

## Reproductive Biology of the Amazon Sailfin Catfish *Pterygoplichthys pardalis* from Tempe Lake, South Sulawesi, Indonesia

Andi Tamsil\*<sup>1</sup>, Hasnidar<sup>1</sup>, Ernaningsih<sup>1</sup>, Hasrun<sup>1</sup>, Andi Muhammad Akram<sup>2</sup>

<sup>1</sup>Faculty of Fisheries and Marine Science, Universitas Muslim Indonesia, Makassar 90231

<sup>2</sup>Faculty of Engineering, Universitas Muslim Indonesia, Makassar 90231

\*Corresponding Author: [andi.tamsil@umi.ac.id](mailto:andi.tamsil@umi.ac.id)

### ARTICLE INFO

#### Article History:

Received: Jan. 28, 2024

Accepted: May 27, 2024

Online: June 7, 2024

#### Keywords:

*P. pardalis*,  
Reproductive,  
Tempe Lake,  
Spawning,  
Maturity,  
Fecundity

### ABSTRACT

*Pterygoplichthys pardalis* is an invasive fish species that grows rapidly in Lake Tempe, Kabupaten Wajo, and has disturbed the existence of native, endemic, and introduced fish. This study aimed to analyze the reproductive biology of this species in terms of population control. The research was conducted from January to April 2023 at Tempe Lake, Wajo Regency, South Sulawesi. Fish samples were caught using gill nets, cleaned, and stored in a coolbox for observation in the laboratory. Reproductive biology observations began with the separation of male and female fish, measuring the length and weight of each with a fish measuring board and analytical scales. Samples were dissected for maturity gonad and fecundity observations. Length-frequency distribution of male fish ranged from 10.08- 41.58cm, while that of female fish ranged from 10.08- 35.28cm. The overall sex ratio between male and female fish was 1:1 (balanced), while the sex ratio between mature males and females (maturity stage IV) was 1:2 (polygamy). The growth pattern exhibited a negative allometric trend, with a long spawning season reaching its peak in March. The species is considered a partial spawner. The length at first maturity of gonads ( $L_m$ ) for male and female fish were 28,99 and 25.98cm, respectively. Moreover, the fecundity ranged from 201– 5,050 eggs with an average of  $1,762 \pm 966$  eggs.

### INTRODUCTION

*Pterygoplichthys* spp. originates from the South American Amazon River (Kottelat & Whitten, A.J., 1996; Armbruster & Page, 2006; Bijukumar *et al.*, 2015; Wahyudewantoro, 2018). Moreover, it has invaded waters around the world, both tropical and sub-tropical waters such as Philippines (Joshi, 1989; Hubilla & Primavera, 2007), Japan (Nakabo, 2002), Indonesia, Malaysia, Singapore (Page & Robins, 2006), Turkey (Özdilek, 2007), Mexico (Wakida-Kusunoki *et al.*, 2007; Orfinger & Goodding, 2018), Bangladesh (Hossain *et al.*, 2008), Europe (Keszka *et al.*, 2008; Piazzini *et al.*, 2010), Taiwan (Wu *et al.*, 2011), the United States (Nico *et al.*, 2012), Israel (Golani & Snovsky, 2013), Vietnam (Zworykin & Budaev, 2013), Sri Lanka (Sumanasinghe & Amarasinghe, 2014), and India (Bijukumar *et al.*, 2015).

The distribution of *Pterygoplichthys* spp. in Indonesia is extensive, and the population is growing very fast including that found in Lake Tempe, Kabupaten Wajo. The population has grown rapidly in the last five years and has affected fish productivity (Hasnidar *et al.*, 2021). According to Hill and Lodge (1999), *Pterygoplichthys* spp. is included in the invasive

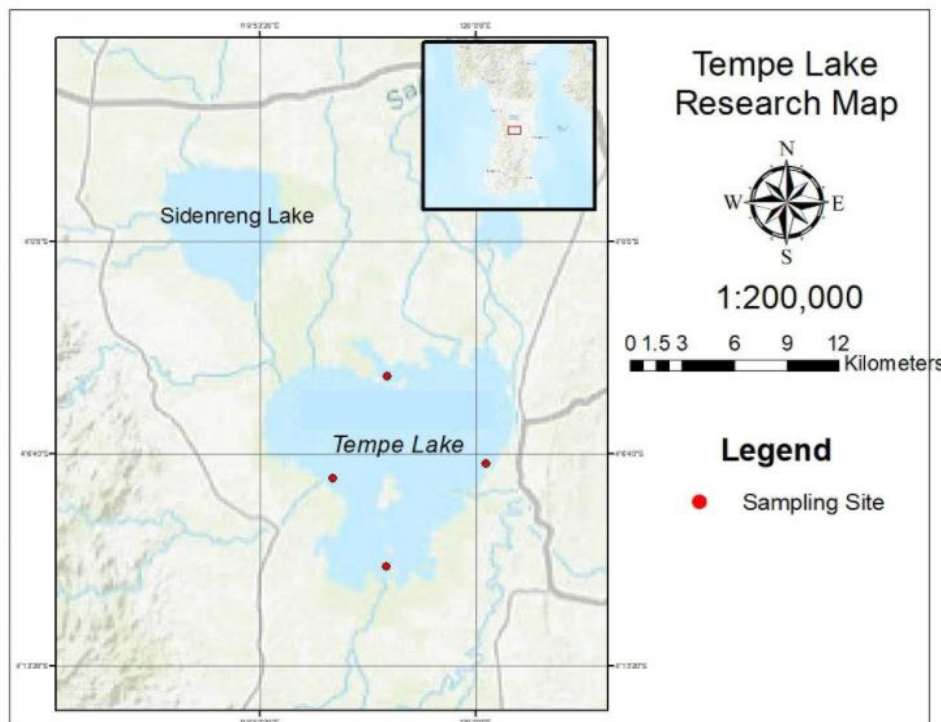
species. Invasive species can be both predators and competitors to native species. The competition between these invasive fish and native species can result in negative impacts on local fisheries (Keszka *et al.*, 2008), reducing biodiversity and contributing to declining native species populations worldwide (Keszka *et al.*, 2008; Vilà *et al.*, 2011; Hossain *et al.*, 2019).

