

## Effect of feed, manure and their combination on the growth of *Cyprinus carpio* (L.) fry and fingerlings

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### ABSTRACT

Two field experiments, each of 120-day, were conducted in twelve earthen ponds, each with dimensions 50 x 20 x 1.0m (length x width x depth) were located at Serow Fish Farm, National Institute of Oceanography And Fisheries, Dakahlia Governorate, Egypt, with a view to study the effect of nutrient inputs (feed, manure and their combination) on the common carp (*Cyprinus carpio*) fry and fingerlings of the fish. The treatments consisted of control (C, T1), only feed (F, T2), only manure (M, T3) and manure plus feed (M+F (T4)). Poultry manure was applied in split doses to ponds of manure treatments (M and M+F). Fry (Experiment one) and fingerlings (Experiment two) of average initial weight 0.67 g and 3.2 g respectively, were stocked seven days after the initial manure application at 5 individuals/m<sup>2</sup>. Fish in F and M+F treatments were provided a fish meal based pelleted diet once daily in the morning at 5% of body weight. The most dominant genera of phytoplankton encountered were *Microcystis*, *Anabaena* and *Microspora*, while among the zooplankton, *Keratella* and Nauplii dominated. M+F treatment had the highest plankton biomass ( $P<0.05$ ). Significant ( $P<0.05$ ) variation in both phyto and zooplankton dry weight was recorded with respect to the study period. Both feeding and manuring, individually and in combination, significantly improved ( $P<0.05$ ) the growth of the fish. The highest specific growth rate, final fish weight, and gross production were recorded in M+F treatment in both the experiments. The difference in survival among the control and treatments was not significant in experiment one ( $P>0.05$ ), whereas F and M+F treatments recorded lower survival ( $P<0.05$ ) in the second experiment. Fish production was comparable under feed (F) and manure (M) treatments ( $P>0.05$ ) in experiment one, but was significantly ( $P<0.05$ ) different in experiment two. The increment in gross fish production over the control was 103.22 and 119.99% in feed, 77.30 and 59.44% in manure and 162.34 and 175.08% in M+F treatments of the two experiments respectively. Carcass protein, fat and ash contents were significantly ( $P<0.05$ ) higher in the three treatments as compared to the control.

**Keywords:** Common carp, Natural food, poultry manure, Supplementary feed; Fish production, carcass composition.

### INTRODUCTION

Presently fish culture mainly depends on the application of organic fertilizers and to some extent on inorganic fertilizers. Fertilization enhances phytoplankton productivity in rearing and stocking ponds (New and Fedoruk, 2003; Bhakta *et al.*, 2004, 2006). Phytoplankton and zooplankton represent rich source of protein (40-60%) on dry weight basis which is sufficient for fish growth at low stocking densities (Silva and Anderson, 1995; Tabinda and Ayub, 2009, 2010; Sun *et al.*, 2010). Artificial feed is seldom applied, due to the cost and herbivore nature of common carp (*Cyprinus carpio*). With the increase in fish demand trend has developed to culture fish more intensively to enhance the present level of fish production. Common

carp has remained neglected in different culture practices, due to its in pond breeding habit and burrowing nature of feeding (Milstein *et al.*, 2002). Recently some farmers are trying to introduce this fish to replace *Cirrihinus mirgala* (a bottom feeder) in combination with indigenous major carps in semi-intensive culture system. The adoption of this fish in polyculture seems more promising because it leads to higher fish production, due to its fast growth and hardy nature. Moreover it keeps nutrients in suspension, due its burrowing nature (Saikia and Das, 2008). Farmers can get double benefit in introducing the common carp in the pond as it increases the availability of nutrients for phytoplankton which in turn enhances fish production. According to Jain (2002), *C. carpio* has the ability to survive under various climatic conditions and is found to be most suitable for many fish farming systems. He is also of the opinion that *C. carpio* has the potential to improve conditions in pond bottom soil, as a result soil perturbation increases the oxygen transfer to the soil, decreases the concentration of toxic compounds and enables more efficient food web recycling and nutrient release (Ritvo *et al.*, 2004; Da Silva *et al.*, 2006; Rahman *et al.*, 2008; Muhammad *et al.*, 2011). Thus through polyculture, farmers can utilize all the different zones in pond water due to different feeding habits of fish and their occupation of different niches and polyculture with *Cyprinus carpio*. In this way farmers can also significantly cut down the overall production cost maintaining the desired water quality (Wahab *et al.*, 2002; Milstein *et al.*, 2002, Alim *et al.*, 2005; Woynarovich, *et al.*, 2011). According to Milstein *et al.* (2003), the common carp as a bottom feeder fish produces a fertilizing effect through a food web that benefits the filter feeding fishes and reduces the application of organic and inorganic fertilizers in aquaculture practices. It grows rapidly with high protein diet and minimum feed coefficient and is considered as a target cultured fish, and plays a key role in pond management. It stimulates efficiency of liming and nutrient availability in the bottom of ponds, so the inclusion of common carp in polyculture is economical to farmers as it lowers the input and management costs and it also benefits the pond water ecosystem (Wahab *et al.*, 2002; Alim *et al.*, 2005, Abbas *et al.*, 2010). The production of fish pond depends on the vegetation, which is dependent on the nutrients in the ponds. It is not possible to increase the production of cultivated fish by giving them the greater quantities of natural food directly. Organic manures and chemical fertilizers can be used to increase the planktonic biomass, on which fish mainly feeds. It stimulates the growth of natural food by providing essential deficient elements, which are utilized by the phyto- and zooplanktons. Fertilization in fish farming is to improve water quality and to increase the variety and quantity of phytoplankton and zooplankton, which eventually leads to high fish yield and economic returns. Hence, the ultimate goal of fertilization is to achieve suitable environmental conditions for the production of natural food for fish, but in comparison with organic manure, fertilizers increase the level of primary productivity, abundance of algae, dissolved oxygen, pH and total phosphate (Afzal *et al.*, 2007; Jana *et al.*, 2001 Abbas *et al.*, 2010). Sustainable and successful freshwater fish culture on scientific basis principally depends upon the use of adequate, economically viable and environment friendly artificial feeds. Since the feed costs vary between 40 to 60% of the total managerial expenditure in fresh water fish culture system, provision of artificial feed increases the fish growth and production in the fertilized ponds and results in higher growth rates and yields than fertilization alone (Diana *et al.*, 1994). With a view of reducing feed input cost in aquacultural practices, it is necessary to develop better feeding strategies by incorporating plant based feed with animal protein based diets in feeding practices. Common carp fed with fish meal, rice bran, mustard oil cake

