tilapia fingerlings fed diets containing different levels of black soldier fly larvae meal

a, b, c and d. Means in the same row having different superscripts differ significantly (P < 0.05).

SEM: Standard error of the mean, NS: Not significant,*: Significant at (P<0.05), BWG: Total body weight gain, FI: Feed intake, FCR: Feed conversion ratio, FCP%: Feed crude protein percentages, CPI: Crude protein intake, PER: Protein efficiency ratio,BSFLM: black soldier fly larvae meal.

Fish body composition of different experimental groups

Data presented in Table (5) indicate that the inclusion of BSFLM at different dietary levels (15, 30, and 45% of the total diet formulation for D2, D3, and D4, respectively) had a significant effect (P < 0.05) on most parameters of the Nile tilapia whole body composition, except for organic matter (OM) content, when compared with the control group (D1).

Incorporation of BSFLM significantly increased crude protein (CP%) and moisture content, while it significantly reduced dry matter (DM%), ether extract (EE%), ash%, and gross energy (GE; kcal/100 g DM). The highest CP value (62.51%) was recorded in fish from D4, which received the 45% BSFLM diet.

Table 5. Fish body composition of initial and different experimental groups of the Nile tilapia fingerlings fed diets with different levels of black soldier fly larvae meal

Item	Fish body	Experimental diets					
	composition of initial fish	Zero BSFLM	15% BSFLM	30% BSFLM	45% BSFLM		Sign.
		G_1	G_2	G_3	G ₄	SEM	P<0.05
Moisture	75.46	70.89 ^d	73.37 ^a	72.31 ^c	72.94 ^b	0.287	*
DM	24.54	29.11 ^a	26.63 ^d	27.69 ^b	27.06°	0.287	*
Chemical analysis on DM basis Chemical analysis on DM basis			ısis	-	-		
OM	83.68	85.10	85.13	86.73	86.17	0.253	NS
CP	56.17	54.63 ^d	55.94°	58.44 ^b	62.51 ^a	0.906	*
EE	27.51	30.47 ^a	29.19 ^b	28.29°	23.66 ^d	0.777	*
Ash	16.32	14.90 ^a	14.87 ^a	13.27 ^c	13.83 ^b	0.211	*
GE1	575.95	595.08 ^a	590345 ^b	596.11a	575.59 ^c	2.501	*
GE2	5.7595	5.9508 ^a	5.9045 ^b	5.9611a	5.7559 ^c	0.0251	*

a, b, c and d: Means in the same row having different superscripts differ significantly (P<0.05).

SEM: Standard error of the mean, NS: Not significant, *: Significant at (P < 0.05), DM: Dry matter

OM: Organic matter, CP: Crude protein, EE: Ether extract, GE1: Gross energy kcal/ 100g.

GE2: Gross energy cal/ g DM, BSFLM: black soldier fly larvae meal.

Energy retention and protein productive value percentages

Data illustrated in Table (6) show that energy retention percentage (ER%) and protein productive value percentage (PPV%) were significantly (P<0.05) increased in the Nile tilapia fed diets containing BSFLM at 15, 30, and 45% (D2, D3, and D4, respectively) compared with the control group (D1) that received 0% BSFLM.

The corresponding ER% values were 16.27, 30.37, 21.36, and 18.75% for D1, D2, D3, and D4, respectively. Similarly, the PPV% values were 18.33, 41.25, 31.25, and 38.40% for D1, D2, D3, and D4, respectively.

Table 6. Energy retention (ER) and protein productive value (PPV) % of the Nile tilapia fingerlings fed diets containing different levels of black soldier fly larvae meal

Item	Experimental diets					
	Zero	15%	30%	45%		
	BSFLM	BSFLM	BSFLM	BSFLM		Sign.
	G ₁	G ₂	G ₃	G ₄	SEM	P<0.05
IW	363	367	366	364	0.685	NS
FW	472°	615 ^a	520 ^b	516 ^b	15.780	*
		Calculation the	energy retention	ı		
ECFBW	5.9508a	5.9045 ^b	5.9611a	5.7559 ^c	0.025	*
TEEBF	2809 ^d	3631a	3100 ^b	2970°	93.275	*
ECIBF		5.75	95		-	-
TESBF	2091	2114	2108	2096	3.976	NS
ERBF	718 ^d	1517a	992 ^b	874 ^c	90.869	*
EFI	4.387	4.409	4.410	4.461	-	-
QFI	1006°	1133a	1053 ^b	1045 ^b	13.961	*
TEFI	4413°	4995a	4644 ^b	4662b	62.758	*
ER%	16.27 ^d	30.37 ^a	21.36 ^b	18.75°	1.615	*
	Calculati	on the protein p	roductive value	(PPV) %		
CPFBC%	54.63 ^d	55.94°	58.44 ^b	62.51a	0.096	*
PR ₁	257.85 ^d	344.03a	303.89°	322.55 ^b	9.618	*
CPIBFC%		56.	17		-	-
PR ₂	203.90	206.14	205.58	204.46	0.385	NS
PR ₃	53.95 ^d	137.89a	98.31°	118.09 ^b	9.427	*
CPFE%	29.26	29.50	29.88	29.43	_	-
TPI	294.36 ^d	334.24a	314.64 ^b	307.54 ^c	4.359	*
PPV%	18.33 ^d	41.25a	31.25°	38.40 ^b	2.687	*

a, b, c and d: Means in the same row having different superscripts differ significantly (P<0.05).