*Pterygoplichthys* spp. has fast growth and high reproductive ability (Hoover *et al.*, 2004; Gibbs *et al.*, 2008). Moreover, it has good adaptation to polluted and oxygen-deficient waters (Sakai *et al.*, 2001; Chavez *et al.*, 2006), and has gills as their main respiratory organ. Additionally, it possesses additional respiratory organs, namely the stomach, which has undergone modifications allowing it to absorb oxygen from outside the water (Armbruster, 1998). It is an omnivore, capable of consuming algae, protozoa, microfungi, organic substances (detritus), and other microbes (Cardoso *et al.*, 2017). Moreover, it lacks predators and competitors (Hill & Lodge, 1999), and it is not commonly consumed by the public, and has no other significant uses.

Research on the reproductive biology of *Pterygoplichthys* spp.; namely, length size distribution, sex ratio, length and weight relationship, stage of gonad maturity, size at first maturity, spawning season and fecundity are needed. Information from the results of this study can be a reference for population control of *Pterygoplichthys* spp., especially in Lake Tempe.

## MATERIALS AND METHODS

*Pterygoplichthys pardalis* specimens were collected from Tempe Lake in Kabupaten Wajo, South Sulawesi, Indonesia (Fig. 1).



**Fig. 1.** Map of locations for collecting *Pterygoplichthys pardalis* from Lake Tempe, Wajo Regency, South Sulawesi

Sampling was carried out every week for four months from January–April 2023 using gill nets. Fish samples that were caught were first cleaned, drained, and then put in a cool box and covered with ice cubes. Observation of samples was carried out in the laboratory.

### Sample measurement

The measurements taken were: 1) Total length was measured using a measuring board with an accuracy of 0.1cm. 2) Weight of fish was measured using a digital scale with an accuracy of 0.01g. 3) Determination of the stage of gonad maturity was based on the morphology of the gonads (shape, size, color), the position of the gonads in the abdominal cavity (Wahidin, 2021), and the diameter of the eggs was measured using an ocular micrometer on a microscope (magnification 40 x).

### Length- frequency distribution

Length-frequency distribution involved determining the number of classes according to Sturges' instructions (Mahendra & Parmithi, 2015). The number of classes was determined using the formula  $k = 1 + 3.3 \log n$ , where  $k$  = the number of classes, and  $n$  = the number of data. While class intervals was determined using the equation  $c = \frac{X_n - X_1}{k}$ , where  $c$  = class interval;  $X_n$  = largest data value;  $X_1$  = smallest data value, and  $k$  = number of classes.

### Sex ratio

The sex ratio of male to female individuals was assessed. Sex ratio was estimated as: M/F (Peña-Mendoza *et al.*, 2005), where M=Number of males, and F=Number of females.

### Length-weight relationship

The length-weight relationship shows the relationship between the weight and total length of the fish using the exponential plot. The exponential plot was done using the expression:  $W = aL^b$  (LeCren, 1951), where  $W$ =Weight of fish (g);  $L$  = Length of fish (cm),  $a$  = Constant (intercept),  $b$  = growth pattern (slope) or exponent showing the length- weight relationship parameters (isometric or allometric). To get this equation, the values of  $L$  and  $W$  were transformed to logarithms (base 10) as follows:  $\log W = \log a + b \log L$ . If the value of  $b = 3$ , it indicates an isometric growth (growth in length with weight), if the value of  $b \neq 3$ , it indicates an allometric growth ( $b > 3$  is positive allometric: weight growth is faster than length;  $b < 3$  is negative allometric: length growth is faster than weight).

### Length at first maturity ( $L_{m50}$ )

The length at first maturity represents the size at which 50% of the female individuals of the fish species are mature. The  $L_{m50}$  was estimated using the log transformed equation of the logistic curve: with :  $\frac{p}{1 + e^{r(L - l_m)}}$  (King, 1995); dimana  $p$  = adjusted population ripe;  $L$  = length of fish (cm), and  $L_m$  = length at first maturity.

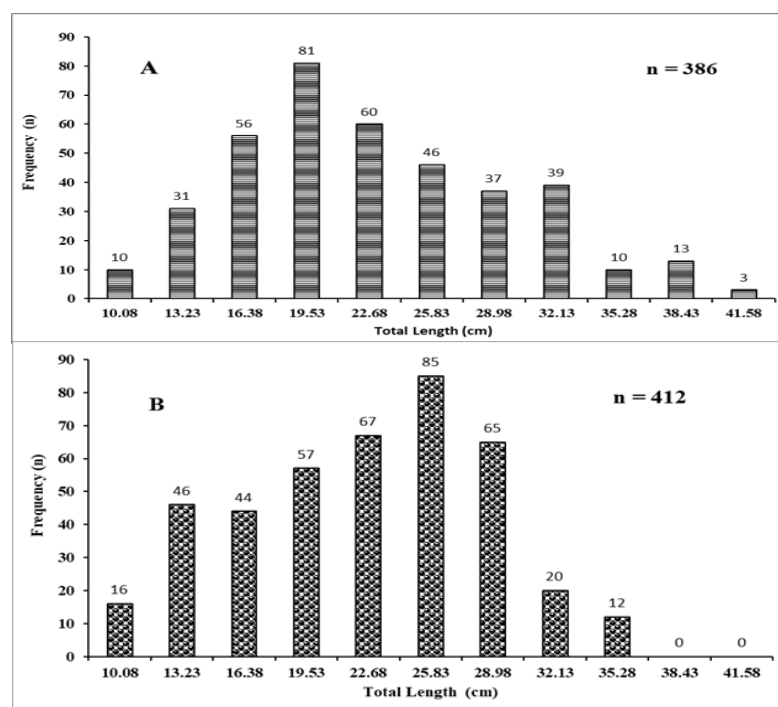
### Fecundity

Mature female fish (stage IV) were dissected, and the gonads were removed, and the overall gonad weight ( $W_g$ ) was measured, then the gonads were taken (anterior, middle and posterior) and the sub-gonad weight ( $W_s$ ) was measured. Sub gonads were given Gilson's fluid (Sivashanthini, 2008). The gravimetric method, as described by LeCren (1951) and Hasan *et al.* (2020), was used to calculate fecundity. This method involves counting the number of eggs contained in the sub-gonad ( $F_s$ ) and applying the formula  $F = (W_g / W_s) \times F_s$ , where  $F$  represents fecundity in eggs;  $W_g$  is the gonad weight in grams;  $W_s$  is the sub-gonad weight in grams, and  $F_s$  is the number of eggs in the sub-gonad.