showed 1.5 and 2.1 times higher fish yield than in the treatments without supplementary feed (Rahman *et al.*, 2006). According to Azim *et al.* (2002) growth, specific growth rate of major carps were higher in fertilized ponds with the provision of supplemental feed than in control (fertilization alone). Nandeeshia *et al.* (2001) also noted that the specific growth rates, protein efficiency ratio as well as growth rate were more pronounced in animal and plant based diet as compared to animal based diet. Fertilization and supplemental feeding are the two important management measures adopted in the semi-intensive system of carp and Tilapia culture in Egypt. Several studies focus on the role of fertilizers in fish production (Garg and Bhatnagar, 2000; Dhawan and Kaur, 2002; Das *et al.*, 2005; Sayeed *et al.*, 2007; Bwala and Omoregie, 2009; Hussein, 2009; Ponce Palafox, 2010. Priyadarshini *et al.*, 2011) and of supplemental feed in systems receiving fertilizers (Aziz *et al.*, 2002; Virk and Saxena, 2003; Ahmed *et al.*, 2005; Waidbacher *et al.*, 2006; Elnady *et al.*, 2010). While supplemental feeding affects fish growth directly, fertilization contributes to growth via the planktonic natural food. In addition to acting as a food for fish, plankton perform other important functions in pond aquaculture: a net producer of dissolved oxygen, which is indispensable for fish growth (Teichert-Coddington and Green, 1993) and the most important sink of ammonia-nitrogen, which is excreted by the fish (Hargreaves, 1998; Jiménez-Montealegre, 2001). The FAO/AADCP Regional Expert Consultation has emphasized the need for a greater understanding of the role of natural food organisms in semi-intensive farming based on systems that optimize pond fertilization, in order to bring down the cost of fish production (NACA/FAO, 2000). The best way to reduce the cost of fish production is to minimize the use of supplemental food that can be best achieved by exploiting the synergetic interaction between natural food and supplemental feed. According to Moav *et al.* (1977), judicious organic manuring of fish ponds can eliminate the need for supplementary feeding. Increase in production by a given regime of supplementary feeding is of great economic importance, but is difficult to predict whether it is related to the amount of natural food available, the density of stocking or the range of other management variables. In the present work, an experiment has been conducted to evaluate the effect of supplementary feeding, manure and their combination in the monoculture of the common carp (*Cyprinus carpio*) fry and fingerlings carried out separately, to get an insight into their contribution to fish growth and production under different treatments.

## MATERIALS AND METHODS

**Description of the study area:** The two parallel experiments were conducted in twelve nursing earthen ponds, each with dimensions 50 x 20 x 1.0m (length x width x depth) located at Serow Fish Farm, National Institute of Oceanography And Fisheries, Dakahlia Governorate, Egypt. These ponds were firstly drained and cleaned, then supplied with drainage freshwater from El-Serow drainage canal to a depth of 0.7 m. The experimental period lasted for 4 months (120 days, initiated on first May till first September). Ponds were sun and air dried for three weeks. Inlets of ponds were properly screened with gauze of fine mesh to avoid the entry of intruder into or exit of fish from the ponds. Tube well was used as source of water. All ponds were watered up to a level of 0.7 m and this water level was maintained throughout the experimental period. They were fertilized with organic manure to stimulate the productivity of the ponds. After two weeks of fertilization, ponds were stocked with 5000 *Cyprinus carpio* fry. At the time of stocking fish were weighed and measured.

Fertilization of all these ponds was done daily with organic manure (poultry manure) 0.10g N /100g fish body weight except control.

**Feeds and feeding:** Common carp fry (mean wt. 0.67 g) in experiment one and fingerlings (mean wt. 3.2 g) in experiment two were stocked in the ponds 7 days after manuring, at 5 individuals/m<sup>2</sup>. Fish in F and M+F treatments were provided with fish meal-based pelleted feed (Table 1) twice daily at 5% of body weight. There were three experimental treatments and a control (T0). All the treatments including control had three replicates. The control did not receive any external input neither poultry manure nor supplementary feed while treatment 1(T1) received regular applications of organic manure (M), treatment 2(T2) was offered supplementary feed only (F) containing 30% protein (Islam, 2002), prepared (Rath, 2002) from different ingredients (Table 1) and T3 received both organic manure and supplementary feed (M+F) as mentioned in T2 above in triplicate. Supplementary feed was fed to fish daily 5% of their wet fish body weight. The amount of feed was increased fortnightly in proportionate to the weight increments. The feed was prepared using finelyground ingredients as per composition shown in Table 1.

They were mixed thoroughly with water to make a dough. The dough was then transferred to an aluminum container and steam cooked in a pressure cooker at 15 psi for 15 minutes. Vitamins and minerals mixture was mixed after cooling the dough. Pellets (2 mm diameter size) were prepared by a hand pelletizer and were dried in an oven at 40°C.

Table 1: Composition of formulated feed

Ingredient	(%)
Fish meal	30
Sunflower meal	30
Rice bran	10
Yellow corn	28
Vitamin <sup>1</sup> -mineral <sup>2</sup> Mixture*	2

1- each one kg of vitamin mixture contains: vitamin A 72000 IU; E 60 mg; B1 6 mg; B3 12000 IU; B6 9mg; B12 0.06mg; C 12mg; Pantothenic acid 60 mg; Nicotinic acid 120mg; Folic acid 6mg; Biotin 0.3 mg; choline chloride 3mg.

