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# Biochemical and Sensory Quality of the African Catfish (Clarias gariepinus) Spreads

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# ABSTRACT

The African catfish (Clarias gariepinus) is known for its resilience in extreme conditions; however, it is not widely accepted by most Egyptian consumers. This study aimed to determine the biochemical and sensory quality of fish spread made from the catfish meat, with varying levels of potato substitution. Five treatments were designed: a control (without potatoes) and four treatments with 5, 10, 15, and 20% potatoes. The results showed that the control spread contained 63.24% moisture, 16.05% crude protein, 16.60% fat, 2.25% ash, and 1.68% carbohydrates. Additionally, the quality indices recorded were pH 6.51, TVN 13.02mg/ 100g, TBA 0.23mg MDA/kg sample, and TPC 43×10<sup>2</sup> cfu/g. These values varied with each treatment, showing significant changes in both protein and carbohydrate content. Regarding sensory quality, the fish spread with 10% potatoes received a high score for color, while the treatment with 15% potatoes achieved high scores in color, spreadability, aftertaste, and overall acceptability. In conclusion, spreads made from the catfish were well accepted by panelists, and the inclusion of potatoes up to 15% improved both the biochemical and sensory quality. This study recommends that underutilized catfish be considered a good source for creating suitable fish spreads for all ages.

# **INTRODUCTION**

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Aquaculture is growing rapidly to cover the high protein demand for 9 billion human populations by 2050 (Gebremichael *et al.*, 2023). Factors of increasing in population growth, disposable incomes, working women and relative preferences are encouraging to the demand for ready to-eat fishery products (Venugopal, 2006; Bochi *et al.*, 2008; Kilinc *et al.*, 2008). The demand for fishery products as modern life style has been increased due to convenience, nutritional adequacy and good taste. However, spreads are limited and expensive (Mohanty *et al.*, 2019; Kakatkar *et al.*, 2022).

Although fish are a cherished delicacy that can help overcome socio-economic, age, religious, and educational barriers (Adebayo-Tayo *et al.*, 2008), they are also extremely susceptible to microbial contamination (Abolagba & Uwagbai, 2011).

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The catfish has gained a wide acceptability due to its unique sensory properties and being a good source of nutritive value required for human (**Abdullahi** *et al.*, **2001**). Fish spreads have good quality and nutritional value in addition to their convenience and shelf life. Moreover, they are regularly consumed and are made from underutilized fish. However, most fish spreads do not contain functional lipid or micronutrients (**Bhattacharya** *et al.*, **2012; Rathod** *et al.*, **2018**). Recently, **Kakatkar** *et al.* (**2022**) studied the shelf life of fish spread processed from Bombay duck (*Harpadon nehereus*) and preserved using gamma rays (5 kGy). They concluded that the developed fish spread was suitable for all consumers. **Mostafa** *et al.* (**2023**) reported that the protein content decreased by adding feta cheese and ketchup to produce the Indian mackerel and pangasius spreads while carbohydrates, fat and nutritional value increased.

In Egypt, freshwater African catfish (*Clarias gariepinus*) exhibits successful growth under extreme conditions due to its strong resistance to poor water quality. However, it is not widely accepted by most consumers, despite its nutritional value being comparable to that of other species such as tilapia. Therefore, this study was designed to determine the biochemical and sensory quality of fish spreads made from the African catfish (*Clarias gariepinus*) with varying levels of potato substitution (0, 5, 10, 15, and 20%).

### MATERIALS AND METHODS

### Materials

About 15kg of live catfish (Clarias gariepinus) samples (Fig. 1A) were purchased from the Elkanater Elkaairia market in December 2023. The average weight and length recorded were  $1.177 \pm 5.74$ kg and  $51.92 \pm 8.40$ cm, respectively. The fish were immediately transferred to the Fish Technology and Processing Laboratory at the National Institute of Oceanography and Fisheries (NIOF), Egypt.