## RESULTS

### 1. Length-frequency distribution

The total count of male *P. pardalis* captured was 386 individuals. Based on the length-frequency distribution, the length of the fish caught ranged from 10.08- 41.58cm. The largest proportion, comprising 81 individuals or 20.98%, was observed in the length category of 19.53cm. Conversely, the smallest proportion accounting for 3 individuals or 0.78% was recorded in the length category of 41.58cm (Fig. 2A). Among the captured female *P. pardalis*, totaling 412 individuals, the length-frequency distribution ranged from 10.08 to 35.28cm. The highest number was observed at 25.83cm, comprising 85 individuals or 20.63%, while the smallest proportion, represented by 3 individuals or 0.73%, was found at 35.28cm (Fig. 2B).



**Fig. 2.** Length-frequency distribution of (A) male and (B) female *P. pardalis*

### 2. Sex ratio

During the study, a total of 798 *Pterygoplichthys pardalis* were observed, comprising 386 males and 412 females. The results of the chi-square test analysis yielded a sex ratio male and female in this study of 1:1 or 48:52% (Fig. 3). Furthermore, the sex ratio from gonad maturity (stage IV) was 38:62% or 1:2 (Fig. 4).

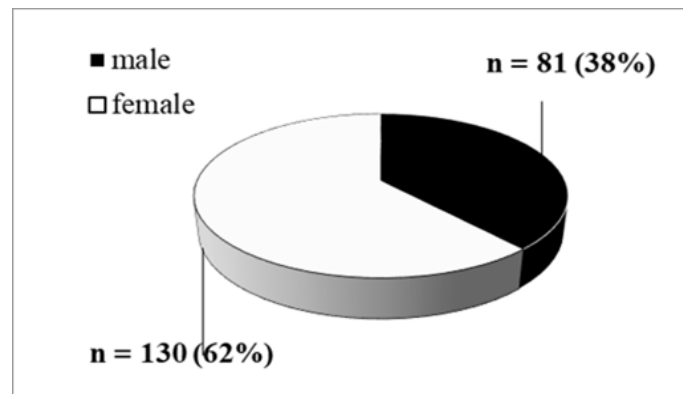


Fig. 3. Number of male and female *P. pardalis*

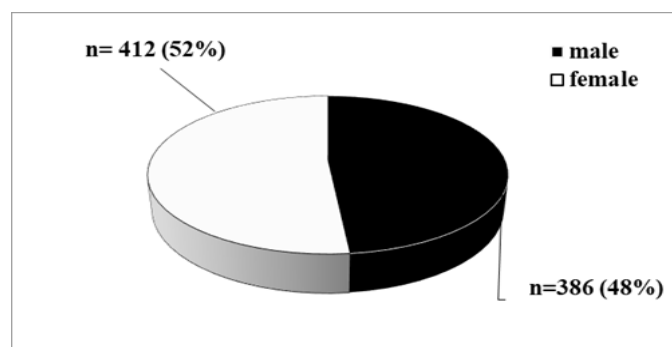


Fig.4. Number of mature male and female *P. pardalis*

### 3. Relationship of length-weight

The analysis of the relationship between length and weight of *P. pardalis* yielded the following equations:  $W = 0.02 L^{2.7069}$  for males and  $W = 0.02 L^{2.6951}$  for females (Fig. 5).

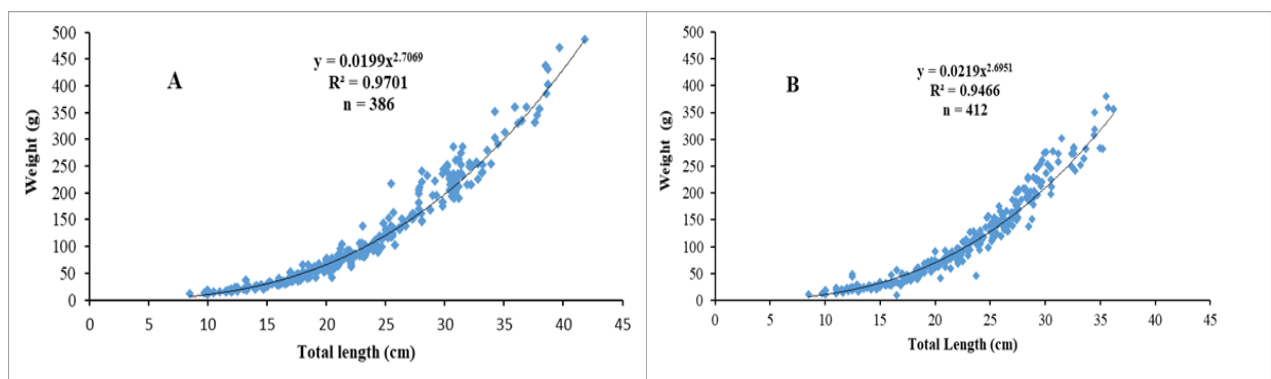


Fig. 5. Relationship of length-weight between (A) male and (B) female *P. pardalis*

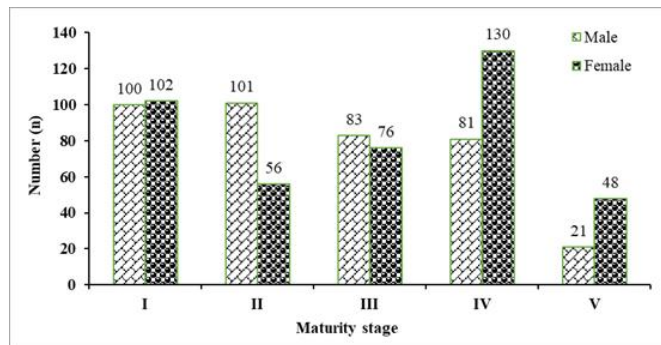
### 4. Gonad maturity stage

Criteria for the gonad maturity stage male and female *P. pardalis* were based on gonadal morphology (shape, size, color, and position of the gonads in the abdominal cavity) and size (egg diameter) (Table 1).