2- each one kg of mineral mixture contains: zinc sulfate heptahydrate 3.0g; cuprous chloride 0.10g; calcium phosphate monobasic 135.8 g; calcium lactate 327.0g; ferric citrate 29.7g; potassium phosphate dibasic anhydrous 239.8 g; sodium phosphate monobasic 87.2; sodium chloride 43.6 g; magnesium sulfate 12.75g; aluminum chloride anhydrous 0.15 g; potassium iodide 0.15 g; cobalt chloride 1.0g; sodium selenite 11mg and L-cellulose 132.25g.

After every thirty days, the stocked fish were captured randomly with nylon net and wet body weight and total length was measured and recorded and the feed quantity was readjusted based on the weight recorded at each sampling. Trial continued for four months. At the termination of experiment all the fish from different treatments (with all the three replicates) were harvested, weighed individually and yield calculated and measured to assess the performance of various inputs on specific growth rate (SGR) and net fish yield.

**Proximate composition of feed ingredients:** The highest values of protein, fat and ash were recorded in fish meal and the lowest values of protein and fat in sunflower meal were recorded and determined. Rice bran had the highest fiber content, while Yellow corn had the highest level of NFE. The feed contained 28.87% protein, 5.1% fat and 30.54% NFE (Table 2).

Table 2: Proximate compositions (%) of ingredients and feed

Parameter	Fish meal	sunflower meal	Rice bran	Yellow corn	Feed
Moisture	6.91±0.17	6.58±0.18	8.40±0.07	9.99±0.42	8.61±0.42
Crude protein	51.83±0.55	37.49±0.05	4.92±0.55	2.46±0.10	28.87±0.10
Fat	10.92±0.06	6.79±0.10	1.63±0.06	0.53±0.10	5.10±0.10
Ash	25.79±0.03	6.50±0.39	17.69±0.03	14.68±0.03	14.68±0.03
Crude fibre	1.90±0.15	10.80±0.21	31.80±0.15	3.60±0.01	12.20±0.01
Nitrogen-free extract <sup>1</sup>	2.65	31.84	35.56	81.68	30.54
Gross energy (kJ/g) <sup>2</sup>	16.41	16.59	7.86	14.81	13.76

1- Calculated by differences.

2- Estimated according to Jobiling (1983).

**Water quality analysis:** Water quality parameters were measured weekly according to **Boyd (1990 and 2000) and APHA (2000)**. Water quality samples were collected weekly from each pond manually from the middle of water column by putting a closed sample bottle and opened in the desired depth. This procedure was done in five different spots in each pond then samples were mixed in a plastic bucket and 1 liter sample was taken as a representative water sample of each pond. These samples were taken one week after fertilizer application. Analysis of water quality including temperature, pH, dissolved oxygen (DO), free carbon dioxide (CO<sub>2</sub>), total alkalinity, phosphate, ammonia, nitrate and nitrite were done at every week, collecting samples was done from the experimental ponds between 09.00 and 10.00hr. Water temperature, dissolved oxygen and pH were measured at 9.00h using a digital thermometer, and dissolved oxygen meter model Orion 835 A, while pH was measured with a digital pH meter model Acumen 25 meter.

**Qualitative and quantitative analysis of plankton:** Phytoplankton and zooplankton samples were collected and measured fortnightly for qualitative analysis by towing 15mm and 60 mm nets respectively. After collecting, zooplankton samples were preserved in LUGOL and phytoplankton in 4% formalin solution. Dry weight of plankton was also determined every fortnight by filtering 100 liters of water from each pond through a plankton net of 15 mm size and drying the filtrate in a hot-air oven at 80°C, till a constant weight was obtained. The quantitative estimation of total plankton was done by the "Direct census method" (Jhingran *et al.*, 1969).

**Proximate composition:** Proximate composition of feed ingredients, feed and fish carcass from experiment one was estimated. Fish carcass was obtained upon harvest by collecting five fish, each from the triplicate ponds and dried at 80°C to a constant weight. The dried carcass of each group was pooled together and ground. Moisture and ash contents were estimated according to AOAC (1995) methods. Crude protein, fat and fibre contents were analyzed using Kjeltex (Tecator, 1002 distilling unit), Soxtec (Tecator, 1043 extraction unit) and Fibretex (Tecator 1017 hot extractor) systems. Carbohydrate content was calculated as nitrogen free extract (NFE) by the difference method of Hastings (1976). The energy value of each ingredient as well as feed was obtained by multiplying protein, lipid and carbohydrate contents by factors 22.6, 38.9 and 17.2 respectively (Mayes, 1990) and expressed in kJ/g.

**Fish growth parameters, survival and production calculation:** After every one month, cultured fish species were captured randomly by using drag net from each experimental treatment and released back into their respective ponds after recording the data for wet body weight (WBW) and specific growth rate (SGR). After one month interval, on the basis of WBW, amount of organic fertilizer and supplementary feed added in fish ponds were determined for each treatment. Specific growth rate (SGR) was estimated by the following formula given by Dhawan and Kaur (2002).

$SGR = \frac{\ln(\text{Final wet body weight}) - \ln(\text{Initial wet body weight})}{\text{Time duration (days)}} \times 100$

**Survival rate and total fish production under different treatments:** At the end of the experiment, total harvested

SR (%) = % of live fish number at harvest.

Production (g) = Mean body weight (g) x Total number of viable fish at harvest.

**Statistical analysis:** Mean values of fish growth parameters at harvest, and carcass proximate composition were compared by one-way ANOVA. All plankton and water quality parameters were subjected to two-way ANOVA with treatment and sampling date as factors. When a main effect was significant, pair-wise comparison of treatment means was done by Duncan's multiple range test ( $P = 0.05$ ) (Duncan, 1955). All analyses were done using the ANOVA procedure of SAS program ver. 9.1 (SAS, 2005)

## RESULTS AND DISCUSSION

### Water quality analysis During the Experimental period:

Results presented in Table 3 revealed that average of water quality data variation for different water quality parameters of different treatments over the study period. Water quality parameters were within acceptable ranges for fish culture. There were no significant differences in water quality parameters between the different treatment ponds.

