Egyptian Journal of Aquatic Biology & Fisheries Zoology Department, Faculty of Science, Ain Shams University, Cairo, Egypt. ISSN 1110 - 6131 Vol. 22(4): 49- 59 (2018) ejabf.journals.ekb.eg



Assessment of polycyclic aromatic hydrocarbons in surface sediments and some fish species from the Gulf of Suez, Egypt

Alaa M. Younis¹; Yosry A. Soliman²; Eman M. Elkady² and Mohamed H. El-Naggar^{2,3}

- 1- Department of Aquatic Environment, Faculty of Fish Resources, Suez University, Suez, Egypt.
- 2- Marine Chemistry Lab., National Institute of Oceanography & Fisheries, Suez, Egypt.
- 3- Department of Chemistry, Faculty of Science, University of Sharjah 27272, UAE.

Corresponding author's email:<u>ala_den@yahoo.com</u>

ARTICLE INFO

Article History: Received: July 2, 2018 Accepted: Aug.6, 2018 Available online: Aug. 2018

Keywords: Hydrocarbons PAHs Suez Gulf Sediments Fish

ABSTRACT

The present study evaluated the origin and distribution of sixteen polycyclic aromatic hydrocarbons (PAHs) included by the United States Environmental Protection Agency (US EPA) within the priority pollutants were considered in the surface sediment as well as eleven fish species of Suez Gulf, Egypt. Sediment samples were collected at 0.5 and 10 meter depth from ten stations in addition to eleven fish species (Liza subviridis, Parupeneus forsskali, Upeneus japonicus, Lethrinus nebulosus, Stephanolepis diaspros, Epinephelus areolatu, Sphyraena chrysotaenia, Pomadasys stridens, Trachurus indicus, Saurida undosquamis, Engyprospon sp.). The concentrations of PAHs were measured by gas chromatography (GC) equipped with FID detector. The total concentration \sum 16 PAHs ranged from 1667.02 to 2671.27 ng/g with the highest levels recorded at stations IV, VII and X (Ras Gharib, Aion Mousa and Abu Zanima). While the total concentration $\sum 16$ PAHs in collected fish species ranged from 621 to 4207 ng/g wet weight with the highest Σ 16 PAHs found in species Saurida undosquamis, while the lowest found in the species Stephanolepis diaspros. The high molecular weight PAHs (HPAHs) were predominant if compared with the low molecular weight PAHs (LPAHs). With LPAHs/HPAHs values less than one for all studied species indicating anthropogenic origin of PAHs in fish. Furthermore, the study of the PAHs diagnostic ratios suggests the predominance of pyrolytic origin for the studied PAHs.

INTRODUCTION

Increasing developments of industrial activities near coastal areas have caused more concerns regarding risks to human health resulting of environmental pollution as well as seafood contamination (Shreadah *et al.*, 2006a & b; Younis & Nafea 2012; Younis *et al.*, 2014; El Zokm *et al.*, 2015; Okbah *et al.*, 2015; Younis *et al.*; 2016)

Polycyclic aromatic hydrocarbons (PAHs) are ubiquitous environmental organic contaminants. PAHs are well recognized for carcinogenic and mutagenic effects. PAHs are relatively water insoluble and highly lipophilic.

ELSEVIER DOAJ





Therefore, they can easily accumulate in fatty tissues of many marine species. Sources of PAHs could be anthropogenic or natural mainly because of incomplete combustion of petroleum as well as fossil fuel (Liang *et al.* 2007)

Suez Gulf was known to be one of the most important fishery grounds in Egypt. It is extends for about 250 km from Suez city in the north to Shadwan island in the south and it is relatively shallow with a mean depth of about 45 m while its width fluctuates between 20 and 40 km. As we know, fish are the most important aquatic products of Suez gulf cities. Lovers of seafood from all walks of life can be found surfing through the colorful variety of meat that offered from Red Sea (Said *et al.* 2006; Younis *et al.*, 2015). In the recent years, the Egyptian government has constructed many projects around the northwestern of Suez gulf area through the planning of development for future economic (Snousy *et al.*, 2016). These projects such as petroleum refining industries, shipyard, marinas, power stations, fertilizer, chemical industries, pipelines and establish the commercial harbors close to the industrial zone. Therefore, the Gulf of Suez coastal water exposed daily to high loads of oil wastes as well as various other anthropogenic discharges from the surrounding industrial areas (Nemr *et al.*, 2006; Snousy *et al.*, 2016).

