Egyptian Journal of Aquatic Biology & Fisheries Zoology Department, Faculty of Science, Ain Shams University, Cairo, Egypt. ISSN 1110 – 6131 Vol. 28(4): 1479 – 1494 (2024) www.ejabf.journals.ekb.eg



Influence of Different Temperature-Salinity Combinations on the Oxygen Consumption in the Juveniles Jinga Shrimp *Metapenaeus affinis*

Ashwaq T. Abaas¹, Abdul-Hussein H. Ghazi^{1*}, Anteesar N. Sultan²

¹Department of Natural Marine Science, Marine Science College, University of Basrah, Iraq ²Department of Fishes and Marine Resources, University of Basrah, Agriculture College *Corresponding Author: <u>gabdulhussein@gmail.com</u>

ARTICLE INFO

Article History: Received: July 18, 2024 Accepted: Aug. 2, 2024 Online: Aug. 16, 2024

Keywords: Oxygen consumption, Temperature-salinity combinations, *Metapenaeus affinis*

ABSTRACT

The Shatt al-Arab River in Iraq has seen a significant spread and subsequent disappearance of certain shrimp species, potentially due to historical fluctuations in temperature and salinity. This study examines the effects of combined temperature and salinity on the oxygen consumption, metabolic rate, and thermal coefficient of juvenile Metapenaeus affinis. Salinity tolerance tests revealed that shrimp survived transfers up to 15ppt but experienced over 50% mortality at 20 ppt, with the highest survival (100%) at 5ppt and the lowest (17%) at 20ppt. Oxygen consumption increased with salinity at 20-21°C, with peak rates at 15 ppt (0.036mg O₂/h) and lowest at 1ppt (0.006mg O₂/h). At 25-26°C, oxygen consumption was higher at 10 and 15 ppt (0.040mg O₂/h) and lower at 5 and 10 ppt (0.010mg O₂/h). At 29-30°C, the highest consumption rate was 0.046mg O₂/h at 1ppt, while the lowest was 0.016 mg O₂/h at the same salinity in the second and third hours. Temperature significantly affected oxygen consumption, with the highest rate (0.018mg O₂/h) at 29-30°C and 15ppt, and the lowest rate (0.006mg O₂/h) at 20-21°C and 1 ppt. Metabolic rates peaked at 0.072 mg O₂/g at 29-30°C and 15 ppt and were lowest at 0.024mg O₂/g at 20-21°C and 1 ppt. These findings provide valuable insights for aquaculturists in developing practices to enhance growth and survival in varying environmental conditions.

INTRODUCTION

Scopus

Indexed in

In the 1980s and 1990s a number of shrimp species spread in the Iraqi local environment, especially in the Shatt al- Arab River, such as *Metapenaeus affinis* (Miquel, 1983), *Atyphaera desmarestii mosopotamica* (Al-Adhub, 1987), *Caridina Babulti basrensis* (Al-Adhub & Hamzah, 1987) and *Exopalaemon styliferus* (Salman & Bishop, 1990). After the 2000s, these species disappeared and other species were recorded as invasive species to the local environment like *Macrobrachium nipponense* (Salman *et al.*, 2006), *Macrobrachium lar* (Ghazi & Hassan, 2021), *Macrobrachium equidens* (Hassan *et al.*, 2023). There are various reasons for this situation, including climate changes especially temperature, pollution, and different levels of salinity in the

ELSEVIER DO

IUCAT

Shatt al-Arab due to the lack of drainage that negatively affect the marine environment in the region (Habashy & Hassan, 2011; Al-Mahmood *et al.*, 2015). One of the threatened species is the jinga shrimp, *Metapenaeus affinis* (H. Milne Edwards, 1837), it is dispersal from the Arabian Sea to South India, and live at depths between 55 to 90m, mainly on mud bottom (Fischer & Bianchi, 1984). The jinga shrimp is one of the dominant and highly valued penaeid shrimps along the coastal waters of Iraqi, and juveniles can travel for long distances from the marine toward the estuary of the Shatt al-Arab (inland waters), moreover the jinga shrimp is considered as a commercial shrimp in this area (Salman *et al.*, 1986; Salman *et al.*, 1990; Abbas & Ghazi, 2021).