The values of water quality parameters monitored weekly ranged as follows. Water temperature: 27.96 to 28.16°C, pH: 8.43 to 8.58, dissolved oxygen: 6.77 to 8.85 mg/L, free carbon dioxide: 2.62 to 3.73 mg/L, total alkalinity ( $\text{CaCO}_3$ ): 139.20 to 151.29 mg/L, phosphate: 0.86 to 1.10  $\mu\text{g/L}$ , ammonia: 0.19 to 0.93  $\mu\text{g/L}$ , nitrate: 0.080 to 108  $\mu\text{g/L}$ , nitrite: 0.008 to 0.047  $\mu\text{g/L}$  (Table 3). Alkalinity and phosphate contents were significantly ( $P < 0.05$ ) higher in M + F treatment. pH, free carbon dioxide, ammonia, nitrate and nitrite values did not differ ( $P > 0.05$ ) between the treatments and the control (Table 3). All the water quality parameters showed difference significant ( $P < 0.05$ ) variation. The interaction effect of treatment and day was significant only for  $\text{CO}_2$  ( $P = 0.04$ ). DO, pH, alkalinity, nitrite and ammonia were the lowest on the first day of sampling.

Table 3: Water quality parameters (mean  $\pm$  S.E.) (Pooled data of the two experiments)

Treatment	Water Temp. (°C)	pH	Dissolved Oxygen (mg/L)	Free CO <sub>2</sub> (mg/L)	Alkalinity (mg/L)	Phosphate (mg/L)	Ammonia (mg/L)	Nitrate (mg/L)	Nitrite (mg/L)
Control (T1)	28.14 $\pm 0.39a$	8.46 $\pm 0.10a$	8.09 $\pm 0.34ab$	3.73 $\pm 0.49a$	139.20 $\pm 2.17c$	0.86 $\pm 0.08b$	0.37 $\pm 0.51a$	0.078 $\pm 0.21a$	0.039 $\pm 0.70a$
Feed (T2)	28.16 $\pm 0.38a$	8.49 $\pm 0.09a$	6.77 $\pm 0.46c$	3.02 $\pm 0.47a$	141.67 $\pm 2.53b$	0.96 $\pm 0.08ab$	0.93 $\pm 0.42a$	0.108 $\pm 0.21a$	0.047 $\pm 0.088a$
Manure (T3)	28.16 $\pm 0.46a$	8.58 $\pm 0.07a$	7.18 $\pm 0.48bc$	2.62 $\pm 0.51a$	141.46 $\pm 2.00b$	1.03 $\pm 0.08ab$	0.19 $\pm 0.44a$	0.080 $\pm 0.18a$	0.008 $\pm 0.63a$
Manure+ Feed (T4)	27.96 $\pm 0.38a$	8.43 $\pm 0.08a$	8.85 $\pm 0.17a$	2.73 $\pm 0.53a$	151.29 $\pm 2.06a$	1.10 $\pm 0.09a$	0.34 $\pm 0.41a$	0.084 $\pm 0.17a$	0.024 $\pm 0.60a$

Different superscripts for values in the same column indicate significant ( $P \leq 0.05$ )

Table (3) shows that the average values of water temperature and pH in the different treatments were similar during the experimental period and varied within a narrow range. pH was in the alkaline range throughout the experimental duration, indicating favorable conditions for biological production. This range was beneficial to fish culture in agreement with results of Hussein (2009) and Muhammad *et al.*,

(2011). Jhingran (1995) who observed that carps thrive well in the temperature range of 18.3°C to 37.8°C. According to Farmanfarmain and Moore (1979), aquatic organisms can tolerate a wider range of temperatures, provided that fluctuations are not severe, sudden and of long duration. DO was high throughout the experimental duration and fluctuated between 6.77 (F) and 8.85 mg/L (M+F treatment). This reflects higher photosynthetic activity in manure plus feed treatment. Dissolved oxygen levels improved due to photosynthesis, while ammonia levels were reduced through assimilation by phytoplankton (Boyd, 1990).

Generally, cyprinids are capable of tolerating low oxygen levels of 3 mg/L (Huet, 1972). The highest value of total alkalinity was recorded in the (T4) treatment (151.29 mg/L) and the lowest occurred in control T1 (139.20 mg/L). Total alkalinity was significantly greater where organic fertilizer and feeds were applied to ponds (Kumar *et al.*, 2005). Alkalinity increases with organic fertilization because bacterially generated CO<sub>2</sub> from manure decomposition dissolves calcium and magnesium carbonate in pond water into calcium and magnesium bicarbonate (Boyd, 1990). Diana *et al.*, (1994) reported that fertilization alone led to low alkalinity. Phosphorus was significantly higher ( $P < 0.05$ ) in M+F treatment in comparison with the control (Table 3). The higher phosphorus concentration may be associated with the increase in phosphorus produced during the decomposition of organic fertilizer and also from the feed through fish excrete. Both soluble organic phosphorus and orthophosphate are released during the process of organic fertilizer decomposition under aerobic conditions (Wudtish and Boyd, 2005; Hussein, 2009). Higher concentrations of ammonia nitrogen are often noticed in fish culture ponds (Edwards, 2008). However, the values of ammonia recorded in the present experiments were low (Table 3). Sugiyama and Kawai (1978) reported that the higher concentrations of dissolved oxygen decreases ammonia level through oxidation. These low concentrations of ammonia may be attributed to ammonia utilization by phytoplankton (Boyd, 1998) or oxidation of ammonia nitrite especially in high dissolved oxygen level conditions (Boyd, 2000). Total ammonia nitrogen fluctuated throughout experiment but remained below 1 mg/L and at the pH levels observed; unionized ammonia probably did not adversely affect fish performance. Major water quality parameters measured during the study remained in the favorable range for fish culture (Boyd, 1990). Comparable results were obtained by Lawson (1995). All ponds were within acceptable range of water quality parameters during the study. The Use of organic fertilizers and supplementary feed improved water quality through stimulation of natural food, mainly phytoplankton and zooplankton, suitable for the filter feeding carp species. Organic fertilizers acts as an energy source for bacterial growth, but the aerobic decomposition of organic matter by bacteria is an important drain of oxygen supplies in ponds (Boyd, 1982).