Fish is an important source for protein, vitamins and unsaturated fatty acids. It is estimated that by 2020, the consumption of fish in some developing countries will rise by about 57% (Delgado *et al.* 2003). This high request for fish needs a sustainable supply of fish products. However, bad practices such as overfishing, discharge of agricultural and industrial waste in lakes and oceans not only decreases the fish stock but also leads to a health hazard due to consumption of contaminated fish. The determination of PAHs in different fish species can give useful information about quality of food supply. Therefore, regular monitoring of PAHs in fish and their environment is highly recommended. In this study the level of 16 priority pollutants PAHs recommended by U.S. EPA were determined in surface sediments and eleven fish species to assess the extent, distribution and source of PAHs contamination in the gulf of Suez.

MATERIALS AND METHODS

Sediments samples

Sediment samples were collected from ten stations; (I) Adabia (II) El-Sokhna (III) Zafrana (IV) Ras Gharib (V) Ras Shokar (VI) Jemsha (VII) Eion Mousa (VIII) Ras Sader (IX) Hamamat Pharons (X) Abu Zanima by van deer grab. Dried samples (10-20 g) were extracted in a Soxhlet-apparatus with dichloromethane. The cycle of siphonation was around 30 min with repetition for at least 10 times. As the process of soxhlet extraction was completed, the solvent was evaporated to about 1 ml using the rotary evaporator. Activated copper powder was used to remove sulfur compounds from the extract (UNEP, 1991).

The final extracted volume (1 ml) of sediment were injected in the Gas chromatography (Hewlett Packard, 5890 series II), with a capillary column, (25m length x 0.2 mm i.d x 0.5 μ m thickness), Ultra-1, coated with 100% dimethyl polysiloxane; non-polar. Nitrogen gas was used as the carrier gas at 4 ml/min flow rate. The temperature-programming rate was 8°C/min from initial hold of 50°C to a final hold of 290°C. The blank used for the GC analysis was a group of standard PAHs mixture including [naphthalene, acenahthylene, acenaphthene, fluorene, phenanthrene, anthracene, fluoranthene, pyrene, benzo (a) anthracene, chrysene, benzo (a) fluoranthene, benzo (k) fluoranthene, benzo (a) pyrene, dibenzo (a,h)

anthracene, benzo (ghi) perylene and indeno (1,2,3-cd) pyrene)] each of which has a concentration of 10 ppm dissolved in n-hexane.



Fig. 1: Sampling locations of surface sediment in the area of study

Fish samples

Eleven fish species (*Liza subviridis, Parupeneus forsskali, Upeneus japonicus, Lethrinus nebulosus, Stephanolepis diaspros, Epinephelus areolatus, Sphyraena chrysotaenia, Pomadasys stridens, Trachurus indicus, Saurida undosquamis, Engyprospon sp.*) were collected from different location of Suez gulf in summer of 2016. The fish samples were individually wrapped in previously hexane cleaned aluminium foil, The transferred to the laboratory and frozen (-20°C) until analysis. An identified number of each organism species were collected and the filet of each fish was homogenized to obtain a representative sample for each species of fish before the analysis. Fish muscle was obtained after cleaning the fish with ethanol, and removing the skin. The samples were analyzed for polycyclic aromatic hydrocarbons according to USEPA (1994).

Extraction and cleanup

All chemicals used for gas chromatographic analyses were chromatography grade and purchased from Merck. Exactly 10 g of fish muscle from each sample was treated with 2 g of anhydrous sodium sulfate and the mixture was blended at high speed for 5 min. Then the mixture was extracted with 250 ml of n-hexane in a soxhlet extractor for 8 h and then re-extracted for 8 h into 250 ml of methylene chloride. The mixture was filtered and the tissue was extracted twice more. Organic solvent fractions were combined and filtered through filter paper with 1 g anhydrous sodium sulfate and concentrated down using a rotary evaporator at 30 °C. The extract was transferred to a round bottom flask, then was saponified the lipids, using dark glass vials, with 10 mL of 1 M KOH in an ethanol solution for 3 h.

Clean-up was an fractionation achieved by flowing the concentrated extract through alumina/silica column. Aromatic hydrocarbons were eluted with 30 ml of a mixture of methylene chloride and n-hexane (50:50) (v/v). The volume of the eluted fraction was reduced to a volume of 1 ml by concentration under a gentle stream of nitrogen and analyzed by a gas liquid chromatography equipped with a flame ionization detector GC/FID.

Analytical Quality Controls

In order to study the assure recovery efficiency, three analyses were conducted on PAH standard reference materials (SRM 2974) provided by national institute of standards and technology (NIST, USA). Recovery studies showed that the efficiency ranged from 90 to 105% with a coefficient of variation (CV) of 6–10% for all 16 PAHs.

The quality assurance was also carried out by analyzing blanks sample spiked with known quantity of each PAHs standard. The values of PAHs in the blanks samples were below the detection of the instrument (0.01 μ g/ml).