All spices and other additives (potatoes, corn flour, butter, sunflower oil, onion, garlic) were purchased from the local market (Table 1). Food grade-multilayers tubes with screw cover (capacity 50g) were carefully sterilized using heated water and drained. **Technological processes** 

Catfish samples were manually filleted, washed, soaked in the acid-brine (10% sodium chloride and 0.02% acetic acid) for 10min at 22°C, rinsed, drained and minced (3mm) using electrical meat mixer. Fish mince was cooked by autoclave at 116°C for 20 minutes. Moreover, powder spices were well mixed and sautéed with sunflower oil by heating. Potatoes were carefully washed, cut into rings shape, put in pot with enough water and boiled for 30 minutes. Additionally, cooked potatoes were drained, gently minced and shells were discarded.

Item	%	*Spices mix	g	%				
Cooked fish mince	65.00	White pepper	0.84	42				
<b>Boiled potatoes as Replacer</b>	0, 5, 10, 15, and 20%	Cumin	0.52	26				
Corn flour	2.44	Cardamom	0.04	2				
Butter	10.00	Cubeb	0.06	3				
Sunflower oil	10.00	Clove	0.04	2				
Carrot	5.40	Red pepper	0.22	11				
Fresh garlic	1.00	Coriander	0.20	10				
Fresh onion	2.00	Ginger	0.04	2				
Sod. chloride	1.50	Cinnamon	0.02	1				
Polyphosphate	0.40	Turmeric	0.02	1				
Sod. carbonate	0.26		1	1				
*Spices mix	2.00							

 Table 1. The recipe used in this work

# Treatments

Five treatments were designed: a control (without potatoes) and treatments with 5,

10, 15, and 20% potato levels replacing fish mince, as shown in Table (2).

# **Preparation of fish spreads**

Cooked fish mince was gently homogenized with sautéed spices and potatoes paste and was then filled in tubes (Fig. 1B). All treatments were autoclaved at 110°C for 10min, tightly closed, cooled and analyzed.



Fig. 1. (A) Raw catfish; (B) Its spreads

Itom	Treatments					
Item	Control	5%	10%	15%	20%	
Cooked fish mince (65%)	650	617.5	585.0	552.5	520.0	
Boiled potatoes	-	32.5	65.0	97.5	130.0	
Additives (35%)						
Butter	100	100	100	100	100	
Sunflower oil	100	100	100	100	100	
Carrot	54	54	54	54	54	
Corn flour	24.4	24.4	24.4	24.4	24.4	
Spices mix	20	20	20	20	20	
Onion	20	20	20	20	20	
Garlic	10	10	10	10	10	
Sod. chloride	15	15	15	15	15	
Sod. bicarbonate	4	4	4	4	4	
Polyphosphate	2.6	2.6	2.6	2.6	2.6	

Table 2. The recipes of fish spreads (kg) used in this study

# **Analytical methods**

Moisture, crude protein (calculated as TN  $\times$  6.25), fat, and ash content of all treatments were determined according to the methods of **AOAC** (2000). Salt content, expressed as sodium chloride, was measured following the method of **Mol** *et al.* (2010). Total carbohydrate content was calculated by difference, as described by **Maclean** *et al.* (2003). The energetic value (kcal/100 g) of the cooked samples was calculated using the formula:

Energetic value =  $[(\% \text{ of carbohydrate } \times 4) + (\% \text{ of protein } \times 4) + (\% \text{ of fat } \times 9)]$ 

The pH value of the filtrate was measured using a digital pH meter (Adwa, AD 131) according to the method outlined by **Egbert** *et al.* (1992). The TBA value was assessed using the method of **Tarladgis** *et al.* (1960), while the total volatile basic nitrogen (TVB-N) and trimethylamine nitrogen (TMA-N) content were determined following the method of AMC (1979). Additionally, total plate count (TPC) was examined according to APHA (1992).

Sensory tests (including appearance, texture, odor, spreadability, taste, aftertaste, and total acceptability) were evaluated using a 9-point hedonic scale (where 9 = like extremely, 8 = like very much, 7 = like moderately, 6 = like slightly, 5 = neither liked nor disliked, 4 = dislike slightly, 3 = dislike moderately, 2 = dislike very much, and 1 = dislike extremely) for catfish spreads (Muzaddadi, 2013). The results obtained were statistically analyzed ( $P \le 0.05$ ) and expressed as mean ± SD using SPSS (Version 16).