Values of water temperature, pH and DO in the diurnal samples showed no effect of treatments. The increase in the values of these parameters with the progress of day and decrease with the progress of night can be related to the presence and absence of light which affects temperature and also dissolution of oxygen in pond water. Further photosynthesis during day time is responsible for the higher DO values, whereas consumption of DO by plankton reduced night time DO. Similarly, pH variations can also be correlated with photosynthetic activity.

#### **Plankton biomass**

Table 4 quantifies the planktonic species encountered in the tank water on the sampling days. Among phytoplankton, Chlorophyceae comprised 15 genera, the major ones being *Microspora*, *Volvox* and *Scenedesmus*. Cyanophyceae was represented by 3 genera, *Microcystis* and *Anabaena* being dominant. Chrysophyceae,

Bacillariophyceae and Dinophyceae were represented by one genus each. *Diaptomus*, *Cyclops*, *Moina* and *Keratella* were the zooplankton species encountered. In addition, nauplii and insect eggs were also found in good numbers.

Table 4: Abundance of phytoplankton species in pond water (cells /L  $\pm$  S.E.) during the experimental period.\* (Pooled data of the two experiments)

Group/Genus	Control T1	Feed T2	Manure T3	Manure+Feed T4
<b>PHYTOPLANKTON</b>				
<b><i>Chlorophyceae:</i></b>				
<i>Gleocapsa</i>	0.04 $\pm$ 0.04	1.12 $\pm$ 0.46	0.40 $\pm$ 0.15	0 0
<i>Golenkinia</i>	3.35 $\pm$ 0.98	7.14 $\pm$ 2.12	11.52 $\pm$ 6.51	3.08 $\pm$ 0.93
<i>Hydrodictyon</i>	0 0	0.36 $\pm$ 0.16	0.13 $\pm$ 0.10	0.27 $\pm$ 0.10
<i>Menoidium</i>	0 0	0 0	0.40 $\pm$ 0.21	0.54 $\pm$ 0.33
<i>Ankistrodesmus</i>	15.09 $\pm$ 3.10	11.79 $\pm$ 2.77	16.74 $\pm$ 4.03	16.65 $\pm$ 5.98
<i>Chlorococccum</i>	10.13 $\pm$ 2.07	17.54 $\pm$ 5.92	12.90 $\pm$ 2.52	31.16 $\pm$ 15.55
<i>Closterium</i>	3.39 $\pm$ 0.53	2.32 $\pm$ 0.61	2.72 $\pm$ 0.53	0.45 $\pm$ 0.18
<i>Staurastrum</i>	22.95 $\pm$ 26.07	32.37 $\pm$ 4.53	28.44 $\pm$ 4.10	60.00 $\pm$ 26.70
<i>Microspora</i>	148.68 $\pm$ 203.28	330.67 $\pm$ 95.78	226.88 $\pm$ 57.55	484.64 $\pm$ 109.31
<i>Pediastrum</i>	7.01 $\pm$ 1.01	24.20 $\pm$ 5.82	3.17 $\pm$ 0.61	10.31 $\pm$ 3.13
<i>Phytoconis</i>	21.12 $\pm$ 5.34	21.12 $\pm$ 4.97	9.38 $\pm$ 2.56	22.99 $\pm$ 3.40
<i>Scenedesmus</i>	59.20 $\pm$ 24.73	37.81 $\pm$ 9.83	75.85 $\pm$ 20.18	71.92 $\pm$ 24.60
<i>Selenastrum</i>	67.41 $\pm$ 18.72	25.94 $\pm$ 4.61	78.08 $\pm$ 28.80	44.82 $\pm$ 10.42
<i>Tetraedron</i>	35.22 $\pm$ 10.47	18.84 $\pm$ 7.37	7.05 $\pm$ 2.34	9.46 $\pm$ 1.44
<i>Volvox</i>	89.87 $\pm$ 23.83	14.64 $\pm$ 2.46	25.54 $\pm$ 7.11	49.20 $\pm$ 15.95
<b><i>Chrysophyceae:</i></b>				
<i>Chrysophyxis</i>	3.66 $\pm$ 1.16	2.01 $\pm$ 0.43	1.47 $\pm$ 0.34	1.38 $\pm$ 0.35
<b><i>Bacillariophyceae :</i></b>				
<i>Anomoeoneis</i>	5.71 $\pm$ 1.58	10.18 $\pm$ 3.26	3.13 $\pm$ 0.62	3.17 $\pm$ 0.68
<b><i>Dinophyceae :</i></b>				
<i>Monomastix</i>	1.29 $\pm$ 0.38	0.45 $\pm$ 0.18	2.28 $\pm$ 0.61	1.21 $\pm$ 0.31
<b><i>Cyanophyceae :</i></b>				
<i>Anabaena</i>	541.61 $\pm$ 205.71	1654.73 $\pm$ 366.36	1550.27 $\pm$ 325.36	2213.26 $\pm$ 314.90
<i>Merismopedia</i>	92.14 $\pm$ 87.44	1.43 $\pm$ 0.33	5.40 $\pm$ 1.31	3.30 $\pm$ 0.85
<i>Microcystis</i>	1099.33 $\pm$ 256.83	2117.72 $\pm$ 317.21	2155.40 $\pm$ 360.98	2139.46 $\pm$ 340.12
<b>Total Phytoplankton</b>	2227.21 $\pm$ 513.44	4332.37 $\pm$ 472.86	4217.14 $\pm$ 628.81	5167.28 $\pm$ 530.43
<b>ZOOPLANKTON:</b>				
<i>Diaptomus</i>	40.18 $\pm$ 6.26	52.99 $\pm$ 12.06	77.23 $\pm$ 15.56	153.70 $\pm$ 87.27
<i>Cyclops</i>	23.44 $\pm$ 3.98	19.24 $\pm$ 2.45	44.46 $\pm$ 9.81	23.30 $\pm$ 3.62
<i>Moina</i>	13.66 $\pm$ 2.98	7.54 $\pm$ 1.27	17.37 $\pm$ 8.53	12.86 $\pm$ 2.16
<i>Keratella</i>	95.49 $\pm$ 48.01	287.54 $\pm$ 159.98	39.06 $\pm$ 12.98	456.25 $\pm$ 220.85
Insect eggs	34.37 $\pm$ 4.40	48.70 $\pm$ 8.61	74.06 $\pm$ 16.02	55.80 $\pm$ 12.32
Nauplius	106.70 $\pm$ 73.96	28.17 $\pm$ 8.57	386.61 $\pm$ 248.60	123.79 $\pm$ 83.01
<b>Total Zooplankton</b>	313.84 $\pm$ 90.71	444.20 $\pm$ 165.22	638.79 $\pm$ 257.65	825.71 $\pm$ 258.11