Apparatus

The examined samples were measured using a Hewlett Packard 5890 series II GC equipped with a flame ionization detector (FID). The apparatus is equipped with splitless injector (3μ l). The detector maintained at 300°C, while the injection port maintained at 290 °C. A fused silica capillary column (30m, 0.32mm, 0.17μ m) coated with HP-1 (100% dimethyl polysiloxane). The oven temperature program was ranged from 60 to 290°C, ramped at 3°C/min and maintained at 290°C for 25 min. Nitrogen gas was used at 1.2 ml/min.

RESULTS AND DISCUSSION

Hydrocarbons in sediment samples

Table 1. Total cone			eight) in seuthent and fish	samples
Sediments sample	Total PAHs	Fish samples	Fish species	Total PAHs
Ι	1894.35	1	Liza subviridis	3470.33
II	2200.48	2	Parupeneus forsskali,	1014.30
III	1667.02	3	Upeneus japonicus	658.82
IV	2661.56	4	Lethrinus nebulosus	1311.94
V	2102.73	5	Stephanolepis diaspros	621.96
VI	2083.79	6	Epinephelus areolatus	1427.49
VII	2671.27	7	Sphyraena chrysotaenia	2309.43
VIII	1875.30	8	Pomadasys stridens	1378.75
IX	1685.97	9	Trachurus indicus	2655.30
Х	1894.35	10	Saurida undosquamis	4207.56
-	-	11	Engyprospon sp.	851.80

Table 1: Total concentration of PAHs (ng/g wet weight) in sediment and fish samples

Total average values of $\Sigma 16$ individual PAHs concentrations in sediments samples ranged from (1667.02-2671.27ng/g). The maximum concentration (2671.27ng/g) was noticed at stationVII. In contrast, the minimum concentration (1667.02ng/g) was noticed at location III (Table 1).

The highest concentrations along the gulf of Suez were recorded at stations IV (Ras Gharib), VII (Aion Mousa) and X (Abu Zanima) respectively. This observation may be due to heavy shipment of oil tanker and oil drilling activities concentrated at Suez Gulf. Acenaphthyle, acenaphthene, fluorene, anthracene, phenanthrene, pyrene, chrysene, benzo (b) fluoranthene, benzo (k) fluoranthene, benzo (a) pyrene and dibenzo (a,h) anthracene not directed in the area during 2016.

Stations	Sand	Silt	Clay	WT	TOM	ΣLMW	ΣHMW
			%			ng	g/g
1	74.64	10.55	14.81	21.52	0.52	5.37	1888.98
2	91.33	5.89	2.69	14.32	0.28	6.24	2194.24
3	88.50	7.44	4.06	17.33	0.61	4.73	1662.29
4	70.55	14.33	15.12	23.89	1.89	7.54	2654.02
5	77.81	9.48	12.71	20.66	0.94	5.96	2096.77
6	89.56	5.66	4.78	18.45	0.88	5.91	2077.88
7	68.55	10.58	20.87	26.33	2.01	7.87	2663.4
8	83.24	9.88	6.88	19.64	0.57	5.3	1870.0
9	80.47	6.65	12.88	20.45	1.03	4.78	1681.19
10	69.22	10.55	20.33	25.33	1.95	7.09	2493.45

Table 2: Sediment characterizations, organic matter percent, high and low molecular weight of PAHs

The concentrations of PAHs in sediment have been previously classified as low, moderate, high and very high contaminated (0-100, 100-1000, 1000-5000, >5000 ng/ g (Baumard *et al.*, 1998). The levels of PAHs in exanimated sediments of the present study ranged between 1667.02 and 2671.27ng/ g therefore, the surface sediments of Suez gulf can be classified from moderate high contaminated.

The pattern of PAHs based on number of aromatic rings shows four rings as the predominance in all examined sediments (Table 3). These results are significantly lower than those reported by Wang *et al.* (2001) for Boston Harbor, United States and La Rocca *et al.* (1996) for Venice, Italy. However, the results of the present study are also close to the concentrations in the Hellenic coastal zone, eastern Mediterranean (Botsou & Hatzianestis, 2011).

Sample	2-3 rings (%)	4 rings (%)	5 rings (%)	6 rings (%)
Ι	0.283	90.859	0.852	8.005
II	0.284	90.859	0.852	8.006
III	0.284	90.859	0.852	8.005
IV	0.283	90.859	0.852	8.005
V	0.283	90.859	0.852	8.005
VI	0.284	90.859	0.852	8.006
VII	0.295	94.408	0.885	4.413
VIII	0.283	90.860	0.853	8.004
IX	0.284	90.859	0.852	7.197
Х	0.284	90.859	1.263	10.675

Table 3: Percent of PAHs collected sediment samples based on the number of aromatic rings

A significant correlation was found in the present study between total organic matter (TOM), grain size and PAHs. This observation confirms that TOM and grain size play the most important role in the behaviour of PAHs distribution in the sediment (Nudi *et al.*, 2007; Mostafa *et al.*, 2009).