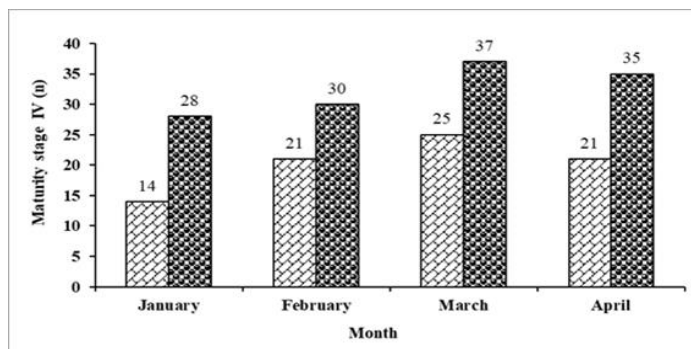
**Table 1.** Gonad maturity stage of female and male *P. pardalis*

Stage	Female (Ovary)	Male (Testes)
I (Immature)	Ovaries are small, elongated in shape, translucent white to pink in color, filled with a viscous liquid, the eggs are not visible, it is estimated that <15% fills the abdominal cavity.	Testes are elongated, very small and clear in color
II (Development)	Ovaries are larger in size, light yellow to red in color, the eggs are visible but the size is very small, the diameter of the eggs ranges from 0.20 to 1.60 mm, it is estimated that 20-40% fills the abdominal cavity.	Testes are larger than maturity stage I and are white.
III (Early mature)	Ovaries are large, yellow in color, the eggs are clearly visible but the eggs are still difficult to separate. There are small and large eggs. Small egg diameter ranges from 0.30 – 0.80 mm, large eggs range from 0.80 – 2.50 mm. It is estimated that 30% -60% fills the abdominal cavity.	Testes have a size larger than maturity stage II, the surface of the testes looks rough and white in color.
IV (Mature)	The ovaries are getting bigger, the color is yellow to orange, the eggs are clearly visible and the eggs are easy to separate. There are medium and large eggs. Small egg diameter ranges from 0.50 – 1.70 mm, large eggs range from 1.50 – 2.7 mm. It is estimated that 50-80% fills the abdominal cavity.	Testes are larger than maturity level III, filled with white viscous fluid.
V (spent)	Ovaries are wrinkled and red in color, containing leftover eggs that are not spawned	Testes are wrinkled and look like stage II maturity.

*Pterygoplichthys pardalis* both male and female were found to reach gonad maturity stage I-V. The highest number of male *P. pardalis* was found at maturity stage II, while females were predominantly at stage IV (Fig. 6). Furthermore, both male and female *P. pardalis* with the highest gonad maturity stage IV were found in March (Fig. 7).



**Fig. 6.** Number of male and female *P. pardalis* based on gonad maturity stage



**Fig. 7.** Distribution of mature (stage IV) male and female in January-April 2022

### 5. Length at first maturity of gonads ( $L_{m50}$ )

The number of *P. pardalis* that matured in relation to length showed that males started to mature at 16.23cm and female at 13.23cm. The highest number of mature male and female *P. pardalis* were 32.13 and 29.98cm, respectively (Fig. 8). Furthermore, based on the length at first maturity of gonads ( $L_m$ ) of *P. pardalis*, males matured at 28.99cm and females at 25.98cm.

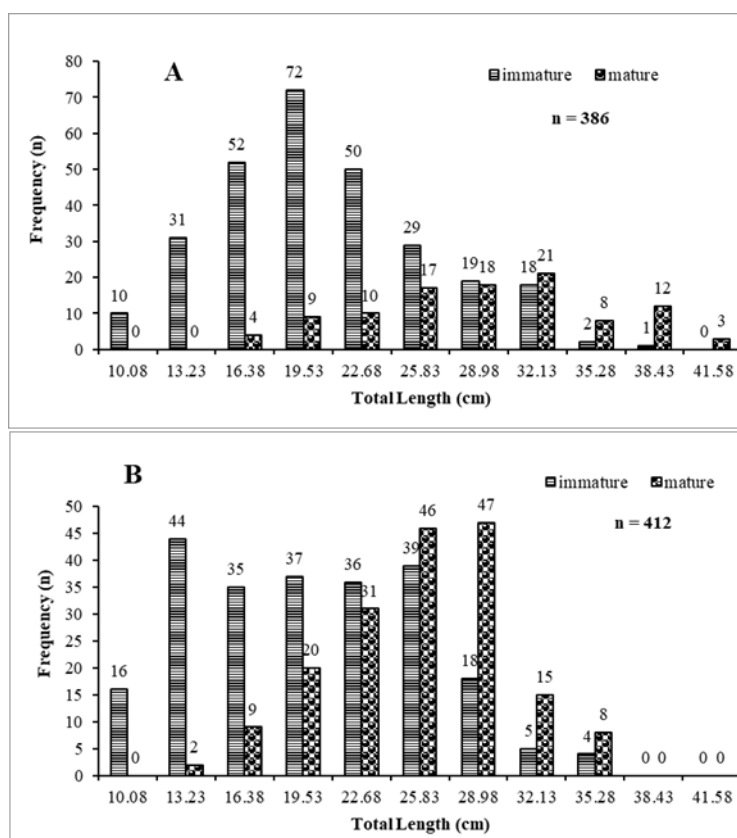


Fig. 8. Number of mature and immature (A) male and (B) female *P. pardalis*

### 6. Fecundity

During the observation, 130 mature female gonads (stage IV) of *Pterygoplichthys pardalis* were recorded, with lengths ranging from 13.23 to 35.28cm and weights from 28 to 380g. The gonad weight varied between 6.6 and 66.4g. Their fecundity ranged from 201 to 5,050 eggs, with an average of  $1,579 \pm 966$  eggs.

## DISCUSSION

The length-frequency distribution of *Pterygoplichthys pardalis* in Tempe Lake ranged from 10.08 to 41.58cm. This distribution varies across different locations and studies. For instance, in the Ciliwung River at Bogor Botanical Gardens, Indonesia, the reported length range is 27.40– 60.70cm (Hariandati, 2015). In Peninsular Malaysia, it's 10.08– 38.00cm (Samat *et al.*, 2008), while in Siak River in Pekanbaru, Indonesia, it's 5.25– 36.87cm (Sari *et al.*, 2015). In the Amazon River, the reported range is 21.00– 37.00cm (Sousa *et al.*, 2019a).

Moreover, in the Gangga-Brahmaputra River in Bangladesh, it's 33.40– 39.40cm (**Hossain *et al.*, 2018**). Other studies have reported lengths of up to 40cm (**Geerinckx *et al.*, 2007**). Moreover, some investigations reported lengths of up to 70 cm (**Kottelat & Whitten, 1996**; **Nelson *et al.*, 2016**; **Elfidasari *et al.*, 2022**). The differences in the length-frequency distribution of *P. pardalis* mentioned above are influenced by their habitats.