Dissolved oxygen (DO) is the major factor for studying water quality since all aquatic organisms need oxygen to survive and grow (Anongponyoskun et al., 2012). The life cycle of *M. affinis* is completed between inland and marine waters. Therefore, this species is considered migratory between brackish water (1- 2ppt) and marine water (30ppt) (Gerami et al., 2012). Moreover, salinity and temperature are important factors and have influence on oxygen consumption (Duan et al., 2022). Knowledge of the effects of different salinity levels on dissolved oxygen will benefit aquaculture by promoting optimal growth and survival (Schuler, 2008; Cobo et al., 2014). Many papers have been written on the combined effects of salinity and temperature on oxygen consumption, metabolic rate, and thermal coefficient (Q10) (Stern et al., 1984; Rosas et al., 2001; Manush et al., 2004; Spanopoulos et al., 2005; Allan et al., 2006; González et al., 2010; Garcia-Guerrero et al., 2013; Vinagre et al., 2015; Rahi et al., 2021; García-Guerrero et al., 2022). The purpose of this research was to determine the effects of different salinity concentrations and temperatures on the oxygen consumption rate in the juveniles of Metapenaeus affinis shrimp, as well as assessing their metabolic rate and thermal coefficient (Q10).

MATERIALS AND METHODS

A total of 60 juvenile shrimps, *M. affinis*, were collected from the al-Mas'hab area, southern sector of Iraqi inland water near Al- Hammar marshes; this is located within an area that lies between 30° 39′ 34.27 " N; 47° 39′ 13.81" E. (Map 1). The shrimp were captured by trawl net (Fig. 1), and they were kept in laboratory aquaria containing water adjustment to the salinity level of resource water, and acclimated for 12 hours at 4ppt prior to the experiment. Additionally, the salinity of the resource water varied between 3 to 4ppt. and specimens weighed from 1.0 to 1.3g. The effect of a rapid change of salinity on the oxygen consumption was estimated every one hour and for four consecutive hours, this is a time when juveniles reach the point of stress due to lack of oxygen to less than 4mg/ l (initial DO 8.5mg/ l). The oxygen consumption was measured with a YSI oximeter, salinity by refractometer and controlled on temperature by a thermostat heater, during this time shrimp were not fed. We used four plastic tanks,

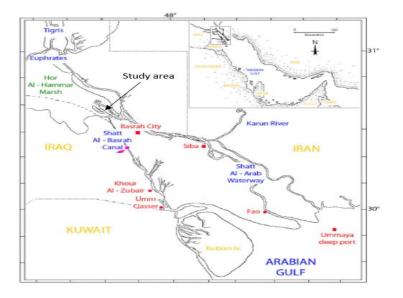
tightly closed to prevent the exchange of external air, the size of one tank is two liters, each tank contains three individuals of shrimp, and tanks were kept in the dark (Fig. 2). The juveniles were transferred from the acclimatization tanks directly to the experimental tanks (rapid change) containing different salinities by adding sea salts (1, 5, 10 and 15ppt). The temperatures were controlled within the specified ranges using an electric heater, and it was noted that the temperature ranges were set at $\pm 1^{\circ}$ C. Therefore, the chosen ranges were 20- 21, 25- 26, and 29- 30°C. Total oxygen consumption (OCT) (mg O₂/h /per shrimp) was calculated as: OCT = (Ct₀ – Ct₁) × (Cv).

Where, Ct_0 is the initial oxygen concentration (mg/L); Ct_1 is the final oxygen concentration, and Cv is water volume in liters.

Oxygen consumption per gram of weight (OCg) was calculated as: OCg (mg/g/h) = OCT/WP, where WP is the shrimp's weight (g).

The thermal cofinance (Q10) value of the shrimp was determined by using the equation $Q10=(R2/R1)^{10/T2-T1}$

Where, R_2 and R^1 are the oxygen consumption at the respective temperatures T_2 and T_1 .



Map 1. A map representing the samples collection area in the Al-Mas'hab region in southern Iraq





Fig. 1. Collection process in trawling nets (right), transportation process using plastic bags (left), and individuals selected to conduct oxygen consumption experiments (bottom)



Fig. 2. Designing an oxygen consumption experiment in plastic tanks with measuring devices and thermometers installed to adjust the temperature

RESULTS

Before studying oxygen consumption, the half lethal concentration (LC50) of salinities was determined to establish the ranges in which oxygen consumption experiments are conducted. It was noted that at a salinity of 1ppt, the survival rate was 95%. At a salinity of 5ppt, there were no deaths (100% survival), and at a salinity of 10ppt, the survival rate was 80%. When the salinity increased to 15ppt, the survival rate dropped to 58%, and at 20ppt, mortality occurred in more than half of the juveniles, resulting in a survival rate of only 17%. Consequently, the highest salinity concentration was excluded from the oxygen consumption experiments (Fig. 3).