\*Numbers are means of 14 samplings. Numbers in italics are standard errors.

The overall phytoplankton population was the highest under M+F treatment (5167.28 cells /L) and the lowest in control (2227.21 cells /L). The number of green algae was lower as compared to blue-green algae in all the ponds (Table 4). The ratio between cyanophyceae and chlorophyceae was the lowest in control and highest in manure+feed (M+F) ponds. Density of phytoplankton and zooplankton (no/L) was also significantly lowest in controlponds (Table 5). The density of both phyto and zooplankton was the lowest ( $P < 0.05$ ) on the first day of sampling and highest on the 120<sup>th</sup> day. The interaction effect of treatment and day was significant for *Staurastrum* ( $P = 0.0197$ ) and nauplii ( $P = 0.007$ ).



Dry weight of phytoplankton was highest under feed (F) treatment and lowest in control. The values in manure (M) and M + F treatments were almost similar (Table 5), whereas the highest dry weight of zooplankton was observed under M + F (2.15 mg/L) treatment, followed by manure (1.69 mg/L), feed (1.65 mg/L) and control (0.77 mg/L). Significant ( $P<0.05$ ) variation in both phyto and zooplankton dry weight was recorded over the experimental period, both being the lowest on zero day and highest on 120<sup>th</sup> day.

Significant ( $P<0.05$ ) variation in both phytoplankton and zooplankton dry weight was recorded with respect to study period. The interaction effect of treatment and day was also significant for both phytoplankton and zooplankton. The number of green algae was lower as compared to blue-green algae in all the experimental ponds. In fish ponds, blue green algae constitute the greater part of phytoplankton; higher alkalinity, nitrate, ammonia and phosphate favour the multiplication of cyanophyceae (Padmavathi and Veeraiah, 2009). Kulkarni (1992), who studied the effect of distillery waste on plankton and fish production, reported a significant ( $P<0.05$ ) correlation between phosphorus level and blue green algae production.

Table 5: Plankton biomass and density ( $\pm$  S.E.) in experimental ponds (Pooled data of the two experiments)

Treatment	Dry weight (mg/L)		Density (no/L)	
	Phytoplankton	Zooplankton	Phytoplankton	Zooplankton
Control	0.77 $\pm$ 0.14b	0.77 $\pm$ 0.24c	2227.21 $\pm$ 513.44b	313.84 $\pm$ 90.71a
Feed	1.98 $\pm$ 0.27a	1.65 $\pm$ 0.15b	4332.37 $\pm$ 472.86a	444.20 $\pm$ 165.22a
Manure	1.73 $\pm$ 0.24a	1.69 $\pm$ 0.17b	4217.14 $\pm$ 628.81a	638.79 $\pm$ 257.66a
Manure $\pm$ Feed	1.70 $\pm$ 0.25a	2.15 $\pm$ 0.24a	5167.28 $\pm$ 530.43a	825.71 $\pm$ 258.11a

Different superscripts for values in the same column indicate significant ( $P<0.05$ ) difference.

**Rahman et al. (2008)** reported that the common carp increased bio-available N and P in the water column and plankton where availability was positively correlated with bio-available N and P. The relationship between provision of manure/feed and plankton biomass observed in the present study can be related to the nutrient input. In addition, fish excretion would have contributed to the level of N and P in pond water, particularly towards the later part of the experiment.

#### **Fish growth, survival and production**

The final weight and length of fish in experiments one and two are given in Tables (6 and 7). The highest final weight was observed in T4 treatment in both the experiments. Growth was similar in F and M treatments in experiment one, but significantly ( $P<0.05$ ) different in experiment two.

Table 6: Growth parameters (average  $\pm$  S.E.) of common carp fry under different treatments (Experiment one)

Parameter	Control T1	Feed T2	Manure T3	Manure+Feed T4
Final weight (g)/fish	18.16 $\pm$ 1.15b	35.86 $\pm$ 0.22a	36.84 $\pm$ 4.50a	43.43 $\pm$ 2.02a
Increment in growth over control (%)	-	97.46	102.86	139.15
Final length (cm)	11.41 $\pm$ 0.7b	14.05 $\pm$ 0.20a	12.78 $\pm$ 0.50ab	13.17 $\pm$ 0.10a
SGR (%)	2.75 $\pm$ 0.09b	3.32 $\pm$ 0.02a	3.32 $\pm$ 0.18a	3.47 $\pm$ 0.07a
Survival (%)	57.58 $\pm$ 7.57a	61.20 $\pm$ 14.2a	52.12 $\pm$ 12.76a	63.03 $\pm$ 2.09a
Production (kg/pond/ 4 months)	1725.39 $\pm$ 105.72c	3506.40 $\pm$ 91.85b	3059.04 $\pm$ 38.95b	4524.63 $\pm$ 173.35a
Increment in production over control (%)	-	103.22	77.30	162.24

Different superscripts for values in the same column indicate significant ( $P<0.05$ ) difference.