SEM: Standard error of the mean, NS: Not significant, *: Significant at (P<0.05), IW: Initial weight, g. FW: Final weight, g, ECFBW: Energy content in final body fish (cal / g), TEBF: Total energy at the end in body fish (E), Energy content in initial body fish (cal / g), TESBF: Total energy at the start in body fish (E0) Energy retained in body fish (E-E0), EFI: Energy of the feed intake (Cal / g feed), QFI: Quantity of feed intake

Economical evaluation of different experimental groups

Data presented in Table (7) show that the incorporation of BSFLM into feed formulation reduced the overall feed cost compared with the control diet. The cost of feed formulation decreased from 21.860 LE/kg feed in the control diet (D1) to 21.810, 20.250, and 20.691 LE/kg feed in diets D2, D3, and D4, respectively.

The net improvement in feeding cost (%) was 0.00, 50.38, 24.00, and 24.14% for D1, D2, D3, and D4, respectively.

Table 7. Economical evaluation of the Nile tilapia fingerlings fed diets containing different levels of black soldier fly larvae meal

Item		Experimental diets				
	Zero	15%	30%	45%		
	BSFLM	BSFLM	BSFLM	BSFLM		
	G_1	G ₂	G_3	G ₄		
Costing of kg feed (LE)	21.860	21.810	20.250	20.691		
Relative to control (%)	100	99.77	92.63	94.65		
Feed conversion ratio (FCR)	9.229	4.569	6.838	6.875		
Feeding cost (LE) per (Kg weight gain)	201.75	99.65	138.47	142.25		
Relative to control (%)	100	49.39	68.63	70.51		
Net improving in feeding cost (%)	Zero	50.38	24.00	24.14		

Diet formulation calculated according to the local prices at year 2024, as presented in Table (1). Feed cost (L.E) FCR×FI. Cost per Kg diet, BSFLM: black soldier fly larvae meal.

DISCUSSION

The proximate composition of black soldier fly larvae meal (BSFLM) in the present study demonstrates its potential as a valuable feed ingredient for aquafeeds. BSFLM contained 35.40% crude protein and 30.00% ether extract, along with considerable ash (17.95%) and high gross and metabolizable energy values (5511 and 432.91 kcal/kg DM, respectively). The calculated protein–energy ratio (81.77 mg CP/kcal ME) further confirms that BSFLM is nutritionally dense and suitable for fish diets. Formulation of the experimental diets showed that all rations were iso-nitrogenous, with crude protein levels ranging from 29.26% to 29.88%, which fall within the recommended protein requirement for the Nile tilapia. Crude fiber remained relatively constant (4.56– 4.71%), while ether extract increased gradually with higher inclusion of BSFLM (3.35%) in D1 to 10.80% in D4). Ash content varied more markedly (9.11–16.80%), which may reflect the mineral contribution of BSFLM. Nitrogen-free extract decreased with increasing BSFLM levels, indicating a shift from carbohydrate-based to lipid-based energy sources. Despite these differences, gross energy values (4387–4461 kcal/kg DM) and metabolizable energy (346.42–353.98 kcal/kg DM) remained within a narrow range, suggesting that the diets were energetically comparable. Protein–energy ratios (83.14– 85.42 mg CP/kcal ME) also confirmed that all formulations were well balanced for tilapia nutritional needs. These results are consistent with earlier reports emphasizing the suitability of insect meals as alternative protein sources in aquafeeds. Tacon and Metian (2008) and Kishawy et al. (2022) highlighted the increasing demand and rising costs of conventional protein sources such as fish meal, animal by-products, and plant proteins due to the intensification of aquaculture. Similarly, **Obirikorang** et al. (2015) argued that modern feed formulation strategies must consider not only nutrient content but also cost, sustainability, and ingredient availability. The present findings align with this perspective, supporting the use of insect meals as sustainable replacements. Several studies have documented the successful incorporation of insect meals into aquafeeds for different species. Fitches and Smith (2018) demonstrated the nutritional adequacy of insect larvae meals for the Nile tilapia, while Cummins et al. (2017) reported similar findings in Pacific white shrimp. Comparable results have also been reported for the rainbow trout (Renna et al., 2017) and Atlantic salmon (Belghit et al., 2019). Abozaid et al. (2024c) further confirmed that Galleria mellonella larvae are highly nutritive, with even higher lipid content than BSFLM, contributing to elevated gross and metabolizable energy values. These collective findings suggest that insect-based ingredients are not only nutritionally sufficient but also offer a sustainable and cost-effective solution for aquaculture feed production.

In the present study, the incorporation of BSFLM at different inclusion levels significantly (P < 0.05) enhanced the growth performance of the Nile tilapia, as reflected by the increased FW, TBWG, ADG, SGR, and RGR values. The best growth responses were obtained with the 15% BSFLM diet (D22), while survival remained 100% with zero mortality across all treatments. These findings confirm the suitability of BSFLM as a safe protein source for the Nile tilapia. Our results are consistent with those of **Kishawy** et al. (2022), who reported that substituting fishmeal with processed BSFLM up to 100% did not significantly affect growth performance of the Nile tilapia. Similarly, several studies have demonstrated that BSFLM can replace fishmeal without compromising performance in different aquaculture species, including the rainbow trout (Renna et al., 2017), the Atlantic salmon (Belghit et al., 2019), the Japanese sea bass (Wang et al., 2019), the European sea bass (Abdel-Tawwab et al., 2020), the Siberian sturgeon (Caimi et al., 2020), and the Nile tilapia (Tippayadara et al., 2021). Devic et al. (2018) also showed that up to 80% fishmeal replacement with BSFLM did not negatively affect final body weight or survival in the Nile tilapia. The positive effects observed in this study may be attributed to the high nutritional value of BSFLM, particularly its balanced amino acid and fatty acid profile, which closely resembles conventional protein sources. Our results are further supported by Abozaid et al. (2024c), who observed improvements in growth performance and survival when soybean meal was partially replaced with insect meal. Likewise, Fayed et al. (2023) reported that inclusion of BSFLM up to 25% significantly improved growth performance, feed conversion, and survival in the Nile tilapia, although performance declined at 30% replacement, likely due to increased chitin content.