Fig. (4) shows the effect of different salinities on oxygen consumption (mg/O2/h/per shrimp) in juvenile shrimp at a temperature of 20- 21°C. We observed an increase in oxygen consumption with increasing salinity. At 1ppt, we noticed a gradual increase in consumption until the third hour: 0.06, 0.016, and 0.018, respectively, after which there is a decrease in oxygen consumption rate to 0.012. While at a salinity of 5ppt, we observed an increase in oxygen consumption in the first hour (0.018), dropping to 0.016 and 0.012 in the second and third hours, respectively, then rising to 0.019 in the fourth hour of the experiment. Oxygen consumption levels converged at 10ppt salinity, with a gradual increase until the third hour of the experiment: 0.020, 0.022, and 0.024, respectively, then a decrease occurred in the fourth hour to 0.019. At high salinity of 15ppt, oxygen consumption in the first hour recorded 0.024 and in the second hour 0.020, rising in the third and fourth hours to 0.036 and 0.034, respectively.

By raising the temperature to 25- 26°C, we noticed differences in the rate of oxygen consumption depending on the difference in salinity. At a salinity of 1ppt, we noticed stability in oxygen consumption in similar ranges, ranging between 0.018, 0.016, 0.016 and 0.014 in the four hours of the experiment, respectively. At a salinity of 5ppt, there was an increase in the consumption rate in the first and second hours, reaching 0.024 and 0.038, respectively, then the consumption decreased in the third hour to 0.014, and at the end of the experiment (fourth hour) it reached 0.01. When the salinity increased to 10ppt, there was an increase in the consumption values for the first three hours, reaching 0.022, 0.034, and 0.040, respectively, and a decrease in the fourth hour of the experiment to 0.010. At high salinity of 15ppt, we noticed that oxygen consumption rose in the first hour to 0.032, and a slight decrease slight in the fourth hour reaching 0.030 (Fig. 5).

When the temperature was risen to an extreme temperature, we generally noticed an increase in oxygen consumption. At a salinity of 1ppt, we noticed an increase in consumption in the first and second hours: 0.024 and 0.046, respectively, and it decreased in the third and fourth hours at the same rate, reaching 0.016. At 5ppt salinity, consumption converged in the first and second hours, reaching 0.023 and 0.024,

respectively, and decreased to 0.030 and 0.026 in the third and fourth hours of the experiment, respectively. At the salinities of 10 and 15ppt, the juvenile shrimp exhibited similar physiological patterns. Oxygen consumption in the first hour was 0.038mg/ O2 at 10ppt and 0.040mg/ O2 at 15ppt. In the second hour, the consumption decreased to 0.022mg/ O2 at 10ppt and 0.030mg/ O2 at 15ppt. However, in the third and fourth hours, there was an increase in consumption. At 10ppt, the rates were 0.024mg/ O2 in the third hour and 0.028mg/ O2 in the fourth hour. At 15ppt, the rates were 0.032mg/ O2 in the third hour and 0.038mg/ O2 in the fourth hour (Fig. 6).

When testing three temperature ranges (20-21, 25-26, and 29- 30°C) with different salinities, a difference was recorded in the rate of total oxygen consumption (mg/O2/h/per shrimp). At a salinity of 1ppt, the consumption according to the difference in temperature ranges reached between 0.024, 0.032, and 0.028, respectively. At 5ppt salinity, the total consumption was 0.032, 0.040 and 0.052, respectively. At 10ppt salinity, the total consumption was 0.044, 0.052 and 0.056, respectively. Total oxygen consumption increases with salinity up to 15ppt, reaching 0.056mg/ O2 in the 20- 21°C temperature range, 0.064mg/ O2 in the 25- 26°C range, and 0.072mg/ O2 in the 29- 30°C range (Fig. 7).

Fig. (8) shows that the metabolic rate (mg/O2/g/) was clearly associated with an increase in temperature and salinity. At a temperature of 20- 21°C, the metabolism increased exponentially with an increase in salinity, and was 0.024, 0.032, 0.040, and 0.058 at salinity (ppt) 1, 5, 10, and 15, respectively. At a temperature of 25- 26°C, metabolic rates were recorded at 0.032, 0.040, 0.052, and 0.064, respectively. At 29- 30°C metabolic rates increased from 0.028 at a salinity of 1ppt, and increased to 0.052 at a salinity of 5ppt, also increased to 0.056 at a salinity of 10ppt, as well as to 0.072 at a salinity of 15ppt.