# RESULTS

#### Nutritional composition of catfish spreads

The nutritional composition (wet weight) of different catfish spreads is shown in Table (3). The control spread contained moisture at 63.24%; crude protein at 16.05%; fat at 16.60%; ash at 2.25%; and salt at 1.41%, and carbohydrates at 1.68%. With regard to the effect of adding potatoes, the average moisture content ranged from 62.20 to 63.81%; crude protein from 10.88 to 13.66%; fat from 16.11 to 16.49%, ash from 2.12 to 2.34%; salt from 1.52 to 1.76%; and carbohydrates from 5.61 to 7.07% across all treatments containing 5, 10, 15, and 20% potatoes. Additionally, the total energy of the treatments ranged from 219.52 to 222.79kcal/ 100g compared to the control spread at 221.04kcal/ 100g.

Constituent (%)	Control	Catfish spreads with potatoes				
Constituent (70)	Control	5%	10%	15%	20%	
Moisture	63.24±0.34	62.20±0.53	63.81±0.02	$62.94 \pm 0.02$	63.27±0.07	
Crude protein	$16.05 \pm 0.93$	13.66±0.96	12.41±0.92	$12.10\pm0.14$	$10.88 \pm 0.00$	
Fat	$16.60 \pm 3.30$	$16.19 \pm 0.00$	16.16±6.31	16.11±1.49	16.49±0.39	
Ash	$2.25 \pm 0.66$	$2.34{\pm}1.50$	$2.12 \pm 0.01$	$2.12\pm0.06$	$2.29 \pm 0.08$	
Salt (sod. chloride)	$1.41\pm0.33$	$1.76\pm0.16$	$1.64 \pm 0.33$	$1.52\pm0.42$	$1.76 \pm 0.66$	
Carbohydrates	1.86	5.61	6.11	6.73	7.07	
Energetic value (kcal/100g)	221.04	222.79	219.52	220.31	220.21	

**Table 3.** Nutritional composition of the catfish spreads products

These results were expressed as mean  $\pm$  SD.

## **Quality criteria**

Table (4) demonstrates some quality indices of the catfish spread products. The control fish spread had a pH of 6.51, total volatile nitrogen (TVN) of 13.02mg/ 100g, trimethylamine (TMA) of 6.94mg/ 100g, and free amino nitrogen (FAN) of 1.88mg/ 100 g. Additionally, it had an acid value (AV) of 0.38mg/ 100g, a TBA value of 0.23mg MDA/ kg sample, while the total plate count (TPC) was  $43 \times 10^2$  cfu/g sample. These values varied across the other treatments, depending on the different levels of potatoes.

## Sensory evaluation

Sensory tests of the catfish spreads products are tabulated in Table (5). The sensory properties were carried out the first day after fish spreads processing by trained panelists. Control fish spread had got slightly scores of appearance, color, flavor, taste, after taste and overall acceptability, while it had got a like moderately of spreadability test. Both treatments containing 5 and 20% potatoes received similar scores for sensory properties. The treatment with 10% potatoes achieved a high score (very much) for color,

while the treatment with 15% potatoes scored highly in color, spreadability, aftertaste, and overall acceptability.

Index	Control	Catfish spreads with potatoes levels				
		5%	10%	15%	20%	
pH value	6.51±0.03	6.24±0.07	6.39±0.01	6.65±0.01	6.45±0.01	
<sup>1</sup> TVBN (mg/100g)	13.02±0.00	$10.42 \pm 0.00$	7.81±0.00	10.42±0.00	7.81±0.00	
2TMA (mg/100g)	6.94±0.00	7.81±0.00	6.08±0.00	5.21±0.00	$6.94 \pm 0.00$	
3FAN (mg/100g)	$1.88 \pm 0.00$	2.23±0.06	2.18±0.28	2.15±0.02	$2.52 \pm 0.00$	
<sup>4</sup> AV (mg/100g)	0.38±0.04	$0.46 \pm 0.01$	0.41±0.07	0.31±0.01	$0.48 \pm 0.08$	
5TBARS (mg MDA/	0.23±0.02	$0.70 \pm 0.08$	0.25±0.08	0.38±0.02	0.34±0.06	
kg)						
<sup>6</sup> TPC (cfu/g)	$43 \pm 4.24 \times 10^{2}$	61±9.90×10 <sup>2</sup>	25±1.41×10 <sup>2</sup>	$56\pm2.82\times10^{2}$	52±1.06×10 <sup>2</sup>	

Table 4. Some quality indices of raw catfish spreads

These results were expressed as mean  $\pm$  SD. <sup>1</sup>TVN: total volatile basic nitrogen; <sup>2</sup>TMA: trimethylamine nitrogen; <sup>3</sup>FAN: free amino nitrogen; <sup>4</sup>AV: acid value; <sup>5</sup>TBARS: thiobarbituric acid reactive substances; <sup>6</sup>TPC: total plate count.