The role of chitin in limiting nutrient digestibility has been highlighted in several studies. **Alegbeleye** *et al.* (2012) and **Karlsen** *et al.* (2017) pointed out that chitin, when included at high levels, can act as a filler with low digestible energy and hinder amino acid absorption, leading to reduced growth rates. This may explain why in our trial the best performance was obtained at 15% BSFLM, while higher levels showed diminishing

responses. Similar declines at high inclusion levels have been reported in the Nile tilapia (**Dietz & Liebert, 2018**), meager (**Guerreiro** *et al.*, **2020**), and other species.

Incorporation of BSFLM into the Nile tilapia diets significantly enhanced feed utilization parameters. Feed intake (FI) and crude protein intake (CPI) increased in groups fed BSFLM-based diets compared to the control group (0% BSFLM). Notably, the group fed 15% BSFLM (D2) exhibited the highest FI (1133 g) and CPI (334.24 g). Feed conversion ratio (FCR) also improved significantly (P < 0.05) in fish fed BSFLM diets (D2, D3, and D4), with the best efficiency observed in D2 (4.569) compared to D1 (9.229), D3 (6.838), and D4 (6.875). Similarly, protein efficiency ratio (PER) improved in all groups receiving BSFLM. These results suggest that partial replacement of conventional protein with BSFLM, particularly at 15%, promotes better nutrient utilization in tilapia. Comparable outcomes have been reported in earlier studies. Kishawy et al. (2022) noted that replacing fish meal protein with partially defatted BSFLM at 25-100% did not adversely affect FCR, specific growth rate (SGR), PER, or protein retention efficiency (PRE), and that all diets were equally accepted by fish. Likewise, Rawski et al. (2020) observed that feed intake did not differ among treatments, but both FCR and PER improved in insect-fed groups, with the greatest improvements at lower inclusion levels (5% BSFLM). These findings support the current results, where the most favorable feed conversion occurred at moderate inclusion (15% BSFLM), while higher levels yielded smaller improvements.

The positive influence of BSFLM on feed utilization may be attributed to several functional properties of insect-derived nutrients. Insects are rich in antimicrobial peptides (AMPs), medium-chain fatty acids (notably lauric acid), and chitin, all of which have been reported to enhance gut health, nutrient absorption, and immune responses (Park et al., 2015; Kasumyan, 2016; Lee et al., 2020; Moretta et al., 2020). Such bioactive components may explain the improved feed conversion and protein efficiency observed in the present study.

In addition, other feed additives such as methionine supplementation and probiotics (e.g., *Saccharomyces cerevisiae*) have been shown to improve feed efficiency in the Nile tilapia. **Abozaid** *et al.* (2024a, b) reported enhanced FCR and PER when methionine or yeast were included in tilapia diets, while **Abozaid** *et al.* (2024c) demonstrated significant improvements in FI, FCR, CPI, and PER when soybean meal was partially replaced with *Galleria mellonella* larvae meal. These findings align with the present study and reinforce the concept that alternative protein sources and functional ingredients can improve nutrient utilization and reduce dependence on traditional fish meal.

Inclusion of BSFLM at different levels (15–45%) significantly affected the whole-body composition of the Nile tilapia, with the exception of organic matter (OM). Incorporation of BSFLM resulted in higher crude protein (CP%) and moisture levels,

whereas dry matter (DM%), ether extract (EE%), ash, and gross energy (GE) declined. The highest CP value (62.51%) was obtained in fish fed 45% BSFLM (D4), suggesting that BSFLM can enhance protein deposition in tilapia muscle tissue. These findings are partly consistent with earlier reports. **Kishawy** *et al.* (2022) observed no significant differences in DM and CP contents of the Nile tilapia fed PD-BSFLM up to 100%, although they reported reductions in lipid and ash content, including calcium and phosphorus, at higher replacement levels. Similarly, studies of **Dumas** *et al.* (2018) and **Amer** *et al.* (2021) on the rainbow trout, and that of **Reyes** *et al.* (2020) on the sea bass found that dietary BSFLM did not alter whole-body CP content, but consistently reduced lipid levels. The reduction in lipid deposition has been attributed to the high lauric acid content of BSFLM, which is easily oxidized and less likely to accumulate in fish tissues (Williams *et al.*, 2006; Liland *et al.*, 2017).

Declines in ash content with higher BSFLM inclusion have also been reported in the rainbow trout (Caimi et al., 2021) and tilapia (Kishawy et al., 2022). This reduction may be linked to the relatively low mineral content of insect meals compared to fish meal. On the other hand, several studies confirmed that insect meal inclusion does not negatively affect protein quality or amino acid composition of fish fillets (Mancini et al., 2018; Belghit et al., 2019; Caimi et al., 2021). However, the relatively low methionine level in BSFLM has been highlighted as a factor that may limit body methionine content unless supplemented (Liland et al., 2017). These findings are consistent with Abozaid et al. (2024c), who reported enhanced ER% in the Nile tilapia fed Galleria mellonella larvae meal, although PPV% declined at higher inclusion levels, highlighting possible differences between insect species or inclusion thresholds. Similarly, Abozaid et al. (2024b) noted significant improvements in ER% and PPV% when diets were supplemented with methionine, while **Abozaid** et al. (2024a) observed enhanced PPV% but reduced ER% when Saccharomyces cerevisiae was incorporated. Collectively, these studies suggest that dietary interventions improving amino acid balance and digestibility can enhance nutrient utilization in tilapia. The improved ER and PPV observed in the current study may be partly attributed to the balanced protein-energy ratio of BSFLM diets, along with functional properties of insect-derived nutrients such as medium-chain fatty acids and chitin that support efficient nutrient metabolism.