The sex ratio of male and female is 1:1 (Fig. 3), meaning that the number of male and female fish is balanced. The ratio of *P. pardalis* in the Air Hitam River, Riau Province, is the same (**Pinem *et al.*, 2016**). However, it differs from that observed in other locations. In Peninsular Malaysia, the sex ratio is reported to be 1: 2 (**Samat *et al.*, 2008**), and in the Ciliwung River at Bogor Botanical Gardens, it's also 1:2 (**Hariandati, 2015**). The sex ratio, which tends to have fewer males than females, is thought to be due to males guarding the nest after spawning. This result is the same as the findings of **Mazzoni and Iglesias-Rios (2002)** and **Liang *et al.*(2005)**, elucidating that male fish are responsible for protecting their eggs and young. In addition, it is also suspected that females are more active than males (**Pinem *et al.*, 2016**). The sex ratio between males and females based on maturity (stage IV) is 1: 2 (Fig. 4). Based on this concept, it is suspected that *P. pardalis* is polygamous, that is, one male is paired with more than one female. The same finding was reported by **Liang *et al.* (2005)** and **Gibbs *et al.* (2008)**, postulating that the number of mature female fish is more than the males in the spawning season. According to **Herjayanto *et al.* (2016)**, each fish species has a different optimal male and female sex ratio for spawning.

The length-weight relationship for both males and females has a negative allometric growth pattern, namely the growth in length is more dominant than weight (Fig. 5). The same is true for *P. pardalis* in Sungai Langat, Malaysia (**Samat *et al.*, 2008**) and the Amazon River (**Sousa *et al.*, 2019**). **Froese (2006)** suggested that, in general the growth pattern (value b) depends on the biological conditions of the fish such as gonad development and food availability. In addition to these factors, physiological conditions and environmental conditions, such as temperature, pH, salinity, geographic location, and sampling technique can also play significant roles.

Gonad maturity stage (ovaries and testes) of *P. pardalis* was classified into five stages (Table 1). The maturity classification is based on morphology, gonad position in the abdominal cavity, and egg diameter. The results showed that with higher maturity, there was an increase in both egg diameter and ovary volume. Variations in egg diameter were also found in the ovaries. **Pinem *et al.* (2016)** also classified the gonadal maturity of *P. pardalis* into five histological stages, non-vitellogenic eggs (stage I and II); yolk, non-vitellogenic and vitellogenic eggs (stage III); yolk, early vitellogenic eggs, vitellogenic, atretic eggs (stage IV); atretic eggs and shriveled ovaries (stage V).

Furthermore, according to **Jumawan and Herrera (2014)**, macro and microscopic observations of the gonads of *P. disjunctivus* showed that it was stage I (immature); stage II (the process of maturation, Ø eggs 1-2.5mm); stage III (mature; before spawning; Ø eggs 2.5-3mm); stage IV (spawning); stage V (after spawning, wrinkled ovaries still contain residual eggs); and stage VI (recovery, re-preparation for maturation of leftover eggs). According to **Samat *et al.* (2016)**, in *P. pardalis* from Peninsular Malaysia, the ovaries contain several groups of eggs, with some reaching Ø 3.3cm. Similarly in *P. disjunctivus*, several egg sizes were found in the ovary. Based on the maturity analysis, it is suspected that *Pterygoplichthys* spp. is a partial spawner (**Gibbs *et al.*, 2008**).

The number of male and female *P. pardalis* varied at each maturity stage (Fig. 6). Based on these maturity variations, it is suspected to have a long spawning time. This behavior was also noted in *P. pardalis* from the Air Hitam River, Riau Province, where stage IV fish were observed every month (**Pinem *et al.*, 2016**). This behavior occurs approximately eight months each year (**Neves & Ruffino, 1998**). Furthermore, based on the number of



mature (stage IV) relationship with the time of observation, the number of mature began to increase in January, the highest was in March and a decrease was detected in April (Fig. 7). Hence, it is suspected that the peak of spawning activity occurred in March. The spawning peaks of *P. pardalis* varied as observed all year round in Florida, USA (Yamamoto & Tagawa, 2000), March–September and sometimes reaching February in the United States (Liang *et al.*, 2005), June–July in the Philippine Marikina River (Gibbs *et al.*, 2008; Nico *et al.*, 2012; Gibbs *et al.*, 2013), April or May to September in US Hawaii (Gestring *et al.*, 2010), July–September and April–October in Taiwan (Jumawan & Herrera, 2014; Wickramaratne, 2018), and in the Ciliwung River, Bogor Botanical Gardens, July and August in Sri Lanka's Victoria & Kalaweva Reservoir (Hariandati, 2015). According to Mendoza-Palmero *et al.* (2009), the highest spawning of fish in sub-tropical areas usually lasts several months in summer, but in other areas, it can occur throughout the year.

Based on the size distribution of male and female length, the size at maturity of male gonads was 16.38cm and for females it was 13.23cm. The highest mature male and female fish were found at 32.13 and 28.98cm, respectively (Fig. 8). Furthermore, the results of the analysis of the length at first maturity of gonads (Lm50) indicated that female fish mature faster (25.98cm) compared to males (28.99cm). The length at first maturity of gonads found by several researchers also varied, including males at 25cm and females at 28cm (Neves & Ruffino, 1998); 25cm (Mendoza-Palmero *et al.*, 2009; Nico *et al.*, 2012); males at 12.5cm and females at 13cm (Samat *et al.*, 2016), and 22.3cm (Sousa *et al.*, 2019). The length at first maturity can vary due to environmental factors, fishing gear used, and fishing effort (Anderson *et al.*, 2008).

The fecundity of *P. pardalis* in this study is 201–5,050 (1,579±967 eggs). The fecundity of *P. pardalis* in several locations varied, such as from the Michoacán-Guerrero dam in Mexico, with 2,447 eggs (±12.6 eggs/g fish) (Rueda-Jasso *et al.*, 2013); from the Ciliwung River, Kebun Raya Bogor, where the fecundity is 207–1,445 eggs (Hariandati, 2015); from the Air Hitam River, Riau Province, with 5,351–48,980 eggs (Pinem *et al.*, 2016); from Sungai Langat, Malaysia, with 1,297–18,791 eggs, influenced by the brood size (Samat *et al.*, 2016); and from Victoria and Kalaweva Reservoirs in Sri Lanka, recording 956±261 and 1,856±817 eggs, respectively (Wickramaratne, 2018). According to Hoover *et al.* (2004) and Gibbs *et al.* (2008), *Pterygoplichthys* spp. are classified as fish with high fecundity; their fecundity ranges from 500–3,000 eggs/brood depending on the size of the brood (GISD, 2015). Variations in fecundity between fish populations are influenced by environmental factors, including water temperature, abundance of food, and different types of species (Alp *et al.*, 2003).

## CONCLUSION

The length distribution of male fish is larger than that of the female fish. Moreover, the overall sex ratio between male and female fish is balanced, and the spawning pairs of *P. pardalis* are polygamous (one male, two females). The growth pattern is negative allometric; the spawning season is long with the peak in March. *Pterygoplichthys pardalis* is a partial spawner, with the size at first maturity of the male and female gonads being 28.99 and 25.98cm, respectively. Fecundity ranges from 201 to 5,050 eggs, with an average of 1,762±966 eggs.

