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# A Promising Approach to Control Aquatic Developmental Stages of *Culex Cx. univittatus* Theobald, 1901 Using 4G Mobile (HSDPA 2100) Radiation

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#### ABSTRACT

The International Telecommunication Union reported more than 7 billion cellphone subscriptions worldwide. Therefore, the net result is the production of a number of biological effects on the whole biomolecules, cells and organisms. Such effects induced changes in the intracellular ion concentrations, the rate of composition of different biomolecules, cell reproduction rates and animal reproductive capacity. Biologically, The emitted electromagnetic field (EMF) impacted the exposed insect populations. Both of the larval instars, and the pupal lifespan were significantly depressed due to exposure to mobile radiation. Additionally, the failure of adult emergence increased significantly. Moreover, the sex ratio of Cx. (Cx.) univitattus was significantly affected by exposure to mobile radiation. The duration of the first larval instar declined from  $1.36 \pm 0.13$ days, compared to  $0.90 \pm 0.34$  (P< 0.01) day for both the control and the exposed larvae, respectively. Meanwhile, the durations of the second larval instar decreased from 2.53  $\pm$  0.27 to 1.2  $\pm$  0.09 days for both the control and the exposed larvae (P < 0.05), respectively. Moreover, the duration of the third larval instar was declined from  $3.78 \pm 0.40$ , compared to  $1.09 \pm 0.05$  days for both the control larvae and the exposed larvae group (P < 0.05), respectively. Additionally, the duration of the fourth larval instar decreased (5.27  $\pm$  0.9 days) for the exposed larvae group compared to the control (8.33  $\pm$  2.18 days; P < 0.001). The total larval duration was  $16.00 \pm 2.98$  days compared to  $8.46 \pm 1.38$  days for both the control and the exposed larvae (P < 0.001), respectively. Meanwhile, the pupal lifespan was significantly affected by the exposure to mobile radiation  $(2.2 \pm 0.1)$ days, and 2.5  $\pm$  0.9 days; P< 0.05) for the control and the exposed pupae, respectively. The adult emergency was significantly affected by the exposure to mobile radiation (4:3, and 16:1; P < 0.001) for the control and the exposed groups, respectively. The same pattern was true for the males and females emergency, which was significantly affected (1:1.8) for males, and (1:2.7) females of both the control group and the exposed groups, respectively.

### INTRODUCTION

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*Culex Culex univittatus* Theobald, 1901 (Diptera: Culicidae) is known as ornithophilic mosquito, which is capable of transmitting various viral diseases such as SINV (Sindbis virus) and USUV (Japanese encephalitis virus serocomplex). The former was first isolated from specimens collected in Cairo in 1952 (**Taylor** *et al.* **1955**), while

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the latter was originally isolated in South Africa in 1959 (Williams *et al.*, 1964). In South Africa, *Cx. (Cx.) univittatus* is well known to be as an efficient vector of WNV (West Nile virus) to birds (Jupp, 1974) and Rift Valley fever (RVFV) (Eisa *et al.*, 1977).

Recent telecommunications industry produced a magnificent prevalence within the number of wireless devices. Mobile services have led to a proliferation of infrastructure support in the form of cell towers that provide a link to and from a mobile phone. With no regulations on the status of cell towers, they are placed closer to schools, nurseries, public playgrounds, commercial buildings, hospitals, university campuses, and amphitheaters of densely populated urban residential areas (**Feynman** *et al.* **2013**). In 2015, the International Telecommunication Union reported more than 7 billion cellphone subscriptions worldwide. This led to changes in the intracellular ion concentrations, the rate of composition of different biomolecules, cell reproduction rates and animal reproductive capacity (Lin-Liu & Adey, 1982; Dutta *et al.*, 1984; Adey, 1988; Goodman *et al.*, 1995; Kwee & Raskmark, 1998; Penafiel *et al.*, 1997; Velizarov *et al.*, 1999; Xenos & Margas, 2003).

Additionally, such probable effects may be relevant for insects, essentially for those which live passionate about learning and memory. The radiation emitted from mobile phones,—could have contributed to the dramatic decline in insect populations (Balmori, 2006, 2009, 2014, 2015, 2021; Hallmann, 2017; Thill, 2020).

Pervasive media reports asserted that cellular phones were responsible for the collapse and disorder of the honey bee colonies (Good Morning America, 2007; Kimmel *et al.*, 2007; Mixson *et al.*, 2007; Sharma & Kumar, 2010; Kumar *et al.*, 2011; Favre, 2011; Sainudeen, 2011; Mal & Kumar, 2014; Taye *et al.*, 2017; Kumar, 2018; Favre & Johansson, 2020).

The biological impacts of the electromagnetic field (EMF) on insects were detected, including the effect on the reproductive capacity of *Drosophila melanogaster* (**Panagopoulos** *et al.*, 2004, 2007); the pupae of the house fly *Musca domestica* (**Stanojević** *et al.*, 2005); ant food sites cues (**Cammaerts** *et al.*, 2012); the developmental periods, the adult longevity, the adult weight, and the fecundity of subsequent generations of *Callosobruchus chinensis*, *Marucavitrata*, *Nysius plebeius* and *Nysius hidakai* (**Maharjan** *et al.*, 2019a, 2019b, 2020) in addition to the impact on the survival of fruit flies and their reproductive organs' morphology (**Sudaryadi** *et al.*, 2020).