Fish growth was significantly ( $P<0.05$ ) poor in the control in both the experiments. SGR values followed the trend of fish growth at harvest.

The overall survival varied from 52.12% in T3 treatment to 63.03% in T4 treatment in experiment one, while it was in the range of 61.67% (T4) to 82.78% (T3) in experiment two. However, difference in survival among the control and the treatments was not significant ( $P>0.05$ ) in experiment one. Net production in this experiment varied from 1725.39 (control) to 4524.63 g/pond /120 days (T4 treatment).

In T2 and T3 treatments production was nearly equal, being significantly ( $P<0.05$ ) higher in the control and lower in T4 treatment (Table 6). In experiment two, the net production was higher compared to experiment one (2518.20 to 6927.03 g/pond/120 days) and varied significantly between the control and all treatments (Table 7), while manuring, individually and in combination, improved the growth of fish significantly ( $P<0.05$ ) in both the experiments (Tables 6 and 7). The highest final weight was recorded in M+F (T4) treatment. Specific growth rate followed the growth trend in both the experiments, while there was no difference in growth of fish between feed (F T2) and (M T3) treatments in experiment one, as it differed significantly ( $P<0.05$ ) in experiment two. Growth under F, M and M + F treatment in experiment one works out to 97.46%, 102.86%, and 139.15% higher respectively over the control. The corresponding values in experiment two are 152%, 36.20% and 184.78%. It is clear that feed treatments (F and M+F) had greater impact on the growth of fingerlings as compared to fry; fingerlings are better equipped in terms of mouth size and digestive enzymes to accept and utilize pelleted diet (Woynarovich, *et al.* 2011). Further, a comparison of the final weights of fish from experiment one with that of experiment two, points out to the difference in growth rate due to life stage. The increase in weight of control fish is only 1.62 g, whereas under manure

Table 7: Growth parameters (average  $\pm$  S.E.) of common carp fingerlings under different treatments (Experiment two)

Parameter	Control T1	Feed T2	Manure T3	Manure+Feed T4
Initial weight(g)/fish	3.21 $\pm$ 0.41a	3.19 $\pm$ 0.41a	3.22 $\pm$ 0.41a	3.18 $\pm$ 0.41a
Final weight (g)/fish	19.78 $\pm$ 0.82d	49.87 $\pm$ 1.62b	26.94 $\pm$ 1.48c	56.33 $\pm$ 2.75a
Increment in growth over control (%)	-	152.12	36.20	184.78
Initial length (cm)	-	5.23 $\pm$ 0.21a	5.23 $\pm$ 0.21a	5.22 $\pm$ 0.21a
Final length (cm)	5.20 $\pm$ 0.21a	5.23 $\pm$ 0.21a	5.23 $\pm$ 0.21a	5.22 $\pm$ 0.21a
SGR (%)	9.50 $\pm$ 1.14c	13.70 $\pm$ 1.01a	12.25 $\pm$ 0.50b	14.07 $\pm$ 0.81a
Survival (%)	1.52 $\pm$ 0.03b	2.29 $\pm$ 0.03a	1.77 $\pm$ 0.04b	2.62 $\pm$ 0.27a
Production (kg/pond/4 months)	70.55 $\pm$ 2.94ab	68.89 $\pm$ 6.83b	82.78 $\pm$ 0.55a	61.67 $\pm$ 2.55b
Increment in production over control (%)	2518.20 $\pm$ 63.85d	5540.01 $\pm$ 107.22b	4015.11 $\pm$ 70.33c	6927.03 $\pm$ 141.09a
	-	119.99	59.44	175.08

Different superscripts for values in the same column indicate significant ( $P<0.05$ ) difference.

treatment (M), there is a reduction of 9.9 g. As against this, in fed treatments there is an increase of 14.01g (F) and 12.9 g (M+F) (Tables 6 and 7). This result showed that the nutrient requirement of fingerlings is not satiated by natural food alone, contrary to that in the case of fry. Boyd (1990) reported a strong positive correlation between fish growth and primary productivity in fertilized ponds without supplementary feeding. Natural food is nutritive and contains 51.1% protein, 27.3% carbohydrate and 7.7% fat, while the calorific value ranges from 6.7 to 23.8 kJ/g (De Silva and Anderson, 1995). It is possible that as the fish grows bigger, it prefers artificial diet when available. Rahman *et al.* (2008) observed that common carp growth, in polyculture with rohu, *Labeo rohita*, was higher in the presence of artificial feed and

negatively correlated with natural food availability. They also recorded higher ingestion of benthic macroinvertebrates, copepods and rotifers, and a lower ingestion of phytoplankton by common carp.

In the ponds receiving no supplementary feed, the growth obtained is entirely attributable to the natural food. This applies to the control as well as manure (M) treatment. In experiment one, growth of fish in M treatment was double that of control, reflecting the ability of common carp fry to extensively feed on the available natural food and convert it into flesh. Though the control ponds did not receive any nutrient input during the experiment. The soil bottom of all ponds used in the present study contained some nutrients accumulated from earlier trials; the effect of these could be considered as equal under all treatments. Common carp as a bottom feeding fish enhances the availability of nutrients to phytoplankton through stirring of the mud bottom (Milstein *et al.*, 2002). Ritvo *et al.* (2004) demonstrated that common carp by perturbations results in appreciable mixing of the sediment; this mixing would bring out nutrients into circulation, facilitating natural food production.