## REFERENCES

- Alp, A.; Kara, C. and Büyükçapar, H. M. (2003).** Reproductive biology of brown trout, *Salmo trutta macrostigma* Dumeril 1858, in a tributary of the Ceyhan River which flows into the eastern Mediterranean Sea. *Journal of Applied Ichthyology*, 19(6): 346–351. <https://doi.org/10.1111/j.1439-0426.2003.00455.x>
- Anderson, M. J.; Gorley, R. N. and Clarke, K. R. (2008).** *PERMANOVA + for PRIMER: PRIMER-E Ltd.*
- Armbruster, J. W. (1998).** Modifications of the digestive tract for holding air in loricariid and scoloplacid catfishes. *Copeia*, 1998(3): 663–675. <https://doi.org/10.2307/1447796>
- Armbruster, J. W. and Page, L. M. (2006).** Redescription of *Pterygoplichthys punctatus* and description of a new species of *Pterygoplichthys* (Siluriformes: Loricariidae). *Neotropical Ichthyology*, 4(4): 401–409. <https://doi.org/10.1590/s1679-62252006000400003>
- Bijukumar, A.; Smrithy, R.; Sureshkumar, U. and George, S.(2015).** Invasion of South American suckermouth armoured catfishes *Pterygoplichthys* spp. (Loricariidae) in Kerala, India - a case study. *Journal of Threatened Taxa*, 7(3): 6987–6995. <https://doi.org/10.11609/jott.o4133.6987-95>
- Cardoso, A. C. F.; Oliveira, M. S. B.; Neves, L. R. and Tavares-Dias, M.(2017).** Metazoan fauna parasitizing *Peckoltia braueri* and *Pterygoplichthys pardalis* (Loricariidae) catfishes from the northeastern Brazilian Amazon. *Acta Amazonica*, 47(2): 147–154. <https://doi.org/10.1590/1809-4392201603232>
- Chavez, J. M.; De La Paz, R. M.; Manohar, S. K.; Pagulayan, R. C. and Carandang VI, J. R.(2006).** New Philippine record of south american sailfin catfishes (Pisces: Loricariidae). *Zootaxa*, 1109, 57–68. <https://doi.org/10.11646/zootaxa.1109.1.6>
- Elfidasari, D.; Puspaningtias, F. C. and Fahmi, M. R.(2022).** Reproductive Biology Pleco (*Pterygoplichthys pardalis* Castelnau 1855) in Ciliwung River. *Jurnal Pembelajaran Dan Biologi Nukleus*, 8(2): 247–262. <https://doi.org/10.36987/jpbn.v8i2.2642>
- Froese, R. (2006).** Cube law, condition factor and weight-length relationships: History, meta-analysis and recommendations. *Journal of Applied Ichthyology*, 22(4): 241–253. <https://doi.org/10.1111/j.1439-0426.2006.00805.x>
- Geerinckx, T.; De Poorter, J. and Adriaens, D. (2007).** Morphology and development of teeth and epidermal brushes in loricariid catfishes. *Journal of Morphology*, 268(9): 805–814. <https://doi.org/10.1002/jmor.10547>
- Gestring, K. B.; Shafland, P. L. and S Stanford, S. (2010).** Status of the exotic orinoco sailfin catfish (*Pterygoplichthys multiradiatus*). *Florida Scientist*, 73(2), 122–137.
- Gibbs, M. A.; Shields, J. H.; Lock, D. W.; Talmadge, K. M. & Farrell, T. M.(2008).** Reproduction in an invasive exotic catfish *Pterygoplichthys disjunctivus* in Volusia Blue Spring, Florida, U.S.A. *Journal of Fish Biology*, 73(7): 1562–1572. <https://doi.org/10.1111/j.1095-8649.2008.02031.x>
- Gibbs, M.; Watson, P.; Johnson-Sapp, K. and Lind, C. (2017).** Reproduction revisited – a decade of changes in the reproductive strategies of an invasive catfish, *pterygoplichthys disjunctivus* (Weber, 1991), in Volusia Blue Spring, Florida. *Aquatic Invasions*, 12(2):

- 225–239. <https://doi.org/10.3391/ai.2017.12.2.10>
- Gibbs, Melissa A.; Kurth, B. N. & Bridges, C. D. (2013).** Age and growth of the loricariid catfish *Pterygoplichthys disjunctivus*. *Aquatic Invasions*, 8(2): 207–218.
- Global Invasive Species Database (GISD).(2015).***Species profile Pterygoplichthys pardalis*. <http://www.iucngisd.org/gisd/species.php?sc=1658>
- Golani, D.; & Snovsky, G. (2013).** Occurrence of suckermouth armored catfish (Siluriformes, Loricariidae, *Pterygoplichthys*) in inland waters of Israel. *BioInvasions Records*, 2(3): 253–256. <https://doi.org/10.3391/bir.2013.2.3.13>
- Hariandati, A.(2015).** *Aspek reproduksi ikan sapu-sapu (Pterygoplichthys pardalis) di Sungai Ciliwung Kebun Raya Bogor*. Institut Pertanian Bogor.
- Hasan, M.; Hosen, M. H. A.; Miah, M. I.; Ahmed, Z. F.; Chhanda, M. S. & Shahriar, S. I. M. (2020).** Fecundity, length at maturity and gonadal development indices of river catfish (*Clupisoma garua*) of the old Brahmaputra River in Bangladesh. *Egyptian Journal of Aquatic Research*, 46(3): 259–263. <https://doi.org/10.1016/j.ejar.2020.08.003>
- Hasnidar; Tamsil, A.; Muhammad Akram, A. & Hidayat, T.(2021).** Analisis kimia ikan sapu-sapu (*Pterygoplichthys pardalis* Castelnau 1855) dari Danau Tempe. *Jphpi 2021*, 24(1): 78–88.
- Herjayanto, M.; Carman, O. & Soelistyowati, D. T.(2016).** Tingkah laku memijah, potensi reproduksi ikan betina, dan optimasi teknik pemijahan ikan pelangi *Iriatherina werner* Meinken, 1974 [Spawning behavior, female reproductive potential and breeding technique optimize of threadfin rainbowfish *Iriatherina werne*. *Jurnal Iktiologi Indonesia*, 16(2): 171–183.
- Hill, A.M. and Lodge, D. M. (1999).** Replacement of resident crayfishes by an exotic crayfish: the roler of compotition and predation. *Ecological Applications*, 9(0), 678-690.
- Hoover, J. J.; Killgore, K. J. & Cofrancesco, A. F. (2004).** Suckermouth catfishes: threats to aquatic ecosystems of the United States? *Aquatic Nuisance Species Research Bulletin*, 4(1): 1–14.
- Hossain, M. Y.; Rahman, M. M.; Ahmed, Z. F.; Ohtomi, J. & Islam, A. B. M. S. (2008).** First record of the South American sailfin catfish *Pterygoplichthys multiradiatus* in Bangladesh. *Journal of Applied Ichthyology*, 24(6): 718–720. <https://doi.org/10.1111/j.1439-0426.2008.01117.x>
- Hossain, M. Y.; Vadas, R. L.; Ruiz-Carus, R. & Galib, S. M. (2018).** Amazon sailfin catfish *pterygoplichthys pardalis* (Loricariidae) in bangladesh: A critical review of its invasive threat to native and endemic aquatic species. *Fishes*, 3(1): 2016–2017. <https://doi.org/10.3390/fishes3010014>
- Hossain, S.; Hossain, S.; Shefat, T.; Latifa, G. A. & Obaida, A.(2019).** First record of invasive croaking gourami , *Trichopsis vittata* ( Cuvier 1831 ) in South-eastern Bangladesh. *Journal of Fisheries and Life Science*, 4(2): 6–9.
- Hubilla, M.; Kis, F. & Primavera, J. (2007).** Janitor fish *Pterygoplichthys disjunctivus* in the Agusan Marsh:A threat to freshwater biodiversity. *Journal of Environmental Science and Management*, 10(1), 10–23.
- Joshi, R. C.(1989).***Invasive alien species ( IAS ): Concerns and status in the Philippines*. 1–23.