Test	Control	Catfish spreads with potatoes levels				
		5%	10%	15%	20%	
Appearance	6.94±1.27	7.81±1.00	7.94±0.77	7.94±0.56	7.81±0.96	
Color	6.81±1.31	7.56±1.29	8.00±1.10	$8.25 \pm 0.76$	$7.69\pm0.88$	
Flavor	6.31±1.33	7.63±0.69	$7.50{\pm}1.20$	$7.69 \pm 1.10$	7.50±0.71	
Spreadability	$7.06 \pm 1.32$	7.50±0.89	7.81±0.84	8.31±0.53	7.44±0.90	
Taste	6.44±1.35	7.19±1.13	7.75±1.16	$7.63 \pm 1.30$	7.75±0.46	
After taste	6.75±0.89	7.38±1.19	7.56±1.45	$8.00 \pm 0.96$	7.75±0.46	
Overall	6.75±1.49	7.63±1.30	7.94±0.86	8.13±0.88	7.81±0.65	
acceptability						

**Table 5.** Sensory evaluation of the catfish spreads

9- like extremely; 8-like very much; 7-like moderately; 6-like slightly; 5-neither liked nor dislike; 4-dislike slightly; 3-dislike moderately; 2-dislike very much; 1-dislike extremely.

## DISCUSSION

In this work, the results align with other studies that indicate the nutritional requirements of ready-to-eat fish products, which are due to their content of moisture, proteins, lipids, minerals, and vitamins. These characteristics have contributed to the increasing demand among consumers (Pagarkar *et al.*, 2011; VKM, 2014; Nikmaram *et al.*, 2018; Emam *et al.*, 2016, 2022).

The moisture content of catfish spreads (Table 3) is consistent with the findings of **Freitas** *et al.* (2012), who reported 62.17% for tilapia spread, as well as final spread products from **Khater and Farag** (2016) and mackerel and pangasius spreads ranging from 54.10 to 63.50% (Mostafa *et al.*, 2023). This moisture content is higher than the 56.12 to 58.12% found in tilapia spreads (Minozzo *et al.*, 2008).

Regarding protein content, all samples except the control (Table 3) were comparable to the 12.75% reported for fish spread by **Kakatkar** *et al.* (2022). This protein level is higher than the 8.77% to 9.69% found in tilapia spreads (**Minozzo** *et al.*,

**2004**), 8.53% (**Minozzo** *et al.*, **2008**), 9.75% (**Freitas** *et al.*, **2012**), and 9.75% (**Khater & Farag, 2016**), but lower than the 19.8% to 21.50% reported by **Mostafa** *et al.* (**2023**).

The results of fat content (Table 3) are lower than 27.41% (Minozzo et al., 2004), 26.12-28.15% (Minozzo et al., 2008), 18.81% (Freitas et al., 2012; Khater & Farag, 2016), and higher than 3.53% (Kakatkar et al., 2022) and 4.10-13.30% (Mostafa et al., 2023). The ash content in this study aligns with the finding of Minozzo et al. (2004), who reported a value of 2.20%, and corroborates with the value of 2.11%, recorded in the studies of Freitas et al. (2012) and Khater and Farag (2016). While, the current result is higher than 1.51%, noted by Kakatkar et al. (2022). However, it is lower than the range of 3.01 to 3.26% reported by Minozzo et al. (2008) and that (4.00 to 4.30%) detected by Mostafa et al. (2023). Additionally, both carbohydrate content and total energy varied depending on the recipes used.

The pH values reported in this study are slightly higher than those found in mackerel (6.15) and pangasius (6.21) spreads (Mostafa *et al.*, 2023). Moreover, it is greater than the values for the salmon (6.09), herring (5.78), and anchovy pastes (5.64) (Khater & Farag, 2016). On the other hand, the total volatile nitrogen (TVN) content for the control treatment is consistent with Khater and Farag (2016), who reported TVN values of 13.80, 13.92, and 14.36mg/ 100g for herring, salmon, and anchovy pastes, respectively.