Moreover, significant correlation was found in the present study between clay sediments and PAHs (Σ LMW+ Σ HMW), the same occurred to water content (WC) (Table 2). Actually, there is a great affinity between the levels of low molecular weight PAHs and organic matter whether it's autochthonous organic matter from living origin or allochthonous organic matter from terrestrial origin.

Sediment								PAHs								
sample number	Indeno(1,2, cd) pyrene (IcdP)		Acenaphthylan (ACY)	Naphthaler (NP)	Pyren (PYR	Dibenzo (a,h) Anthracene (DahA)	Fluoranther (FLA)	Benzo(b) Fluoranther (BbF)	Acenaphther (ACE)	Chrysen (CHR)	Benzo(s pyrene (BaP)	Fluoren (FL)	Benzo(K) fluoranther (BkF)	Benzo (ghi) Perylene (BghiP)	Anthracer (ANT)	Phena- nthrene (PHE)
I	151.65	870.13	ND	5.37	ND	ND	851.06	ND	ND	ND	16.14	ND	ND	ND	ND	ND
II	176.16	1010.74	ND	6.24	ND	ND	988.59	ND	ND	ND	18.75	ND	ND	ND	ND	ND
III	133.45	765.71	ND	4.73	ND	ND	748.93	ND	ND	ND	14.2	ND	ND	ND	ND	ND
IV	213.07	1222.53	ND	7.54	ND	ND	1195.74	ND	ND	ND	22.68	ND	ND	ND	ND	ND
V	168.33	965.84	ND	5.96	ND	ND	944.68	ND	ND	ND	17.92	ND	ND	ND	ND	ND
VI	166.82	957.14	ND	5.91	ND	ND	936.17	ND	ND	ND	17.75	ND	ND	ND	ND	ND
VII	117.87	1274.91	ND	7.87	ND	ND	1246.97	ND	ND	ND	23.65	ND	ND	ND	ND	ND
VIII	150.1	861.4	ND	5.3	ND	ND	842.5	ND	ND	ND	16	ND	ND	ND	ND	ND
IX	134.97	774.42	ND	4.78	ND	ND	757.44	ND	ND	ND	14.36	ND	ND	ND	ND	ND
Х	200.18	1148.57	ND	7.09	ND	ND	1123.4	ND	ND	ND	21.3	ND	ND	ND	ND	ND

Table 4: Levels of PAH (ng/g) in sediment of Suez Gulf during 2016.

Hydrocarbons in fish samples

Results for PAHs analysis in the eleven fish species are summarized in Table 5. The distribution of PAHs studied shows different patterns for the eleven species studied. The total concentration of the PAHs varies in the range of 621.92 to 4207.56 ng/g wet weight (Table 1). The fish species *Stephanolepis diaspros* showed the lowest concentration 621.29 ng/g, while the highest (4207.56 ng/g) was found in *Saurida undosquamis* species.

					Fish	ı sample n	umber				
PAH	1	2	3	4	5	6	7	8	9	10	11
Naphthalene(NP)	5.22	11.11	10.51	2.39	4.93	8.55	6.10	23.63	10.78	18.57	12.64
Acenaphthylene (ACY)	ND	1.98	ND	ND	ND	5.42	1.56	ND	ND	ND	ND
Acenaphthene (ACE)	ND	4.16	ND	ND	ND	5.92	1.80	ND	3.02	ND	ND
Fluorene (FL)	ND	6.99	ND	ND	ND	8.45	1.13	1.45	ND	ND	ND
Phenanthrene (PHE)	ND	1.86	ND	1.42	ND	10.22	8.51	ND	24.06	95.66	ND
Anthracene (ANT)	140.54	1.90	10.73	5.66	ND	3.99	12.67	2.41	ND	ND	9.84
Fluoranthene (FLA)	1932.05	628.98	254.66	364.61	348.95	589.77	418.15	697.41	598.40	1001.29	402.66
Pyrene (PYR)	636.78	38.99	ND	ND	ND	19.24	188.40	ND	479.44	633.62	ND
Chrysene (CHR)	27.85	14.36	ND	230.21	ND	ND	ND	26.06	295.70	44.04	ND
Benzo(a) anthrathene (BaA)	84.81	256.82	215.18	197.94	179.23	441.64	211.03	299.40	175.92	476.08	132.15
Benzo(K)fluoranthene (BkF)	3.53	3.24	ND	11.73	ND	1.67	1.40	2.42	90.81	ND	ND
Benzo(b)fluoranthene(BbF)	12.11	12.57	ND	11.49	ND	9.32	26.92	ND	133.59	5.37	ND
Benzo(a)pyrene (BaP)	4.62	2.77	2.39	3.88	1.24	6.11	3.02	3.10	22.59	6.74	1.08
Dibenzo(a,h) anthracene (DahA)	457.78	12.40	99.50	356.33	82.24	191.28	1364.56	128.81	595.63	1796.83	284.70
Benzo(ghi) perylene (BghiP)	139.74	5.20	55.07	105.44	ND	91.67	12.94	55.43	191.43	64.88	2.91
Indeno(1,2,3-cd)pyrene (IcdP)	25.30	10.97	10.78	20.84	5.37	34.24	51.24	138.63	33.93	64.48	5.82
Total ND not detected	3470.33	1014.30	658.82	1311.94	621.96	1427.49	2309.43	1378.75	2655.30	4207.56	851.80