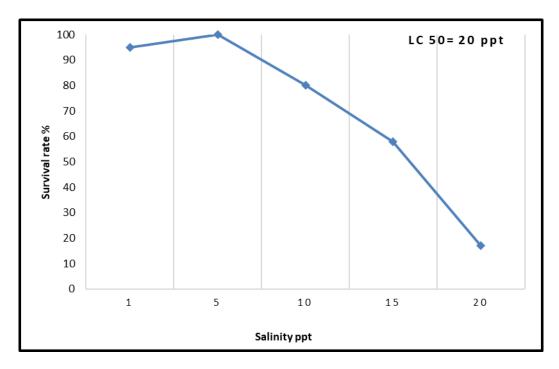


Fig. 3. The lethal concentration (LC 50) for shrimp *Metapnaeus affinis* within two hours of a sudden increase in salinity to concentrations 1, 5, 10, 15, and 20ppt

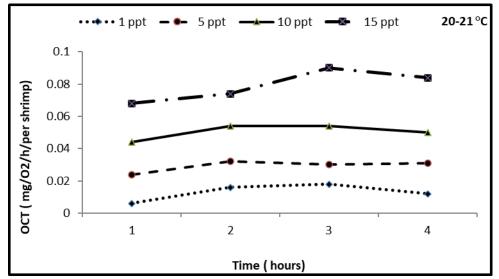


Fig. 4. Oxygen consumption rate (mg/h/ind./ww) of Juveniles *Metapenaeus affinis* at different salinity at constant temperature 20- 21°C

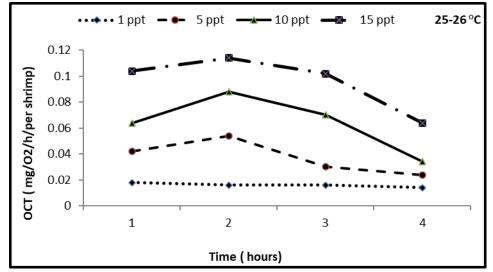


Fig. 5. Oxygen consumption rate (mg/h/ind./ww) of Juveniles *Metapenaeus affinis* at different salinity at constant temperature 25- 26°C

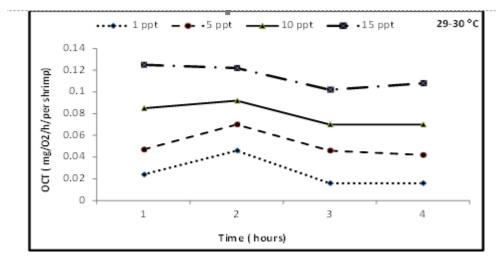


Fig. 6. Oxygen consumption rate (mg/h/ind./ww) of juveniles *Metapenaeus Affinis* at different salinity at constant temperature of 29- 30°C

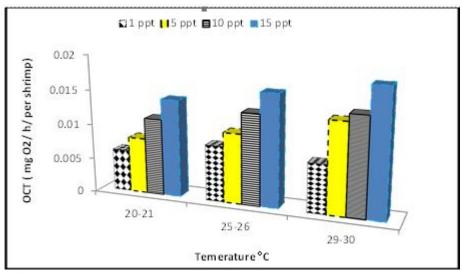


Fig. 7. Combined between different temperatures and salinities on total oxygen consumption (mg/h/ind./ww) after four hours for juveniles *Metapenaeus affinis*

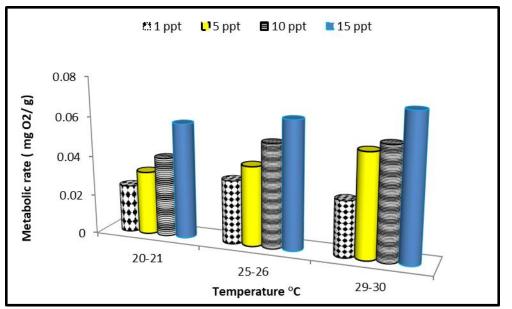


Fig. 8. Metabolic rate (mg O2 /g) for *M. affinis* juveniles at four experimental salinities and three temperature degree combination