For the other treatments, while TVN values concur with previous studies, trimethylamine (TMA) content varied; **Kakatkar** *et al.* (2022) found TMA values of 3.51mg/ 100g. In contrast, our results showed a notable decrease in TVN compared to the higher values of 32.08 and 21.67mg/ 100g for the mackerel and pangasius spreads, respectively (Mostafa *et al.*, 2023). Overall, the TVN and TMA values in this study remain within the acceptable limits established by Connell (1990), which are 35 and 15mg/ 100g, respectively.

The thiobarbituric acid (TBA) values (Table 4) are lower than those reported for anchovy, salmon, and herring pastes (1.73, 2.26, and 1.30mg MDA/kg, respectively) (**Khater & Farag, 2016**); whereas, they are lower than the 1.67mg MDA/kg found by **Kakatkar** *et al.* (2022) and the values for mackerel (2.03mg MDA/kg) and pangasius spreads (2.67mg MDA/kg) (**Mostafa** *et al.*, 2023).

The total plate count (TPC) in this study (Table 4) is lower than the values reported for salmon, herring, and anchovy pastes (5.34, 5.35, and 5.53 log cfu/g, respectively) (**Khater & Farag, 2016**) and 2.42 log cfu/g from **Kakatkar** *et al.* (2022). However, it is higher than the TPC values for mackerel and pangasius spreads, which ranged from  $1.8 \times 102$  to  $3.1 \times 102$  and  $8.1 \times 103$  to  $9.7 \times 103$  cfu/g, respectively (**Mostafa** *et al.*, 2023). This indicates a relatively good microbial quality for the catfish spreads developed in this study.

Regarding sensory properties, the catfish spreads (Table 5) did not exhibit any fishy odor, though a strong fishy taste was detected during sensory analysis. Despite this, all

fish spread products were accepted, aligning with the findings of Aquerreta *et al.* (2002). The absence of fishy odor in our study can be attributed to the pre-soaking of fish fillets in acid-brine, the addition of spices, and the deodorization achieved through autoclaving during cooking. This is consistent with the outcomes of **Bhattacharya** *et al.* (2012), who noted that autoclaving effectively minimized strong fishy odors. Additionally, **Freitas** *et al.* (2012) found that spreads seasoned with salt were preferred by the largest number of panelists.

Our results agree with those of **Mostafa** *et al.* (2023), who reported that sensory scores for mackerel and pangasius spreads ranged from very good to good. Moreover, **Kakatkar** *et al.* (2022) observed a correlation between sensory scores and microbiological quality, reinforcing the importance of these factors in consumer acceptance.

Overall, the variations between our results (Table 3) and those of previous studies can be attributed to differences in fish species, types and quantities of ingredients used, as well as processing conditions. These factors play a significant role in determining the sensory qualities of fish products.

### CONCLUSION

The catfish is often considered unappealing to many consumers in Egypt, likely due to its appearance, feeding behavior, and associated sounds. This study aimed to develop fish spreads that leverage the nutritional benefits of the catfish while addressing these disadvantages. The findings revealed that both the raw catfish and its spreads possess high nutritional value and quality indices. Furthermore, the incorporation of potatoes significantly enhanced the biochemical, safety, and sensory qualities of the catfish spreads.

In conclusion, this work recommends utilizing the catfish to create delicious products viz. spreads that can appeal to consumers of all ages. By promoting these spreads, it may be possible to improve the acceptance of the catfish in the market and expand its culinary applications.

#### REFERENCES

Abdullahi, S. A.; Abolude, D. S. and Ega, R. A. (2001). Nutrient quality of four oven dried freshwater catfish in Northern Nigeria. J. Trop. Biosci., pp: 70.

- Abolagba, O.J. and Uwagbai, E.C. (2011). A comparative analysis of the microbial load of smoke-dried fishes (*Ethmalosa fimbriata* and *Pseudotolithus elongatus*) sold in Oba and Koko markets in Edo and Delta States, Nigeria at different seasons. Australian Journal of Basic and Applied Sciences, 5 (5): 544-550.
- Adebayo-Tayo, B.C.; Odu, N.N; Igiwiloh, N.J.P.N. and Okonko, I.O. (2012). Microbiological and physicochemical level of fresh catfish (Carius hendelotic)

from different markets in Akwa Ibom state, Nigeria. New York Science J., 5 (4):46-52.