Table 5: Concentration of individual PAHs (ng/g wet weight)

The percent distribution of PAHs based on number of aromatic rings indicated the predominance of four rings PAHs in all studied species except with *Sphyraena chrysotaenia* species where the five membered rings prevailed (Table 6). Among the different components of PAHs, fluoranthene was the most abundant one with the highest value found in *Liza subviridis* (1932.05 ng/g) and lowest value found in *Upeneus japonicus* (254.66 ng/g), with stark contrast found in species *Sphyraena Chrysotaenia* and *Saurida undosquamis* where dibenzo (a,h) anthracene was the highest PAHs with concentrations 1364.56 ng/g and 1796.83 ng/g respectively. Generally the HPAHs were highly prevail compared with LPAHs. This finding is consistent with that reported by Nwaichi and Ntorgbo(2016) for PAHs assessment in some fish and seafood from Niger Delta.

Sample	Fish species	2-3 rings (%)	4 rings (%)	5 rings (%)	6 rings (%)
1	Liza subviridis	4.200	77.269	13.775	4.756
2	Parupeneus forsskali,	2.761	92.591	3.054	1.594
3	Upeneus japonicus	3.224	71.315	15.466	9.995
4	Lethrinus nebulosus	0.722	60.427	29.226	9.625
5	Stephanolepis diaspros	0.793	84.922	13.422	0.863
6	Epinephelus areolatus	2.981	73.601	14.598	8.820
7	Sphyraena chrysotaenia	1.376	35.402	60.443	2.779
8	Pomadasys stridens	1.994	74.188	9.743	14.075
9	Trachurus indicus	1.426	58.353	31.734	8.487
10	Saurida undosquamis	2.715	51.218	42.993	3.074
11	Engyprospon sp.	2.639	62.786	33.550	1.025

Table 6: Percent of PAHs collected fish species based on the number of aromatic rings

Depuration or biotransformation may be the cause of the absence of some PAHs (Deb *et al.* 2000). These concentrations seem higher than that reported by Malik *et al.* (2008) (0.207-3.365 ppm) and those reported for fish muscles from Pearl river delta (49.59 ng/g wet wt; Kong *et al.*, 2005) and Red Sea fish (12.29 ng/g wet wt; DouAbul *et al.*, 1997), where the authors found high concentration of LPAHs. However, comparing the PAHs concentration between different studies is limited by several variables, such as the methodology used, number of samples taken, biological variability (age, diet, size) and environmental parameters (Meador *et al.*, 1995; Vives *et al.*, 2004).

PAHs composition and sources.

The most important sources of PAHs in the marine environment are either petrogenic or pyrogenic. Petrogenic source comes from natural seep of petroleum or petroleum products into the environment, while pyrogenic comes from incomplete combustion of fossil fuels. (Neff 1979; Brown 2002). Several studies have used various PAHs concentration diagnostic ratios to study the source of PAHs (Hoffman et al. 1984; Brown and Peake 2006). For example, the low molecular weight PAHs (LPAHs)/high molecular weight(HPAHs) ratio. The LPAHs include two to three rings while the HPAHs contain four to six rings and are highly carcinogenic. With LPAHs/HPAHs> 1 indicates petrogenic while more than one indicates pyrogenic (Zhang et al., 2008). In our result for all stations as well as the eleven species we found the ratios are less than one which is suggested a pyrogenic origin (Table 7). Several authors used the ratios of individual PAHs levels for identification the source of these concentrations (Kavouras et al., 2001; Katsayiannis et al., 2007; Kim et al., 2008). Sicre et al. (1987) indicated that fluorancene/pyrene (FLA/PYR)<1 suggested petrogenic origin, while higher than one refers to pyrolytic source. Table 6 shows FLA/PYR ratios for the analyzed species, all detectable values were higher than 1 indicated pyrolytic origin. More significant diagnostic ratios applied to the present results were PHE/ANT, with PHE/ANT>10 indicates petrogenic and lower than 10 indicates pyrogenic source, Budzinski et al. (1997). The PHE/ANT ratios for our samples were all less than 10 again suggested pyrogenic source. Moreover, ANT/(ANT+PHE) diagnostic ratio suggested pyrogenic origin with value <0.1 indicate petrogenic source while values higher than 0.1 suggest pyrogenic (Pies et al. 2008). The values of ANT/(ANT+PHE) for species Parupeneus forsskali, Lethrinusn ebulosus Epinephelus areolatu and Sphyraena chrysotaenia were 0.505, 0.799, 0.281