Table (1) shows the thermal efficiency coefficient at temperature ranging from 20-25, 25-30, and 20-30°C, and at different salt concentrations of 1, 5, 10, and 15ppt. At salinity of 1ppt, the highest Q10 rate was achieved in the range of 20-25°C, and reached 1.77, and the lowest rate was achieved at a temperature of 25-30°C and reached 0.76, while in the thermal abrasion of 20- 30°C, it reached 1.36. On the other hand, at 5ppt salinity, the Q10 average ranged between 1.56, 1.69, and 2.64, respectively. Additionally, at 10ppt salinity, the Q10 ranged between the highest rate in the

temperature ranging 20- 30°C and reached 1.61, and the lowest rate in the temperature ranged between 25- 30°C, and reached 1.51, while in the temperature range of 20- 25°C the Q10 reached 1.40. A similar pattern was observed at a salinity of 15ppt, with the highest Q10 value of 1.28 recorded in the 20- 30°C temperature range. The lowest Q10 value of 1.12 was observed in the 25- 30°C range, while in the 20- 25°C range, the Q10 value reached 1.14.

Temperature range °C	Salinities (ppt)			
	1	5	10	15
20-25	1.77	1.56	1.40	1.14
25-30	0.76	1.69	1.15	1.12
20-30	1.36	2.64	1.61	1.28

Table 1. Estimation of the thermal coefficient (Q10) for *M affinis* juveniles at different salinities

DISCUSSION

M. affinis have an migrate phase in their life cycle and thus get exposed to wide fluctuations in salinity, since salinity is an important environmental variable that has a significant influence on dissolve oxygen of estuarine and marine animals, knowledge of the oxygen consumption under different salinity regimes will be useful to aquaculturists in formulating improved culture practices for maximizing growth and survival (Gerami, et al., 2012). Aquatic organisms in high salinity environments often expend more energy to osmotic balance and regulate the flow of water and salts across their membranes (Anongponyoskun et al., 2012). In this study, the salinities of 1 and 5ppt achieved a higher survival rate, this is because the salinity of the source is close to the salinity of the experiment. On the other hand, the survival rate decreased to 80% in the salinity of 10ppt and decreased to 58% in the salinity of 15ppt due to the extent of the salinity being far from the salinity of the source water. What confirms this is the decrease in the survival rate to 17%, meaning the percentage of deaths reached less than half the number when the salinity increased to 20ppt. Therefore, the experiment was stopped. Generally, salinity decreases the solubility of oxygen in water, this is because the presence of salts reduces the amount of space available for oxygen molecules. Consequently, high salinity environments tend to have lower dissolved oxygen levels compared to freshwater environments. Salinity seems to be a crucial factor for the survival due to its impact on shrimp respiration (Duan et al., 2022).

Oxygen needs to be increased when shrimp move to marine waters due to the increase in energy expended in metabolic processes accompanied by osmoregulation

(Havird et al., 2014; Rahi et al., 2020). The current study discussed the oxygen consumption and metabolic rate in shrimp when they suddenly moved to salt concentrations different from the salinity of the source water. A relative increase in the rate of oxygen consumption was observed when they suddenly moved to a salinity of 15ppt and a relative decrease when the salinity concentration decreased to 1ppt. These changes in the level of oxygen consumed with different salinity are associated with changes in the effective transport of ions (Rhai et al., 2018). There are a number of studies that addressed changes in metabolic rates when moving to environments with different salinity concentrations (Tantulo & Fotedar, 2006; Joseph & Philie, 2007; Ye et al., 2009; Rahi et al., 2018; Rahi et al., 2020; Jaffer et al., 2020; Rhai et al., 2021), these sources indicated that there are physiological changes associated with a change in salinity concentration that make the organism tend toward a short-term loss of energy. In many organisms, high salinity resulted in decreased oxygen consumption, the results in current study, agree with previous studies of Stern et al. (1984), Rosas et al. (2001), Spanopoulos et al. (2005), Garcia- Guerrero et al. (2013) and García-Guerrero et al. (2022). The interactions between salinity and temperature are complex and can significantly influence oxygen consumption in aquatic ecosystems. Changes in temperature and salinity can alter community structures and ecosystem functioning, leading to shifts in species within local habitats, particularly among aquatic organisms. In the current investigation, we observed differences in the rate of oxygen consumption depending on variations in salinity and temperature.

Higher temperatures and salinities together can significantly raise metabolic rates, leading to greater oxygen consumption (Villarrea *et al.*, 1994). Organisms in such environments may experience increased metabolic stress as they struggle to balance oxygen intake with elevated energy demands for maintaining homeostasis (Bett & Vinatea, 2009; Kieffer & Wakefilld, 2009; Abdul Wafi *et al.*, 2021). If oxygen becomes critically low, it can lead to reduced growth, impaired physiological functions and some species may migrate to areas with more favorable conditions (Allan *et al.*, 2006). Hence, we believe that high temperatures and salinities in past periods may have caused some species to shift from or become depleted in the local environment.