Economic evaluation further confirmed the advantages of BSFLM inclusion. Feed costs decreased from 21.860 LE/kg in the control diet to 21.810, 20.250, and 20.691 LE/kg in D2, D3, and D4, respectively, with net feed cost improvements of 50.38%, 24.00%, and 24.14%. This aligns with **Kishawy** *et al.* (2022), who reported reduced feed cost and enhanced economic efficiency with partial or full replacement of fish meal by BSFLM. Similar reductions in feed cost and improvements in profitability with BSFLM inclusion have been observed in tilapia (Abdel-Tawwab *et al.*, 2020; Fayed *et al.*, 2023) and other aquaculture species (Stejskal *et al.*, 2020; Rawski *et al.*, 2021). Luna *et al.* (2019) further emphasized that reliance on fish meal and fish oil is increasingly

unsustainable and uneconomical, underscoring the importance of alternative protein sources such as insect meals.

CONCLUSION

The present study demonstrated that partial replacement of fishmeal with black soldier fly larvae meal (BSFLM) at levels of 15–45% in the Nile tilapia diets significantly improved energy retention (ER%) and protein productive value (PPV%) compared to the control diet. The best results were obtained at 15% and 30% inclusion levels, indicating efficient nutrient utilization and protein deposition. Moreover, incorporation of BSFLM substantially reduced feed formulation costs and improved economic efficiency, confirming its potential as a cost-effective and sustainable alternative to fishmeal. Therefore, BSFLM can be recommended as a promising protein source in the Nile tilapia diets to enhance growth performance, nutrient retention, and profitability, while contributing to the sustainability of aquaculture production systems.

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REFERENCES

- **Abdel-Tawwab, M.; Khattab, Y.A.E.; Ahmad, M.H. and Shalaby, A.M.E.** (2006). Compensatory growth, feed utilization, whole-body composition, and hematological changes in starved juvenile Nile tilapia, *Oreochromis niloticus* L. J. Appl. Aquac., 18, 17-36.
- **Abdel-Tawwab, M.; Khalil, R.H.: Metwally, A.A.; Shakweer, M.S.; Khallaf, M.A.** and **Abdel-Latif, H.M.R.** (2020). Effects of black soldier fly (*Hermetia illucens L.*) larvae meal on growth performance, organs-somatic indices, body composition and hemato-biochemical variables of European sea bass, *Dicentrarchus labrax*. Aqua., 522: 735136.
- **Abozaid1, H.; Elnadi, A.S. M.; Aboelhassan, D. M.; El-Nomeary, Y.A. A.; Omer H.A. A. and Abbas, W. T.** (2024a). Using the dried Yeast (*Saccharomyces cerevisiae*) as a growth promoter in the Nile Tilapia (*Oreochromis niloticus*) diets. Egypt. J. Aquat. Biol. Fish., 28 (2): 699-716.
- **Abozaid, H.; Elnadi, A.S. M.; Omer, H.A. A.; El-Nomeary, Y. A.A. Abelhassan D. M. and** Abbas **W.T.** (2024b). Productive performance, feed utilization, biochemical parameters, and economic evaluation of the Nile Tilapia (*Oreochromis niloticus*) fed diets containing different levels of methionine. Egypt. J. Aquat. Biol. Fish., 28 (4): 161-176 (2024)

- Abozaid, H.; Elnadi, A.S.M..; Omer, H.A.A.; El-Nomeary, Y.A.A.; Aboelhassan, D M.; Awad, E.; Abbas, W. T.; Ebadah, I. M.A. and Moawad S. S. (2024c). Effect of replacing dietary soybean meal with *Galleria mellonella* larvae powder on growth performance of the Nile Tilapia (*Oreochromis niloticus*). Egypt. J. Aquat. Biol. Fish., 28(5): 123-148.
- **Ai, C. H.; Li, B. and Xia, J.H.** (2022). Mapping QTL for cold-tolerance trait in a GIFT-derived tilapia line by ddRAD-seq." *Aquaculture* 556 (2022): 738273.
- **Aini, N.; Nugroho, R.A. and Hariani, N.** (2018). Growth and survival evaluation of *Oreochromis sp.* fed *Hermetia illucens* larva and *Manihot esculenta* leaves meal. Biosaintifika: J. Biol. Biol. Educ., 10: 565-573.
- Alegbeleye, W.O.; Obasa, S.O.; Olude, O.O.; Otubu, K. and Jimoh, W. (2012). Preliminary evaluation of the nutritive value of the variegated grasshopper (*Zonocerus variegatus* L.) for African catfish *Clarias gariepinus* (Burchell. 1822) fingerlings. Aquacult. Res., 43: 412-420.
- Amer, A.A.; El-Nabawy, E.S.M.; Gouda, A.H. and Dawood, M.A. (2021). The addition of insect meal from Spodoptera littoralis in the diets of Nile tilapia and its effect on growth rates, digestive enzyme activity and health status. Aquac. Res., 52 (11): 5585-5594.
- **AOAC** (2016). Official Methods of Analysis, 18th ed. Association of Official Analytical Chemists, Washington, DC, USA (2005). All content following this page was uploaded by Hayder N. Al-Mentafji on 02 February 2016.
- Barroso, F.G.; de Haro, C.; Sanchez-Muros, M.J.; Venegas, E.; Martinez-Sanchez, A. and Perez-Banon, C. (2014). The potential of various insect species for use as food for fish. Aqua., 422, 193-201.
- Belghit, I.; Liland, N. S.; Waagbø, R.; Biancarosa, I.; Pelusio, N.; Li, Y. and Lock, E.J. (2019). Black soldier fly larvae meal can replace fish meal in diets of sea-water phase Atlantic salmon (*Salmo salar*). Aquaculture, 503, 609-619. https://doi.org/10.1016/j.aquaculture.2018.12.032
- **Benli, A.C.K.; Koksal, G. and Ozkul, A.** (2008). Sublethal ammonia exposure of Nile tilapia (*Oreochromis niloticus* L.): Effects on gill, liver and kidney histology. Chemosphere, 72, 1355-1358.
- **Blaxter, K.L.** (1968). The energy metabolism of ruminants. 2^{nd} ed. Charles Thomas Publisher. Spring field. Illinois, USA.
- Bordignon, F.; Gasco, L.; Birolo, M.; Trocino, A.; Caimi, C.; Ballarin, C.; Bortoletti, M.; Nicoletto, C.; Maucieri, C. and Xiccato, G. (2022). Performance and fillet traits of rainbow trout (*Oncorhynchus mykiss*) fed different levels of *Hermetia illucens* meal in a low-tech aquaponic system. Aqua., 546, 737279.