and 0.598 respectively which confirm pyrogenic origin. BaA/ (BaA+CHR) ratio has been implemented to identify the source of PAHs with values higher than 0.35 indicating combustion source and less than 0.2 indicating petrogenic (Ynker *et al.*, 2002; Akyüz and Çabuk, 2010).

Two more diagnostic ratios were used in this study FLA/(FLA+PYR) and IcdP/(IcdP+BghiP). The former with ratio <0.4 for petrogenic, 0.4-0.5 fossil fuel combustion and >0.5 for wood and coal combustion (De La Torre-Roche *et al.* 2009). While the later with ration <0.2 indicating petrogenic, 0.2-0.5 petroleum combustion and >0.5 for wood and coal combustion (Yunker *et al.*, 2002).

Sample	Fish species	LPAHs/	PHE/	FLA/	ANT/	BaA/	FLA/	IcdP/	Origin
no.		HPAHs	ANT	PYR	(ANT+PHE)	(BaA+CHR)	(FLA+PYR)	(IcdP+BghiP)	
1	Liza subviridis	0.043843	ND	3.034	ND	ND	0.752	0.153	Pyrogenic
2	Parupeneus forsskali,	0.028389	0.979	16.132	0.505	0.947	0.942	0.678	Pyrogenic
3	Upeneus japonicas	0.033313	ND	ND	ND	ND	ND	0.164	Pyrogenic
4	Lethrinus nebulosus	0.007271	0.251	ND	0.799	0.462	ND	0.165	Pyrogenic
5	Stephanolepis diaspros	0.00799	ND	ND	ND	ND	ND	ND	ND
6	Epinephelus areolatu	0.030723	2.561	30.653	0.281	ND	0.968	0.272	Pyrogenic
7	Sphyraena chrysotaenia	0.013949	0.672	2.219	0.598	ND	0.689	0.798	Pyrogenic
8	Pomadasys stridens	0.020344	ND	ND	ND	0.920	ND	0.714	Pyrogenic
9	Trachurus indicus	0.014465	ND	1.248	ND	0.373	0.555	0.151	Pyrogenic
10	Saurida undosquamis	0.027906	ND	1.580	ND	0.915	0.612	0.498	Pyrogenic
11	Engyprospon sp	0.027107	ND	ND	ND	ND	ND	0.667	Pyrogenic

Table 7: PAHs diagnostic ratios for source identification in the collected fish species.
--

CONCLUSION

In this study, 16 polycyclic aromatic hydrocarbons (PAHs) were tested in ten samples of surface sediment as well as eleven fish species collected from Suez gulf. The result indicated the total average levels in sediments samples ranged from 1667.02 to 2671.27ng/g. The results for PAHs analysis in the eleven fish species showed the predominance of PAHs with high molecular weight (4-6 rings). The smallest total PAHs found in *Stephanolepis diaspros* (621.9 ng/g), while the highest found in *Saurida undosquamis* (4207.5ng/g). The identification of the source based on the results obtained from several diagnostic ratios suggested pyrogenic source for PAHs in the studied fish. This study provides useful information about the extent and source of PAHs pollutants in different fish species from the Gulf of Suez, which will be helpful for future comparison.

REFERENCES

- Akyüz M. and Cabuk H. (2010). Gas-particle partitioning and seasonal variation of polycyclic aromatic hydrocarbons in the atmosphere of Zonguldak, Turkey.Science of the Total Environment 408: 5550-5558.
- Baumard P.; Budzinski H.; Garrigues P. (1998b). Polycyclic aromatic hydrocarbons in sediments and mussels of the western Mediterranean sea. Environmental Toxicology and Chemistry. 17: 765-776.
- Baumard, P.; Budzinski, H.; Garrigues, P. (1998). Polycyclic aromatic hydrocarbons in sediments and mussels of the western Mediterranean sea. Environ. Toxicol. Chem. 17: 765–776.