Thermal coefficient (Q10) is a measure of how temperature affects the rates of biochemical reactions, expressing the percentage increase in reaction rate when the temperature is raised by 10°C, changes in temperature and salinity affect the osmotic pressure inside the cells, and high or low osmotic pressure can change the activity of enzymes and other chemicals and thus affect Q10. During the current study, shrimp showed a similarity in Q10 values with differences in temperature and salinity, and this explains the ability of this species to tolerate wide ranges of temperatures between 20-28°C, in addition to its wide tolerance to differences in salinity, as it is a migratory species between the marine and freshwater (Salman *et al.*, 1990). The current study agrees with a study of Retes *et al.* (2008) that concluded that the value of Q10 ranges

between 2-3. Sudden changes in temperature can cause environmental stress to the shrimp, which leads to changes in biochemical processes and increases the shrimp's energy consumption (Haas, 2007; Huang *et al.*, 2023).

CONCLUSION

The research clearly shows that salinity and temperature levels have a direct effect on the rate of oxygen consumption in juvenile shrimp. As salinity and temperature increase, the rate of oxygen consumption also rises. Juvenile shrimp demonstrate a noticeable adaptation to decreasing oxygen levels by reducing their consumption rate in proportion to the oxygen concentration in their environment.

REFERENCES

Abbas, A.T and Ghazi, A.H. (2021). Commercial shrimp landings of two penaeid shrimps in the main markets of Basrah Province, Iraq. Mesopot. J. Mar. Sci., 36(1): 73 – 78.

Abdul Wafi; Ariadi, H.; Abdul Muqsith; Mahmudi, M. and Fadjar, M. (2021). Oxygen Consumption of *Litopenaeus vannamei* in Intensive Ponds Based on the Dynamic Modeling System. Journal of Aquaculture and Fish Health, 10:1, 18-24.

Al-Adhub, A.H. Y. (1987). On a new sub species of freshwater shrimp (Decapoda, Atyidae) from the Shat Al-Arab River, Iraq. Crustaceana, 53(1), E. J. Brill, Leiden.

Al-Adhub, A.H.Y. and Hamzah, H. A. (1987). *Caridina Babulti basrensis* from Shatt Al-Arab region, Iraq (Decapoda, Atyidae). Crustaceana, 52(3): 225-228.

Allan, E.; Froneman, P. and Hodgson, A. (2006). Effects of temperature and salinity on the standard metabolic rate of the shrimp *Palaemon peringueyi*. Journal of Experimental Marine Biology and Ecology, 337:103–108.

Al-Mahmood, H.K.H.; Hassan, W.F.; Alhello, A.Z.; Hammood, A.I. and Muhson, N.K. (2015). Impact of low discharge and drought of the water quality of the Shatt Al-Arab and Al-Basrah Rivers (south of Iraq). J. Int. Acad. Res. Multidisciplinary, 3(1): 285-296.

Anongponyoskun M.; Choksuchart A.; Salaenoi J. and Aranyakananda P. (2012). Dissolved oxygen budget for Pacific White Shrimp (*Litopenaeus vannamei*) Culture in Earthen Ponds. Kasetsart J. (Nat. Sci.) 46: 751 – 758.

Bett, C. and Vinatea, L. (2009). Combined effect of body weight, temperature and salinity on shrimp *Litopenaeus vannamei* oxygen consumption rate. Brazilian Journal of Oceanography, 57(4): 305-314.

Cobo, M. L.; Sonnenholzner S.; Wille M. and Sorgeloos P. (2014). Ammonia tolerance of *Litopenaeus vannamei* (Boone) larvae. Aquaculture Research, 45:470-475.

Duan, Y.; Li, M.; Sun, M.; Wang, A.; Chai, Y.; Dong, J.; Chen, F.; Yu, Z.; Zhang, X. (2022). Effects of Salinity and Dissolved Oxygen Concentration on the Tail-Flip Speed and Physiologic Response of Whiteleg Shrimp, *Litopenaeus vannamei*. Sustainability, 14, 15413.

Fischer, W. and Bianchi, G. (1984). FAO species identification sheets for fisheries purposes Western Indian Ocean (Fishing area 51), Volume 5, FAO, Rome.