The difference in survival of fish in the control and treatment ponds was not significant ( $P>0.05$ ) in the first experiment, whereas feed treatments (F and M+F) recorded lower survival ( $P<0.05$ ) in the second experiment. This could be due to some natural mortality of fish in ponds of the two treatments, since water quality was similar in all treatments, but for higher alkalinity and phosphate levels in M+F treatment. Gross fish production was influenced both by fish weight and survival. Production was the highest in M+F treatment in both the experiments. In experiment one, production was comparable ( $P>0.05$ ) in F and M treatments. The increment in gross fish production over the control was 103.22% in F, 77.30% in M and 162.34% in M+F treatments (Table 6). In experiment two, production was significantly ( $P<0.05$ ) higher in the feed treatment (F) compared to the manure (M) treatment, again indicating the significance of feeding in the case of fingerlings. The corresponding figures of increment for experiment two are 119.99%, 59.44% and 175.08% (Table 7). Abbas *et al.* (2010) and Priyadarshini1 *et al.* (2011) reported highest gross production of carps in the treatment with the combination of organic and inorganic fertilizers and supplementary feeding, compared to combinations of any two of these.

### Proximate composition

The proximate composition of fish carcass from experiment one is shown in Table (8). Crude protein, fat and ash contents were significantly ( $P<0.05$ ) higher in the 3 treatments compared to the control. No difference was found between the crude protein content of F and M treatments and fat content of F, M and M+F treatments.

Table 8: Proximate composition of fish (% wet weight)

Parameter	Control T1	Feed T2	Manure T3	Manure+Feed T4
Moisture	73.50±2.60a	70.19±2.03a	68.90±2.99a	70.72±1.71a
Crude protein	17.21±0.08c	19.20±0.05b	19.27±0.06b	21.60±0.07a
Fat	1.08±0.06b	1.35±0.02a	1.31±0.11a	1.36±0.04a
Ash	5.78±0.02c	6.16±0.01b	6.54±0.01b	6.13±0.03a

Different superscripts for values in the same column indicate significant ( $P<0.05$ ) difference.

Initial weight and length of fry were 0.67±0.06 g and 2.32±0.11 cm respectively.

Proximate analysis of fish carcass revealed that the treatments affected crude protein and fat, both being lowest in control and highest in M + F(T4) treatment. However, there was no difference in moisture level among the treatments and control (Table 8). This is indicative of protein accretion and true growth involving an increase

in the structural tissue such as muscle and various organs (Fafioye *et al.*, 2005). The type of feed ingested and their nutritional quality is known to be one of the main factors affecting fish carcass composition (Reinitz and Hitzel, 1980; Priyadarshini *et al.* 2011).

The results obtained in this study clearly point out to the importance of natural food in fish culture. Growth of fish in experiment one indicates similar potential of poultry manure and the feed provided in inducing growth of common carp fry. In contrast, fish growth in experiment two was significantly better under fed treatments (F and M+F). This shows that nutritional requirement of common carp fingerlings is not fully met by natural food alone, contrary to that of fry. The findings can be used in developing feeding strategy for fish at different life stages during culture.

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### Arabic Summary

تأثير التغذية، والروث، والجمع بينهما على نمو يرقات وأصبعيات سمك المبروك العادي (سيبرينس كاريبو)

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في هذا البحث تم إجراء تجربتين، كل واحدة مدتها 120 يوم، في 12 حوض حضانة تربية، أبعاد كل حوض  $1.0 \times 20 \times 50$  متر في مزرعة أسماك السرورب المعهد القومي لعلوم البحار والمصايد، محافظة الدقهلية، مصر، وتم استخدام يرقات وإصبعيات سمك المبروك العادي (*Cyprinus carpio*)، وذلك بهدف دراسة تأثير المغذيات (الأعلاف، والسماذ، والجمع بينهما) في مراحل النمو المختلفة من الأسماك. تتألف المعاملات من الكنترول (C T1) و معاملة الأعلاف فقط (F T2)، ومعاملة السماذ العضوى فقط (MT3) بالإضافة إلى معاملة السماذ العضوى + الأعلاف (M + F T4). تم تطبيق سماذ روث الدواجن في جرعات مقسمة إلى أحواض المعاملات (M + F و M). كانت اليرقات مخزنة (تجربة رقم واحد) والأصبعيات (تجربة رقم 2) وكان متوسط الوزن الأبتدائي 0.67 و 3.2 جرام على التوالي، وبعد سبعة أيام من تطبيق السماذ الأولي تم التخزين في الأحواض بمعدل 5000 سمكة / حوض. وقدمت للأسماك في المعاملات M + F و F وجبة غذاء مكعبات مرة واحدة يوميا في الصباح بمعدل 5% من وزن الجسم. كانت معظم الأجناس السائدة من العوالق النباتية هي *Microcystis*، *Anabaena* و *Microspora*، في حين كانت العوالق الحيوانية هي *Keratella* واليرقات. وكانت الكتلة الحية في المعاملة (F + M) أعلى العوالق ( $P < 0.05$ ). وسجلت معنوية ( $P < 0.05$ ) التباين في الوزن الجاف فيما يتعلق بفترة الدراسة في كلا الكائنات الحية النباتية والحيوانية. كلا من التغذية والسماذ، فرادى أو مجتمعة، حسنت بشكل ملحوظ ( $P < 0.05$ ) على نمو الأسماك. وسجلت أعلى معدل نمو محدد، ووزن الأسماك النهائي، والإنتاج الإجمالي في المعامل (M + F) على حد سواء. وكان الاختلاف في البقاء على قيد الحياة بين الكنترول والمعاملات ليس كبيرا في التجربة الأولى ( $P > 0.05$ )، في حين المعاملات (M + F و F) سجلت أدنى بقاء ( $P < 0.05$ ) في التجربة الثانية. وكان الإنتاج السمكي غير ملحوظ بالمقارنة في معاملات الأعلاف (F) والروث (M) ( $P > 0.05$ ) في تجربة الأولى، بل كان معنويا ( $P < 0.05$ ) ومختلفة في التجربة الثانية. وكانت الزيادة في إجمالي الإنتاج السمكي للكنترول 103.22 و 119.99% وفي العلف، 77.30 و 59.44% وفي السماذ 162.34 و 175.08%. وفي المعاملة (سماذ + علف) على التوالي. وكانت محتويات الذبحة من البروتين والدهون والرماد عالية ( $P < 0.05$ ) وكانت أعلى في المعاملات الثلاثة بالمقارنة مع الكنترول.