- AMC, (1979). Alteration Methods of Chemistry for the Examination of Fish and Fish Products. Analyst: 104-433.
- AOAC (2000). Official Methods of Analysis of the Association of Analytical Chemists. 18<sup>th</sup> Ed., Washington, D.C.
- **APHA** (1992). American Publish Health Association Compendium of Methods for the Microbiological Examination of Foods. Washington D.C, USA.
- Aquerreta, Y.; Astiasarana, I.; Mohino, A.; Bello, J. (2002). Composition of pâté elaborated with mackerel flesh (*Scomber scombrus*) and tuna liver (*Thunnus thynnus*): comparison with commercial fish pâté. Food Chemistry, Oxford, vol. 77 (2): 147-153. C.F. Freitas *et al.*, (2012). http://dx.doi.org/10.1016/S0308-8146(01)00310-7.
- Arvanitoyannis, I.S.; Tsitsika, E.V. and Panagiotaki, P. (2005). Implementation of quality control methods (physicochemical, microbiological and sensory) in conjunction with multivariate analysis towards fish authenticity. International Journal of Food Science and Technology, 40: 237-263. doi:10.1111/j.1365-2621.2004.00917.x
- Bhattacharya, T.; Bhowal, J.; Ghosh, M.; Bhattacharyya, DK. (2012). Studies on preparation of functional lipid and micronutrient enriched Bhola Bhetki fish (*Nibea soldado*) spread. 1:310. doi:10.4172/scientificreports.310
- Bochi, V.C.; Weber, J.; Ribeiro, C.P.; Victorio, A.M. and Emanuelli, T. (2008). Fish burgers with silver catfish (*Rhamdia quelen*) filleting residue. Biosource Technology, 99: 8844- 8849.
- **Connell, J.J.** (1990). Methods of assessing and selecting for quality. Control of fish quality, (3rd edn), Fishing News Books, Oxford, UK, pp. 122-150.
- Egbert, W. R.; Huffman, D. L.; Chen, C. M. and Jones, W. R. (1992). Microbial and oxidative changes in low-fat ground beef during simulated retail distribution. J. Food Sci., 57: 1269-1269.
- Emam, O. A.; Ibrahim, S. M. and Saber, M. B. (2016). Effect of recipes ingredients on carp burgers quality. J. Food and Dairy Sci., Mansoura Univ., vol. 7 (10): 451 – 455.
- Emam, O.A.; Ibrahim, S.M.; El-Basiouny, M. Ghada. and Saber, M.B. (2022). Determination of nutritional value of snacks containing grass carp powder. SJSE., Issue (20):626-652.
- Freitas, D. G. C.; Resende, A. L. S. S.; Furtado, A. A. L.; Tashima, L. and Bechara, H. M. (2012). The sensory acceptability of a tilapia (*Oreochromis niloticus*) mechanically separated meat-based spread Braz. J. Food Technol, Campinas, vol. 15 (2): 166-173.