García-Guerrero, M.; Orduña-Rojas, J. and Cortés-Jacinto, E. (2013). **Oxygen** Consumption of the Prawn *Macrobrachium americanum* over the Temperature Range of its Native Environment and in Relation to its Weight, North American Journal of Aquaculture, 73:3, 320-326.

García-Guerrero1, M.; Avilés-Espinoza, N.; Lizarraga-Sanchez, G. Herrera-Rodríguez, G.; Valdez-Martínez, D. and Hernández-Sandoval, P. (2022). Maximum critical temperature and oxygen consumption during thermoregulation in *Macrobrachium americanum* (Bate, 1868) adult prawns. Latin American Journal of Aquatic Research, 5(2): 301-309.

Gerami, M.H; Paighambari, S. Y.; Ghorbani, R. and Mohammad Momeni, M. (2012). Population Structure, Growth and Mortality Rates of Jinga Shrimp, *Metapenaeus affinis* in Fishing Grounds of Hormozgan Province, Iran. Caspian Journal of Applied Sciences Research, 1(8): 29-35.

Ghazi, A.H. and Hassan, H.F. (2021). New Record of *Macrobrachium lar* (Fabricius, 1798) (Crustacea: Decapoda: Palaemonidae) from Al-Hammar Marsh, Southern Iraq. Journal Biological and Applied Environmental Research, 5(1): 33-43.

González, R, Díaz, F., Licea, A., Re, A.D., Sánchez, L.N. and García-Esquivel, S. (2010). Thermal preference, tolerance and oxygen consumption of adult white shrimp *Litopenaeus vannamei* (Boone) exposed to different acclimation temperatures. Journal of Thermal Biology, 35: 218-224.

Haas, R. (2007). The role of coenzyme Q in cellular metabolism: Current biological andclinical aspects. Mitochondrion, 7 (Suppl. 1): 1-18.

Habashy, M.M. and Hassan, M.M. (2011). Effects of temperature and salinity on growth and reproduction of the freshwater prawn, *Macrobrachium rosenbergii* (Crustacea- Decapoda) in Egypt. Inter. J. of Envi. Sci. and Engine., 1:83-90.

Hassan, H.F; Abaas, T.A. and Ghazi, A.H. (2023). New record of *Macrobrachium equidens* shrimp (Dana, 1852) (Crustacea: Decapoda: Palamonidae) as invasive species in Al- Mashaab River, East of Al- Hammar marsh, Southern Iraq. Mesopot. J. Mar. Sci., 38(1):1-8

Havird, J.C.; Santos, S.R. and Henry, R.P. (2014). Osmoregulation in Hawaiian anchialine shrimp *Halocaridina rubra* (Crustacea: Atyidae): Expression of ion transporters, mitochondria-rich cell proliferation and hemolymph osmolality during salinity transfers. J. Exp. Biol., 217, 2309–2320.

Huang, Y., Ge, R.; Lou, G.; Jiang, N.; Zhu, X and Guo, Y. (2023). The influence of dietary Coenzyme Q10 on growth performance, antioxidant capacity and resistance against Aeromonas hydrophila of juvenile European eel (Anguilla anguilla). Fish and Shellfish Immunology, 138, 108834.

Jaffer, Y.D.; Saraswathy, R.; Ishfaq, M.; Antony, J.; Bundela, D.S. and Sharma, P.C. (2020). Effect of low salinity on the growth and survival of juvenile Pacific white shrimp, *Penaeus vannamei*: A revival. Aquaculture, 515, 734561.

Joseph, A. and Philip, R. (2007). Acute salinity stress alters the haemolymph metabolic profile of Penaeus monodon and reduces immunocompetence to white spot syndrome virus infection. Aquaculture, 272: 87–97.

Kieffer, J.D. and Wakefield, A.M. (2009). Oxygen consumption, ammonia excretion and protein use in response to thermal changes in juvenile atlantic salmon Salmo salar. Journal of Fish Biology, 74, 291-603.

Manush, M. K.; Pal, T.; Chatterjee, S.; and Mukherjee, C. (2004). Thermal tolerance and oxygen consumption of *Macrobrachium rosenbergii* acclimated to three temperatures. Journal of Thermal Biology, 29:15–19.

Miquel, L. C. (1983). Supplementary notes on species of *Metapenaeus* (Decapoda: Penaeidae). Crustaceana, 45: 71-76.