- Botsou F. and Hatzianestis I. (2009). Polycyclic aromatic hydrocarbons (PAHs) in marine sediments of the Hellenic coastal zone, eastern Mediterranean: levels, sources and toxicological significance. J Soils Sediments, 12:265–277.
- Brown, J. (2002). Partitioning of chemical contaminants in urban stormwater. Dissertation, University of Otago, Dunedin, New Zealand.
- Brown, J. and Peak, B. (2006). Source of heavy metals and polycyclic aromatic hydrocarbons in urban stormwater runoff. Science of the total environment, 359: 145-155.
- Budzinski H.; Jones I.; Bellocq J.; Pierard C.; and Garrigues P. (1997). Evaluation of sediment contamination by polycyclic aromatic hydrocarbons in the Gironde estuary. Marine Chemistry, 58, 85-97.
- De La Torre-Roche R.J.; Lee W.Y.; Campos-Díaz S.I. (2009). Soil-borne polycyclic aromatic hydrocarbons in El Paso, Texas: analysis of a potential problem in the United States/Mexico border region. Journal of Hazardous Materials 163: 946-958.
- Deb S.C.; Araki T.; Fukushima T. (2000). Polycyclic aromatic hydrocarbons in fish organs, Marine Pollution Bulletin, 40(10): 882-885
- Delgado, C.L.; Wasa, N.; Rosegrant, M. W.; Meijer, S., and Muhfuzuddin, A. (2003). Fish to 2020. Supply and demand in changing global markets. Washington: International Food Policy Research Institute/ World Fish centre
- DouAbul A.A.Z.; Heba H.M.A.; Fareed K. H. (1997). Polynuclear aromatic hydrocarbons (PAHs) in fish from the Red Sea coast of Yemen, Hydrobiologia, 352: 251-262.
- El Zokm, G.M.; Okbah, M.A. and Younis, A.M. (2015). Assessment of Heavy Metals Pollution Using AVS-SEM and Fractionation Techniques in Edku Lagoon Sediments, Mediterranean Sea, Egypt. Journal of Environmental Science and Health, Part A, 50: 1-14.
- El-Nemr, A.; Khaled, A.; El-Sikaily, A.; Said, T.O.; Abd-Alla A.M.A. (2006b). Distribution and sources of polycyclic aromatic hydrocarbons in surface sediments of the Suez Gulf. Environ Monit Assess. doi:10.1007/s10661-005-9009-4.
- Hoffman, E.; Millers, G.; Latimer, J.; Quinn, J. (1984). Urban runoff as a source of polycyclic aromatic hydrocarbons to coastal waters. Environmental Science & Technology, 18: 580-587.
- Katsayiannis, A.; Terzi, E.; Cai, Q.Y. (2007). On the use of PAH molecular diagnostic ratio in sewage sludge for understanding of PAH sources. Chemosphere, 69: 1337-1339.
- Kavouras, I.; Koutrakis, P.; Tsapakis, M.; Lagoudaki, E.; Stephanous, E.; Baer, D.; Oyola, P. (2001). Source appointment of urban aliphatic and polyaromatic hydrocarbons (PAHs) using multivariate methods. Environmental Science and Technology, 35: 2288-2294.
- Kim, M.; Kennicutt, M.C.I.; Qian, Y. (2008). Source characterization using compound composition and stable carbon isotope ratio of PAHs in sediments from lakes harbor and shipping waterway. Science of Total Environment, 389: 367-377.
- Kong K.Y.; Cheung K.C.; Wong C.K.C.; Wong M.H. (2005). The residual dynamic of polycyclic aromatic hydrocarbons and organochlorine pesticides in fishponds of the Pearl River delta, South China, Water Research, 39(9): 1831-1843.