- Bruni, L.; Belghit, I.; Lock, E.J.; Secci, G.; Taiti, C. and Parisi, G. (2020). Total replacement of dietary fish meal with black soldier fly (*Hermetia illucens*) larvae does not impair physical, chemical or volatile composition of farmed Atlantic salmon (*Salmo salar* L.). J. Sci. Food Agric., 100, 1038-1047.
- Bruni, L.; Milanovic, V.; Tulli, F.; Aquilanti, L. and Parisi, G. (2022). Effect of diets containing full-fat *Hermetia illucens* on rainbow trout microbiota: A dual cultivation-independent approach with DGGE and NGS. Aqua., 553, 738109.
- Caimi, C.; Renna, M.; Lussiana, C.; Bonaldo, A.; Gariglio, M.; Meneguz, M. and Gasco, L., (2020). First insights on Black Soldier Fly (*Hermetia illucens L.*) larvae meal dietary administration in Siberian sturgeon (*Acipenser baerii Brandt*) juveniles. Aqua., 515, 734539.
- Caimi, C.; Biasato, I.; Chemello, G.; Oddon, S.B.; Lussiana, C.; Malfatto, V.M. and Gasco, L. (2021). Dietary inclusion of a partially defatted black soldier fly (*Hermetia illucens*) larva meal in low fishmeal-based diets for rainbow trout (*Oncorhynchus mykiss*). J. Anim. Sci. Biotechnol., 12, 50.
- Cummins, V.C.; Rawles, S.D.; Thompson, K.R.; Velasquez, A.; Kobayashi, Y.; Hager, J. and Webster, C.D. (2017). Evaluation of black soldier fly (*Hermetia illucens*) larvae meal as partial or total replacement of marine fish meal in practical diets for Pacific white shrimp (*Litopenaeus vannamei*). Aqua., 473, 337-344.
- **Devic, E.; Leschen, W.; Murray, F. and Little, D.C.** (2018). Growth performance, feed utilization and body composition of advanced nursing Nile tilapia (*Oreochromis niloticus*) fed diets containing Black Soldier Fly (*Hermetia illucens*) larvae meal. Aquac. Nutr., 24, 416-423.
- **Diener, S.; Zurbrugg, C. and Tockner, K. (2009)**. Conversion of organic material by black soldier fly larvae: Establishing optimal feeding rates. Waste Manag. Res., 27, 603-610.
- **Dietz, C. and Liebert, F. (2018)**. Does graded substitution of soy protein concentrate by an insect meal respond on growth and N-utilization in Nile tilapia (*Oreochromis niloticus*)? Aquac. Rep., 12, 43-48.
- **Dumas, A.; Raggi, T.; Barkhouse, J.; Lewis, E. and Weltzien, E.** (2018). The oil fraction and partially defatted meal of black soldier fly larvae (*Hermetia illucens*) affect differently growth performance, feed efficiency, nutrient deposition, blood glucose and lipid digestibility of rainbow trout (*Oncorhynchus mykiss*). Aqua., 492: 24-34.
- **Duncan, D.B.** (1955). Multiple Rang and Multiple F-Test Biometrics, 11: 1-42. https://doi.org/10.2307/3001478https://www.jstor.org/stable/3001478