Rahi, M.L.; Ferdusy, T.; Ahmed, S.W.; Khan, M.N.; Aziz, D. and Salin, K.R. (2020). Impact of salinity changes on growth, oxygen consumption and expression pattern of selected candidate genes in the orange mud crab (*Scylla olivacea*). Aquac. Res., 51, 4290–4301.

Rahi, M.L.; Mahmud, S.; Dilruba, K.J.; Sabbir, W.; Aziz, D. and Hurwood, D.A. (2021). Temperature induced changes in physiological traits and expression of selected candidate genes in black tiger shrimp (*Penaeus monodon*) larvae. Aquac., Rep. 19, 100620.

Rahi, M.L.; Moshtaghi, A.; Mather, P.B. and Hurwood, D.A. (2018). Osmoregulation in decapod crustaceans: Physiological and genomic perspectives. Hydrobiologia, 825, 177–188.

Rahi,M.L.; Azad, K.N.; Tabassum,M.; Irin, H.H.; Hossain, K.S.; Aziz, D.; Moshtaghi, A. and Hurwood, D.A. (2020). Effects of Salinity on Physiological, Biochemical and Gene Expression Parameters of Black Tiger Shrimp (Penaeus monodon): Potential for Farming in Low-Salinity Environments. Biology, 2021,10,2020: 1-6.

Reyes, B.A.; Pendergast, J.S. and Yamazaki, S. (2008). Mammalian peripheral circadian oscillator are temperature compensated. J Biol Rhythms, 23(1): 8-95.

Rosas, C.; López, N.; Mercado, P. and Martínez, E. (2001). Effect of salinity acclimation on oxygen consumption of juveniles of the white shrimp *litopenaeus vannamei*. journal of crustacean biology, 21(4): 912–922.

Salman, D.S., Ali, M.H. and Al-Adhub, A.H. (1986). The Penaeid shrimp Metapenaeus affinis within the Iraqi waters. Oceanography of Khor Al-Zubair, Marine Science Center, 7: 417-447.

Salman, S. D. and Bishop, J.H. (1990). *Exopalaemon styliferus* in the Northern Arabian Gulf and in the inland water of Iraq (Decapoda, Caridea, Palaemonidae). Crustaceana, 59 (3): 281 – 288.

Salman, S. D., Ali, M.H. and Al-Adhub, A.H.Y. (1990). Abundance and seasonal migration of the penaeid shrimp *Metapenaeus affinis* within Iraqi waters. Hydrobiologia, 196: 79-90.

Salman, S.D.; Page, T.J.; Naser, M.D. and Yasser, A.G. (2006). The invasion of *Macrobrachium nipponense* (De Haan, 1849) (Caridea: Palaemonidae) into the Southern Iraqi marshes. Aquat. Invasions, 1(3): 109–115.

Schuler D. J. (2008). Acute toxicity of ammonia and nitrite to white shrimp (*Litopenaeus vannamei*) at low salinities. Master's thesis, Virginia Polytechnic Institute and State University, Blacksburg, 76 pp.

Spanopoulos, M., C.; Mart'inez-Palacios, R.; Venegas, C. and Rosas, L. (2005). The combined effects of salinity and temperature on the oxygen consumption of juvenile shrimps *Litopenaeus stylirostris* Stimpson, 1874. Aquaculture, 244:341–348.

Stern, S.; Borut, A. and Cohen, D. (1984). The effect of salinity and ion composition on oxygen consumption and nitrogen excretion of *Macrobrachium rosenbergii*. Comparative Biochemistry and Physiology - Part A: Molecular & Integrative Physiology, 79: 271-274.

Tantulo, U. and Fotedar, R. (2006). Comparison of growth, osmoregulatory capacity, ionic regulation and organosomatic indices of black tiger prawn (*Penaeus monodon* Fabricius, 1798) juveniles reared in potassium fortified inland saline water and ocean water at different salinities. Aquaculture, 258, 594–605.

Villarreal, H.; Hinojosa, P. and Naranjo, J. (1994). Effect of temperature and salinity on the oxygen consumption of laboratory produced *Penaeus vannamei* postlarvae. Comp. Biochem. Physiol., 108A, 331-336.

Vinagre, C.; Leal, I.; Mendonça, V. and Flores, A.V. (2015). Effect of warming rate on the critical thermal maxima of crabs, prawn and fish. Journal of Thermal Biology, 47: 19-25.

Ye, L.; Jiang, S.; Zhu, X.; Yang, Q.; Wen, W. and Wu, K. (2009). Effects of salinity on growth and energy budget of juvenile Penaeus monodon. Aquaculture, 2009, 290, 140–144.