- Liang, Y.; Tse, M.F.; Young, L. and Wang, M.H. (2007). Distribution pattern of polycyclic aromatic hydrocarbons (PAHs) in the sediments and fish at Mai Po Marshes Nature Reserve, Hong Kong. Water Research, 41: 1301-1311.
- Meador, J.P.; Stein, J.E.; Reichert, W.L.; Varanasi U. (1995). Bioaccumulation of polycyclic aromatic hydrocarbons by marine organisms. Reviews of Environmental Contamination and Toxicology, 143:79-165.
- Mostafa, A.R.; Wade, T.L.; Sweet, S.T.; Al-Alimi, A.K.A. (2009). Distribution and characterization of polycyclic aromatic hydrocarbons (PAHs) in sediments of Hadhramout coastal area, Gulf of Aden, Yemen. J. Mar. Syst. 78:1–8.
- Neff, J. (1979). Polycyclic aromatic hydrocarbons in the aquatic environment: sources, fates, and biological effects. London: Applied Science Publisher Ltd.
- Nudi, A.H.; Wagener, A.L.R.; Francioni, E.; Scofield, A.L.; Sette, C.B.; Veiga, A. (2007). Validation of Ucidescordatus as a bioindicator of oil contamination and bioavailability in mangroves by evaluating sediment and crab PAH records. Environ. Int., 33: 315–327.
- Nwaichi, E.O. and Ntorgbo S.A. (2016). Assessment of PAHs levels in some fish and seafood from different coastal water in the Niger Delta, Toxicology Report, 3: 167-172.
- Okbah, M. A.; El-Zokm, G.M.; Younis, A.M. (2015). Heavy metals fractionation and acid volatile sulfide (AVS) in the Bardawil Lagoon Sediments, Northern Sinai, Egypt. Dev Anal Chem, 2:1–9.
- Pies, C.; Hoffmann, B.; Petrowsky J.; Yang Y.; Ternes, T.A. and Hofmann, T. (2008). Characterization and source identification of polycyclic aromatic hydrocarbons (PAHs) in river bank soils. Chemosphere, 72: 1594-1601.
- Said, T.O.; Farag, R.S.; Younis, A.M.; Shreadah, M.A. (2006). Organotin Species in Fish and Bivalves Samples Collected from the Egyptian Mediterranean Coast of Alexandria, Egypt. Bulletin of Environmental Contamination and Toxicology, 77: 451-458.
- Shreadah, M.A.; Said, T.O., Younis, A.M.; Farag, R.S. (2006a). Speciation of Organotin Compounds in Sediments of Semi-Closed Areas along the Mediterranean Coast of Alexandria," Chemistry and Ecology, 22 (5): 395-404.
- Shreadah, M.A.; Said, T.O.; Younis, A.M. and Farag, R.S. (2006b). Physicochemical characteristics of the semi-closed areas along the Mediterranean coast of Alexandria. Egypt. Egypt. J. Aquat. Res., 32:38-48.
- Sicre, M.A.; Marty, J.C.; Saliot A. (1987). Aliphatic and aromatic hydrocarbons in different sized aerosols over the Mediterranean sea: occurrence and origin. Atmospheric Environment, 21: 2247-2259.
- Snousy, M.G.; Zawrah, M.F.; Abdel-Moghny, T., Ebiad, M.A.; Rashad, A.M.; Khalil, M.; Abu El Ella, E.M.; El-Sayed, E. and Tantawy M.A. (2016). Mobility and Fate of Pollutants in the Aquifer System of the Northwestern Suez Gulf, Egypt. P. de Voogt (ed.), Reviews of Environmental Contamination and Toxicology,v. 240, p. 170-192, DOI 10.1007/3985.
- UNEP. (1988). Determination of DDTs and PCBs by capillary Gas chromatography and electron capture detector. Reference methods for marine pollution studies No. 67p
- UNEP/IOC/IAEA. (1991). Sampling ofselected marine organisms and sample preparation for analysis of chlorinated hydrocarbons. References methods for marine pollution studies no. 12, revision 2. Nairobi: United Nations Environment Program 17.

- United States Environmental Protection Agency (USEPA) (1994). Method of Soxhlet Extraction (EPA Method 3540C).
- Vives, I.; Grimalt, J.O.; Fernandez, P. and Rosserand, B. (2004). Polycyclic aromatic hydrocarbons in fish from remote and high mountain lakes in Europe and Greenland. Science of the Total Environment. 324: 67-77.
- Younis A.M.; Nafea E.M.A.; Mosleh, Y.Y.I. and Hefnawy M.S. (2016). Low cost biosorbent (Lemnagibba L.) for the removal of phenol from aqueous media. Journal of Mediterranean Ecology, 14: 55-62.
- Younis, A.M. and Nafea, S.M. (2012). Impact of Environmental Conditions on the Biodiversity of Mediterranean Sea Lagoon, Burullus Protected Area, Egypt. World Applied Sciences Journal, 19: 1423-1430.
- Younis, A.M.; Amin, H.F.; Alkaladi, A. and Mosleh, Y.Y.I. (2015). Bioaccumulation of Heavy Metals in Fish, Squids and Crustaceans from the Red Sea, Jeddah Coast, Saudi Arabia. Open Journal of Marine Science, 5: 369-378.
- Younis, A.M., El-Zokm, G.M. and Okbah, M.A. (2014). Spatial Variation of Acid-Volatile Sulfide and Simultaneously Extracted Metals in Egyptian Mediterranean Sea Lagoon Sediments. Environmental Monitoring and Assessment, 186: 3567-3579.
- Yunker, M.B.; Macdonald, R.W.; Vingarzan, R.; Mitchell, R. H.; Goyette, D. and Sylvestre, S. (2002). PAHs in the Fraser River basin: a critical appraisal of PAHs ratios as indicators of PAHs source and composition. Organic Geochemistry, 33:489-515.
- Zhang, W.; Zhang, S.; Wan, C.; Yue, D.; Ye, Y. and Wang X. (2008). Source diagnostics of polycyclic aromatic in urban road runoff, dust, rain and canopy throughfall. Environmental Pollution, 153: 594-601.