- Elangovan, A.V.; Udayakumar, A.; Saravanakumar, M.; Awachat, V.B.; Mohan, M.; Yandigeri, M.S.; Krishnan, S.; Mech, A.; Rao, S.B.N.; Giridhar, K. and *et al.* (2021). Effect of black soldier fly, *Hermetia illucens* (Linnaeus) prepupae meal on growth performance and gut development in broiler chicken. Int. J. Trop. Insect Sci., 41, 2077–2082.
- **El-Sayed, A.F.M. and Teshima, S.** (1992). Protein and energy requirements of Nile tilapia, Oreochromis niloticus, fry. Aqua., 103, 55-63.
- **El-Sayed, A.F.M.** (1999). Alternative dietary protein sources for farmed tilapia, *Oreochromis spp.* Aqua., 179, 149-168.
- **El-Sayed, A.F.M.; Mansour, C.R. and Ezzat, A.A.** (2003). Effects of dietary protein level on spawning performance of Nile tilapia (*Oreochromis niloticus*) brood stock reared at different water salinities. Aqua., 220, 619-632.
- El-Sayed, A.F.M. (2006) Tilapia Culture; CABI Publishing: Wallingford, UK,
- **El-Sayed, A.F.M.** (2016). On-farm feed management practices for Nile tilapia (*Oreochromis niloticus*) in Egypt. In On-Farm Feeding and Feed Management in Aquaculture; Hasan, M.R., New, M.B., Eds.; FAO Fisheries and Aquaculture Technical Paper No 583; FAO: Rome, Italy, 2016; pp. 101-129.
- **Elwert, C.; Knips, I. and Katz, P.** (2010). A novel protein source: Maggot meal of the black soldier fly (*Hermetia illucens*) in broiler feed. In Tagung Schweine- und Geflügelernährung; Gierus, M., Kluth, H., Eds.; Institut für Agrar- und Ernährungswissenschaften, Universität Halle-Wittenberg: Halle, Germany, 2010; pp. 140-142.
- Fabrikov, D.; Barroso, F.G.; Sanchez-Muros, M.J.; Hidalgo, M.C.; Cardenete, G.; Tomas-Almenar, C.; Melenchon, F. and Guil-Guerrero, J.L. (2021). Effect of feeding with insect meal diet on the fatty acid compositions of sea bream (*Sparus aurata*), tench (*Tinca tinca*) and rainbow trout (*Oncorhynchus mykiss*) fillets. Aqua., 545, 737170.
- **FAO.** (2022). The State of World Fisheries and Aquaculture, towards Blue Transformation; FAO: Rome, Italy, 2022.
- **Fayed, W.M.; Mansour, A.T.; Zaki, M.A.; Omar, E.A.; Moussa, Nour A.A., Taha E.M. and Sallam G.R.** (2023). Water quality change, growth performance, health status in response to dietary inclusion of black soldier fly larvae meal in the diet of Nile tilapia, *Oreochromis niloticus*. Ann. Anim. Sci., 24(2): 533-544.
- **Fisher, H.J.; Collins, S.A.; Hanson, C.; Mason, B.; Colombo, S.M. and Anderson, D.M.** (2020). Black soldier fly larvae meal as a protein source in low fish meal diets for Atlantic salmon (*Salmo salar*). Aqua., 521: 734978.

- **Fitches, E.C. and Smith, R.** (2018). Protein SECT: Insects as a Sustainable Source of Protein, Edible Insects in Sustainable Food Systems. Springer, pp. 421-433.
- **Fukuda, E.P.; Cox, J.R.; Wickersham, T.A. and Drewery, M.L.** (2022). Evaluation of Black Soldier Fly larvae (*Hermetia illucens*) as a protein supplement for beef steers consuming low-quality forage. Transl. Anim. Sci., 6, txac018.
- **Gunasekera, R.M.; Shim, K.F. and Lam, T.J.** (1996a). Effect of dietary protein level on spawning performance and amino acid composition of eggs of Nile tilapia, *Oreochromis niloticus*. Aqua., 146, 121-134.
- **Gunasekera, R.M.; Shim, K.F. and Lam, T.J.** (1996b). Influence of protein content of brood stock diets on larval quality and performance in Nile tilapia, *Oreochromis niloticus* (L). Aqua., 146, 245-259.
- Guerreiro, I.; Castro, C.; Antunes, B.; Coutinho, F.; Rangel, F.; Couto, A.; Serra, C.R.; Peres, H.; Pousão-Ferreira, P. and Matos, E. (2020). Catching black soldier fly for meager: Growth, whole-body fatty acid profile and metabolic responses. Aqua., 516: 734613.
- **Hardy, R.W.** (2010). Utilization of plant proteins in fish diets: effects of global demand and supplies of fishmeal. Aquacult, Res., 41: 770-776.
- Karlsen, Ø.; Amlund, H.; Berg, A. and Olsen, R.E. (2017). The effect of dietary chitin on growth and nutrient digestibility in farmed Atlantic cod, Atlantic salmon and Atlantic halibut. Aquacult. Res., 48: 123-133.
- **Kasumyan, A.O.** (2016). Taste attractiveness of free amino acids and their physiochemical and biological properties (as exemplified by fishes). J. Evol. Biochem. Physiol., 52: 271-281.
- Kishawy, A.T.Y.; Mohammed, H.A., Zaglool, A. W.; Attia, M.S.; Hassan, F.A.M., Roushdy, E.M.; Ismail, T. A. and Ibrahim, D. (2022). Partial defatted black solider larvae meal as a promising strategy to replace fish meal protein in diet for Nile tilapia (*Oreochromis niloticus*): Performance, expression of protein and fat transporters, and cytokines related genes and economic efficiency. Aqua., 555, 738195
- Kroeckel, S.; Harjes, A.G.; Roth, I.; Katz, H.; Wuertz, S.; Susenbeth, A. and Schulz, C. (2012). When a turbot catches a fly: Evaluation of a pre-pupae meal of the Black Soldier Fly (*Hermetia illucens*) as fish meal substitute: Growth performance and chitin degradation in juvenile turbot (*Psetta maxima*). Aqua., 364: 345-352.
- **Li, S.; Ji, H.; Zhang, B.; Zhou, J. and Yu, H.** (2017). Defatted black soldier fly (*Hermetia illucens*) larvae meal in diets for juvenile Jian carp (*Cyprinus carpio* var. Jian): Growth performance, antioxidant enzyme activities, digestive enzyme activities, intestine and hepatopancreas histological structure. Aqua., 477: 62-70.