- Jumawan, J. C. & Herrera, A. A. (2014).** Ovary morphology and reproductive features of the female suckermouth sailfin catfish, *Pterygoplichthys disjunctivus* (Weber 1991) from Marikina River, Philippines. *Asian Fisheries Science*, 27(1): 75–89. <https://doi.org/10.33997/j.afs.2014.27.1.006>
- Keszka, S.; Panicz, R. & Tański, A. (2008).** First record of the leopard pleco, *Pterygoplichthys gibbiceps* (Actinopterygii, Loricariidae) in the Brda river in the centre of Bydgoszcz (Northern Poland). *Acta Ichthyologica et Piscatoria*, 38(2): 135–138. <https://doi.org/10.3750/AIP2008.38.2.08>
- King, M. (1995).** *Fisheries Biology, Assessment and Management*. Blackwell Publishing Company.
- Kottelat, M. and Whitten, A. J. K. (1996).** *Freshwater Fishes of Western Indonesia and Sulawesi: Additions and Corrections*. 3, 830. <https://doi.org/10.2307/1447208>
- LeCren, E. D. (1951).** The length-weight relationship and seasonal cycle in gonad weight and condition in the perch (*Perca fluviatilis*). *Journal of Animal Ecology*, 20(2): 201–219.
- Liang, S. H.; Wu, H. P. and Shieh, B. Sen. (2005).** Size structure, reproductive phenology, and sex ratio of an exotic armored catfish (*Liposarcus multiradiatus*) in the Kaoping River of Southern Taiwan. *Zoological Studies*, 44(2): 252–259.
- Mahendra, W. E. and Parmithi, N. N. (2015).** *Statistik dasar dalam penelitian pendidikan*.
- Mazzoni, R. and Iglesias-Rios, R. (2002).** Distribution pattern of two fish species in a coastal stream in Southeast Brazil. *Brazilian Journal of Biology*, 62(1): 171–178. <https://doi.org/10.1590/S1519-69842002000100019>
- Mendoza-Palmero, C. A.; Sereno-Uribe, A. L. and Salgado-Maldonado, G. (2009).** Two new species of *Gyrodactylus* Von Nordmann, 1832 (Monogenea: Gyrodactylidae) parasitizing *Girardinichthys multiradiatus* (Cyprinodontiformes: Goodeidae), an endemic freshwater fish from Central Mexico. *The Journal of Parasitology*, 95(2): 315–318.
- Nakabo, T. (2002).** *Fishes of Japan: with pictorial keys to the species* (T. Nakabo (ed.)). Tokai University Press.
- Nelson, J.S.; Grande, T.C.; Wilson, M. V. (2016).** *Fishes of the World* (fifth). John Wiley & Sons. <https://www.ptonline.com/articles/how-to-get-better-mfi-results>
- Neves, A. M. B. & Ruffino, M. L. (1998).** Aspectos reproductivos do acari-bodo *Liposarcus pardalis* (Pisces, Siluriformes, Loricariidae) (Castelnau, 1855) do medio Amazonas. *Boletim Do Museu Paraense Emilio Goeldi. Serie Zoologia*, 14(1): 77–94.
- Nico, L. G.; Butt, P. L.; Johnston, G. R.; Jelks, H. L.; Kail, M. & Walsh, S. J. (2012).** Discovery of South American suckermouth armored catfishes (Loricariidae, *Pterygoplichthys* spp.) in the Santa Fe River drainage, Suwannee River basin, USA. *BioInvasions Records*, 1(3): 179–200. <https://doi.org/10.3391/bir.2012.1.3.04>
- Orfinger, A. B. & Goodding, D. D. (2018).** The global invasion of the suckermouth armored catfish genus *Pterygoplichthys* (Siluriformes: Loricariidae): Annotated list of species, distributional summary, and assessment of impacts. *Zoological Studies*, 57(7): 1–16. <https://doi.org/10.6620/ZS.2018.57-07>
- Özdilek, Ş. Y. (2007).** Possible threat for middle east inland water : an exotic and invasive