- Gebremichael, A.; Szabó, A.; Sándor, Z.J.; Nagy, Z.; Ali, O. and Kucska, B. (2023). Chemical and physical properties of African Catfish (*Clarias gariepinus*) fillet following prolonged feeding with insect meal-based diets. Aquaculture Nutrition, vol. 2023, Issue 1 Article ID 6080387, 9 p. https://doi.org/10.1155/2023/6080387
- Kakatkar, A. S.; Gautam, R. K.; Mishra, P. K.; Kumar, V.; Debbarma A. and Chatterjee, S. (2022). Development of nutritious ready to eat fish spread from *Harpodon Nehereus* and its shelf-life extension using Gamma irradiation. J. Biotech Biores. 3(5). JBB. 000574.
- Khater, D. F. and Farag, S. E. S. (2016). Evaluation of bacterial and chemical quality of new manufactured pasted fish products in a large scale fish processing plant, Egypt. Benha Vet. Med. J., 31(2):63-72. doi:10.21608/bvmj.2016.31263
- Kilinc B.; Cakli, S. and Tolasa, S. (2008). Quality changes of sardine (*Sardine pilchardus*) patties during refrigerated storage. J. Food Quality, **31**: 366–381.
- Maclean, W., Harnly, J., Chen, J., Chevassus-Agnes, S., Gilani, G., Livesey, G., and Warwick, P. (2003). Food energy- methods of analysis and conversion factors. Food and Agriculture Organization of the United Nations Technical Workshop Report (vol. 77).
- Minozzo, M. G.; Waszczynskyj, N. and Beirão, L. H. (2004). Características físicoquímicas do patê de Tilápia do nilo (*Oreochromis niloticus*), comparado a produtos similares comerciais. Alimentos e Nutrição, Araraquara, vol., 15 (2): 101-105. 2004. C.F. Freitas *et al.*, (2012).
- Minozzo, M. G.; Waszczynskyj, N. and Boscolo, W. R. (2008). Utilização de carne mecanicamente separada de Tilápia (*Oreochromis niloticus*) para a produção de patês cremoso e pastoso. Alimentos e Nutrição, Araraquara, vol., 19 (3): 315-319. C.F. Freitas *et al.*, (2012).
- Mohanty, B. P.; Mahanty, A.; Ganguly, S.; Mitra, T.; Karunakaran, D. and Anandan, R. (2019). Nutritional composition of food fishes and their importance in providing food and nutritional security. Food Chemistry, 293: 561-570.
- Mol, S.; Cosansu, S.; Alakavuk, D. U. and Ozturanm S. (2010). Survival of Salmonella enteritidis during salting and drying of horse mackerel (*Trachurus* trachurus) fillets. International Journal of Food Microbiology, 13: 36–40.
- Mostafa, M. M.; Youssef, Kh. M.; Abouzied, A.S. and Amin, H. F. (2023). Quality changes during frozen storage of hot smoked fillets and spreads processed from Indian Mackerel (*Rastrelliger kanagurta*) and Pangasius (*Pangasius hypophthalmus*) fish. EJABF, vol. 27(1): 47 67.
- Muzaddadi, A. U. (2013). Naturally evolved fermented fish products of Northeast India (Seedal and Shidal)-A comparative study. Indian Journal of Natural Products and Resources, 4 (2): 170-177.

- Nikmaram, N.; Budaraju, S.; Barba, F.J.; Lorenzo, J.M. an Cox R.B.; *et al.* (2018). Application of plant extracts to improve the shelf-life, nutritional and healthrelated properties of ready-to-eat meat products. Meat Sci., 145: 245-255.
- Pagarkar, A.U.; Joshi, V.R.; Baug, T.E. and Kedar, J.G. (2011). Value addition is need of seafood industries. Fish Coops, 23: 8-17.
- Pegg, R.B. (2004). Lipids. Lipid oxidation/stability. In Handbook of Food Analytical Chemistry (R.E. Wrolstad, T.E. Acree, E.A. Decker, M.H. Penner, D.S. Reid, S.J. Schwartz, C.F. Shoemaker, D.M. Smith and P. Sporns, eds.) pp. 513–564, John Wiley & Sons, Inc., Hoboken, NJ.
- Rathod, N. B.; Pagarkar, A. U.; Pujari1, K. H.; Shingare, P. E.; Satam, S. B.; Phadke, G. G. and Gaikwad, B. V. (2018). Status of valuable components from Pangasius: A review. International Journal of Current Microbiology and Applied Sciences, 7(4): 2106-2120. doi: 10.20546/ijcmas.2018.704.241.
- Tarladgis, B. G.; Watts, B. M.; Younathan, M. T. and Dugan, Jr. L. (1960). A distillation method for the quantitative determination of Malonaldehyde in rancid foods. Journal of Am. Oil Chem. Soc., 37: 44-48.
- Venugopal, V. (2006). Mince and mince-based products. In V. Venugopal (ed.), seafood processing, adding value through quick freezing, Retort able Packaging, and cookchilling (pp.215-258). Boca Raton, FL: Tayler & Francis.
- VKM (Vitenskapskomiteen for matteygghet: Norwegian scientific committee for food safety) (2014). Benefit-risk assessment of fish and fish products in the Norwegian diet – an update. Scientific Opinion of the Scientific Steering Committee. VKM Report 15 [293 pp], Oslo, Norway. Available online: www.vkm.no.