- Li, Y.; Kortner, T.M.; Chikwati, E.M.; Belghit, I.; Lock, E.J. and Krogdahl, Å. (2020). Total replacement of fish meal with black soldier fly (*Hermetia illucens*) larvae meal does not compromise the gut health of Atlantic salmon (*Salmo salar*). Aqua., 520: 734967.
- Liland, N.S.; Biancarosa, I.; Araujo, P.; Biemans, D.; Bruckner, C.G.; Waagbø, R.; Torstensen, B.E. and Lock, E.J. (2017). Modulation of nutrient composition of black soldier fly (*Hermetia illucens*) larvae by feeding seaweed-enriched media. PLoS One, 12: e0183188.
- **Lim, C.E.** (2006). Tilapia–Biology, Culture, and Nutrition; Webster: Springfield, MA, USA, 2006.
- **Luna, M.; Llorente, I. and Cobo, A.** (2019). Integration of environmental sustainability and product quality criteria in the decision-making process for feeding strategies in sea bream aquaculture companies. J. Clean. Prod., 217, 691–701.
- MacRae, **J. and** Lobley G.E. (2003). Some factors which influence thermal energy losses during the metabolism of ruminants Quelques facteurs des pertes d'energie thermique liées au métabolisme des ruminants Einige Faktoren mit Einfluss auf die Wärmeenergieverluste während des Stoffwechsels bei Wiederkäuern. <u>Livestock Production Science Volume 9, Issue 4, July 1982, Pages 447-45 Accepted 16 December 1981, Available online 1 October 2003. https://doi.org/10.1016/0301-6226(82)90050-1</u>
- Mancini, S.; Medina, I.; Iaconisi, V.; Gai, F.; Basto, A. and Parisi, G. (2018). Impact of black soldier fly larvae meal on the chemical and nutritional characteristics of rainbow trout fillets. Animal, 12: 1672-1681.
- Mengistu, S.B.; Mulder, H.A.; Benzie, J.A.H. and Komen, H.A (2020). Systematic literature review of the major factors causing yield gap by affecting growth, feed conversion ratio and survival in Nile tilapia (*Oreochromis niloticus*). Rev. Aquac., 12, 524-541.
- Meric, I.; Wuertz, S.; Kloas, W.; Wibbelt, G.; Schulz, C. (2011). Cottonseed oilcake as a protein source in feeds for juvenile tilapia (*Oreochromis niloticus*). Antinutritional effects and potential detoxification by iron supplementation. Isr. J. Aquac., Bamidgeh,. 63: 568-576.
- Moretta, A.; Salvia, R.; Scieuzo, C.; Di Somma, A.; Vogel, H.; Pucci, P.; Sgambato; Wol, M. and Falabella, P. (2020). A bioinformatic study on antimicrobial peptides identified in the black soldier fly (BSF) *Hermetia illucens* (*Diptera: Stratiomyidae*). Sci. Rep., 10: 16875.

- Munguti, J.; Odame, H.; Kirimi, J.; Obiero, K.; Ogello, E. and Liti, D. (2021). Fish feeds and feed management practices in the Kenyan aquaculture sector: Challenges and opportunities. Aquat. Ecosyst. Health Manag., 24(1): 82-89.
- Newton, G.L.; Sheppard, D.C.; Watson, D.W.; Burtle, G.J.; Dove, C.R.; Tomberlin, J.K. and Thelen, E.E. (2005). The Black soldier fly, *Hermetia illucens*, as a manure management/resource recovery tool. In Proceedings of the Symposium on the State of the Science of Animal Manure and Waste Management, San Antonio, TX, USA, 5-7 January 2005.
- **Ng, W.K. and Romano, N.** (2013). A review of the nutrition and feeding management of farmed tilapia throughout the culture cycle. Rev. Aquac., 5, 220-254.
- **Nguyen, T.T.X.; Tomberlin, J.K. and Vanlaerhoven, S.** (2015). Ability of Black Soldier fly (*Diptera Stratiomyidae*) larvae to recycle food waste. Environ. Entomol., 44, 406-410.
- **NRC** (2011). National Research Council. Nutrient Requirement of Fish. National Academy Press, Washington, DC, USA.
- **Obirikorang, K.A.; Amisah, S.; Fialor, S.C. and Skov, P.V.** (2015). Local agroindustrial by-products with potential use in Ghanaian aquaculture: a review. Aquac. Int., 23: 403-425.
- Park, S.; Kim, J. and Yoe, S. (2015). Purification and characterization of a novel antibacterial peptide from black soldier fly (*Hermetia illucens*) larvae. Dev. Comp. Immunol., 52: 98-106.
- Prabu, E.; Rajagopalsamy, C.B.T.; Ahilan, B.; Jeevagan, I.J.M.A. and Renuhadevi, M. (2019). Tilapia—An excellent candidate species for world aquaculture: A review. Annu. Res. Rev. Biol., 31, 1-14.
- Rawski, M.; Mazurkiewicz, J.; Nczy, k B. K. and Józefiak, D. (2020). Black Soldier Fly Full-Fat Larvae Meal as an Alternative to Fish Meal and Fish Oil in Siberian Sturgeon Nutrition: The Effects on Physical Properties of the Feed, Animal Growth Performance, and Feed Acceptance and Utilization. Anim., 10: 2119.
- Rawski, M.; Mazurkiewicz, J.; Kiero'nczyk, B. and J'ozefiak, D. (2021). Black soldier Fly full-fat larvae meal is more profitable than fish meal and fish oil in Siberian sturgeon farming: the effects on aquaculture sustainability, economy and fish GIT development. Anim., 11, 604.
- Renna, M.; Schiavone, A.; Gai, F.; Dabbou, S.; Lussiana, C.; Malfatto, V.; Prearo, M.; Capucchio, M.T.; Biasato, I. and Biasibetti, E. (2017). Evaluation of the suitability of a partially defatted black soldier fly (*Hermetia illucens* L.) larvae meal as ingredient for rainbow trout (*Oncorhynchus mykiss* Walbaum) diets. J. Anim. Sci. Biotechnol., 8: 1-13.