- species, *Pterygoplichthys disjunctivus* (Weber, 1991) in Asi River, Turkey (Pisces: Loricariidae). *E.Ü. Journal of Fisheries and Aquatic Sciences*, 24, 303–306.
- Page, L. M. & Robins, R. H.** (2006). Identification of sailfin catfishes (Teleostei: Loricariidae) in Southeastern Asia. *Raffles Bulletin of Zoology*, 54(2), 455–457.
- Peña-Mendoza, B.; Gómez-Márquez, J. L.; Salgado-Ugarte, I. H. & Ramírez-Noguera, D.** (2005). Reproductive biology of *Oreochromis niloticus* (Perciformes: Cichlidae) at Emiliano Zapata dam, Morelos, Mexico. *Revista de Biología Tropical*, 53(3–4), 515–522. <https://doi.org/10.15517/rbt.v53i3-4.14666>
- Piazzini, S.; Lori, E., Favilli, L.; Cianfanelli, S.; Vanni, S. and Manganeli, G.** (2010). A tropical fish community in thermal waters of Southern Tuscany. *Biological Invasions*, 12(9): 2959–2965. <https://doi.org/10.1007/s10530-010-9695-x>
- Pinem, F. M.; Pulungan, C. P. & Efison, D.**(2016). Reproductive biology of *Pterygoplichthys pardalis* in the air hitam river Payung Sekaki District, Riau Province. *Journal Online*, 3(1): 1–14. <https://jom.unri.ac.id/index.php/JOMFAPERIKA/article/view/9111/8777%0A%0A>
- Rueda-Jasso, R. A.; Campos-Mendoza, A.; Arreguín-Sánchez, F.; Díaz-Pardo, E. and Martínez-Palacios, C. A.**(2013). The biological and reproductive parameters of the invasive armored catfish *Pterygoplichthys disjunctivus* from Adolfo López Mateos El Infiernillo Reservoir, Michoacán-Guerrero, Mexico. *Revista Mexicana de Biodiversidad*, 84(1): 318–326. <https://doi.org/10.7550/rmb.26091>
- Sakai, A.K.; Allendorf, F.W.; Holt, J.S.; Lodge, D.M.; Molofsky, J.; With, K.A.; Baughman, S.; Cabin, R.J.; Cohen, J.E.; Ellstrand, N.C.; Mccauley, D.E.; O'neil, P.; Parker, I.M.; Thompson, J.N.; Weller, S. G.** (2001). The population biology of invasive species. *Annual Reviews Ecology System*, 32(0): 305–320.
- Samat, A.; Shukor, M. N.; Mazlan, A. G.; Arshad, A. and Fatimah, M. Y.** (2008). Length-weight Relationship and Condition Factor of *Pterygoplichthys pardalis* (Pisces: Loricariidae) in Malaysia Peninsula. *Research Journal of Fisheries and Hydrobiology*, 3(2): 48–53.
- Samat, A.; Yusoff, F. M.; Arshad, A.; Ghaffar, M. A.; Nor, S. M.; Magalhaes, A. L. and Das, S. K.**(2016). Reproductive biology of the introduced sailfin catfish *Pterygoplichthys pardalis* (Pisces: Loricariidae) in peninsular Malaysia. *Indian Journal of Fisheries*, 63(1): 35–41. <https://doi.org/10.21077/ijf.2016.63.1.44937-05>
- Sari, D. P.; Efison, Deni; Pulungan, C. P.** (2015). *Relationship between body length and the length of other body parts of Pterigoplihtys pardalis from the Air Hitam and Siak Rivers Riau* (Vol. 0, Issue 0). <https://doi.org/10.3969/j.issn.1008-0813.2015.03.002>
- Sivashanthini, K.**(2008). Reproductive Biology of the Whipfin Silverbiddy *Gerres filamentosus* Cuvier, 1829 from the Parangipettai Waters (SE coast of India). *Asian Fisheries Science*, 21(2): 127–145. <https://doi.org/10.33997/j.afs.2008.21.2.001>
- Sousa, R. G. C.; Oliveira, C. M.; Sant'Anna, I. R. A.; Marshall, B. G. and Freitas, C. E. de C.** (2019). Growth parameters and yield per recruit analysis for the armoured catfish *pterygoplichthys pardalis* sampled in the low reach of the amazonas river. *Boletim Do Instituto de Pesca*, 45(2). <https://doi.org/10.20950/1678-2305.2019.45.2.396>

- Sumanasinghe, H., and Amarasinghe, U.(2014).** Population dynamics of accidentally introduced Amazon sailfin catfish, *Pterygoplichthys pardalis* (Siluriformes, Loricariidae) in Pologolla reservoir, Sri Lanka. *Sri Lanka Journal of Aquatic Sciences*, 18(0): 37–45. <https://doi.org/10.4038/sljas.v18i0.7040>
- Vilà, M.; Espinar, J. L.; Hejda, M.; Hulme, P. E.; Jarošík, V.; Maron, J. L.; Pergl, J.; Schaffner, U.; Sun, Y. and Pyšek, P.(2011).** Ecological impacts of invasive alien plants: A meta-analysis of their effects on species, communities and ecosystems. *Ecology Letters*, 14(7): 702–708. <https://doi.org/10.1111/j.1461-0248.2011.01628.x>
- Wahidin, L. O.(2021).***1st \_ Buku Penuntun Praktikum BIOPER* (Issue January). <https://doi.org/10.13140/RG.2.2.34531.58403>
- Wahyudewantoro G.(2018).** Sapu-sapu (*Pterygoplichthys* spp.), ikan pembersih kaca yang bersifat invasif di Indonesia (Sailfin Armoured Catfish, *Pterygoplichthys* spp.: A Tank Cleaner has Become One of the Invasive Fish in Indonesia). *Warta Iktiologi*, 2(2): 22–28.
- Wakida-Kusunoki, A. T., Ruiz-Carus, R., & Amador-Del-Angel, E.(2007).** Amazon sailfin catfish, *Pterygoplichthys pardalis* (Castelnau, 1855) (Loricariidae), another exotic species established in southeastern Mexico. *Southwestern Naturalist*, 52(1): 141–144. [https://doi.org/10.1894/0038-4909\(2007\)52\[141:ASCP\]2.0.CO;2](https://doi.org/10.1894/0038-4909(2007)52[141:ASCP]2.0.CO;2)
- Wickramaratne, I.(2018).** Reproductive Biology of Vermiculated Sail Fin Cat Fish *Pterygoplichthys disjunctivus* in Victoria & Kalaweva Reservoirs in Sri Lanka. *E International Conference on Fisheries and Aquaculture*, 4: 01–14. <https://doi.org/10.17501/23861282.2018.4101>
- Wu, L. W.; Liu, C. C. and Lin, S. M.(2011).** Identification of exotic sailfin catfish species (*Pterygoplichthys*, Loricariidae) in Taiwan based on morphology and mtDNA sequences. *Zoological Studies*, 50(2): 235–246.
- Yamamoto, M. N. and Tagawa, A. W.(2000).***Hawaii's Native and Exotic Freshwater Animals*. Mutual Publishing.
- Zworykin, D. D. and Budaev, S. V.(2013).** Non-indigenous armoured catfish in Vietnam: Invasion and systematics. *Ichthyological Research*, 60(4): 327–333. <https://doi.org/10.1007/s10228-013-0356-9>