- Reyes, M.; Rodríguez, M.; Montes, J.; Barroso, F.G.; Fabrikov, D.; Morote, E. and S'anchez- Muros, M.J. (2020). Nutritional and growth effect of insect meal inclusion on sea bass (*Dicentrarchuss labrax*) feeds. Fish., 5: 16.
- **Siddiqui, A.Q.; Al-Hafedh, Y.S. and Ali, S.A.** (1998). Effect of dietary protein level on the reproductive performance of Nile tilapia, *Oreochromis niloticus* (L.). Aquac. Res., 29, 349-358.
- Smetana, S.; Palanisamy, M.; Mathys, A. and Heinz, V. (2016). Sustainability of insect use for feed and food: Life Cycle Assessment perspective. J. Clean. Prod., 137, 741-751.
- **SPSS** (2020). Statistical Package for Social Science (**Software version:** 22.0).
- Stejskal, V., Tran, H.Q., Prokesova, M., Gebauer, T., Giang, P.T., Gai, F. and Gasco, L. (2020). Partially defatted *Hermetia illucens* larva meal in diet of eurasian perch (*Perca fluviatilis*) juveniles. Anim., 10, 1876.
- Su, J.; Gong, Y.; Cao, S.; Lu, F., Han, D.; Liu, H.; Jin, J.; Yang, Y.; Zhu, X. and Xie, S. (2017). Effects of dietary *Tenebrio molitor* meal on the growth performance, immune response and disease resistance of yellow catfish (*Pelteobagrus fulvidraco*). Fish Shellfish Immunol., 69: 59-66.
- **Suresh, A.V. and Lin, C.K. (1992)**. Effect of stocking density on water quality and production of Red tilapia in a recirculated water system. Aquac. Eng., 11, 1-22.
- **Sutthi, N. and Thaimuangphol W.** (2020). Effects of yeast (Saccharomyces cerevisiae) on growth performances, body composition and blood chemistry of Nile tilapia (*Oreochromis niloticus* Linnaeus, 1758) under different salinity conditions. Iran, J. Fish.Sci., 19 (3): 1428-1446 DOI: 10.22092/ijfs.2019.119254
- **Tacon, A.G. and Metian, M.** (2008). Global overview on the use of fish meal and fish oil in industrially compounded aqua feeds: trends and future prospects. Aqua., 285, 146-158.
- **Tippayadara, N.; Dawood, M.A.; Krutmuang, P.; Hoseinifar, S.H.; Doan, H.V. and Paolucci, M.** (2021). Replacement of fish meal by black soldier fly (*Hermetia illucens*) larvae meal: effects on growth, haematology, and skin mucus immunity of Nile Tilapia, *Oreochromis niloticus*. Anim., 11: 193.
- Tran, N.; Chu, L.; Chan, C.Y.; Peart, J.; Nasr-Allah, A.M. and Charo-Karisa, H. (2022). Prospects of fish supply-demand and its implications for food and nutrition security in Egypt. Mar. Policy, 146:105333.

- **Vo, V.** (2019). Development of Insect Production Automation: Automated Processes for the Production of Black Soldier Fly (*Hermetia illucens*). Master's Thesis, School of Electrical Engineering, Aalto University, Espoo, Finland.
- Wang, Y.S. and Shelomi, M. (2017). Review of Black Soldier Fly (*Hermetia illucens*) as Animal Feed and Human Food. Foods, 6, 91.
- Wang, G.; Peng, K.,; Hu, J.; Yi, C.; Chen, X.; Wu, H. and Huang, Y. (2019). Evaluation of defatted black soldier fly (*Hermetia illucens L.*) larvae meal as an alternative protein ingredient for juvenile Japanese sea bass (*Lateolabrax japonicus*) diets. Aqua., 507, 144-154.
- Wardani, W.W.; Alimuddin, A.; Zairin, M.; Setiawati, M.; Nuryati, S. and Suprayudi, M.A. (2021). Growth performance, robustness against stress, serum insulin, IGF-1 and GLUT4 gene expression of Red tilapia (*Oreochromis sp.*) fed diet containing graded levels of creatine. Aquac. Nutr., 27, 274-286.
- Williams, I.; Williams, K.; Smith, D. and Jones, M. (2006). Polka-dot grouper, Cromileptes altivelis, can utilize dietary fat efficiently. Aquac. Nutr., 12: 379-387.
- Yu, M.; Li, Z.M.; Chen, W.D.; Rong, T.; Wang, G. and Ma, X.Y. (2019a). *Hermetia illucens* larvae as a potential dietary protein source altered the microbiota and modulated mucosal immune status in the colon of finishing pigs. J. Anim. Sci. Biotechnol., 10, 1-16.
- Yu, M.; Li, Z.M.; Chen, W.D.; Rong, T.; Wang, G.; Li, J.H. and Ma, X.Y. (2019b). Use of *Hermetia illucens* larvae as a dietary protein source: Effects on growth performance, carcass traits, and meat quality in finishing pigs. Meat Sci., 158, 107837.
- **Zhang, J.B.; Tomberlin, J.K.; Cai, M.M.; Xiao, X.P.; Zheng, L.Y.; Yu, Z.N.** (2020). Research and industrialization of *Hermetia illucens* L. in China. J. Insects Food Feed, 6, 5-12.
- **Zhou, J.; Liu, S.; Ji, H. and Yu, H.** (2018). Effect of replacing dietary fish meal with black soldier fly larvae meal on growth and fatty acid composition of Jian carp (*Cyprinus carpio* var. Jian). Aquacult. Nutr., 24: 424-433.