The main goal of the existing experimental study aimed to record the expected insecticidal effects of mobile phone radiation on the developmental stages of Cx (Cx) *univittatus* (Diptera: Culicidae).

# **MATERIALS AND METHODS**

#### Field collection, identification and mass rearing

In October 2020, a laboratory colony of Cx. (Cx.) univitattus Theobald, 1901 was initiated by collecting larvae from a breeding site in East Gara, Sakaka, Aljouf, Saudi Arabia. Random samples were killed in boiling water, dehydrated in alcohol and preserved in alcohol (70%). The preserved larvae were mounted in euparal until microscopic examination. The larvae were identified according to Harbach (1988). For mass rearing, larvae were maintained under controlled laboratory conditions at 24- 25°C 75-85% relative humidity with а light: twilight: dark and cycle of 12L:1twilight:10D:1dawn. To feed the adult female mosquitoes, a domestic pigeon (Columba livia domestica) was used as a blood meal source, which was provided to the mosquito cages twice a week, following the method of Galal and Seufi (2017).

### **Radiation impact experiments**

Fifty newly hatched mosquito larvae (n=50) were subjected to a single (acute) discontinuous EMF signals produced by using a 4G mobile phone (HSDPA 2100). Two mobile phones were hanged between bottles containing the larvae, and all were surrounded by screen covered to prevent environmental bias. EMF radiation ranged from 900/ 1900 MHz, and power approximately 0.03 mW/cm. Larvae were exposed to that EMF for four hours, followed by observations without radiation treatment. Larval duration, pupal duration, percentage of the adult emergence (%), and the sex ratio were calculated for both the controlled, and the exposed larvae. The experiment was repeated thrice.

#### Statistical analyses

Data were compiled, and calculated using descriptive statistics (the means, standard errors and ranges). Statistical analyses were carried out using SPSS ver. 19 program (SPSS Inc., Chicago, IL). Unpaired, two- tailed Student's *T*- *test* were carried out to compare between group means and determine the significance at (P < 0.05).

# **RESULTS AND DISCUSSION**

The duration of the 1<sup>st</sup> larval instar was  $1.36 \pm 0.13$  days and  $0.90 \pm 0.34$  day for both the control and the exposed larvae (P < 0.01), respectively. Meanwhile, the duration of the 2<sup>nd</sup> larval instar recorded  $2.53 \pm 0.27$  days for the control against  $1.2 \pm 0.09$  day for the exposed larvae (P < 0.05). While, the duration of the 3<sup>rd</sup> larval instar reached  $3.78 \pm 0.40$  days for the control compared to  $1.09 \pm 0.05$  day for the exposed larvae (P < 0.05). For the duration of the 4<sup>th</sup> larval instar, it was  $8.33 \pm 2.18$  days compared to  $5.27 \pm 0.9$  days for both the control and the exposed larvae (P < 0.001), respectively. Additionally, the total

larval duration (16.00  $\pm$  2.98 days) of the control significantly exceeded the whole lifespan of the treated larvae (8.46 $\pm$ 1.38 days, *P*<0.001) (Table 1).

	Duration of larval instar/day					Pupal duration /day
	$1^{st}$	$2^{nd}$	3 <sup>rd</sup>	$4^{th}$	Total larval	
					duration	
Control	(2-1)	(3-2)	(2-5)	(12-3)	(22-8)	(2-4)
	$1.36\pm0.13$	$2.53 \pm$	$3.78 \pm$	$8.33 \pm 2.18$	$16.00\pm2.98$	$2.2 \pm 0.1$
		0.27	0.40			
Exposed	(2-0.4)	(2-1)	(2-1)	(8-3)	(14-5.4)	(2-4)
_	$0.90 \pm .34$	$1.2 \pm$	1.09 ±0.05*	5.27±	8.46±1.38***	$2.5\pm0.9^*$
	**	0.09*		0.9***		

**Table 1.** Effect of mobile radiation on the larval and pupal duration of Cx. (Cx) *univitattus* 

The symbol (\*) refers to a statistically significant difference at P < 0.05; n= 50.

The pupal duration was significantly affected (P < 0.05) by the exposure to mobile radiation since the pupal duration of the control was  $2.2 \pm 0.1$  days, compared to  $2.5 \pm 0.9$  days for the exposed pupae (Table1). In this context, **Dimitrijević** *et al.* (2014) concluded that, ELF magnetic field shortens the developmental time of *Drosophila subobscura*, and **Galal and Seufi** (2022) reported a significant reduction in the developmental time of immature stages of RE-exposed *Culex pipiens* when compared to the control.

The percentage of mortality (%) for all the larval instars, and the pupal stage were significantly (P= 0.00) affected by the exposure of mobile radiation. The mortality percentages of the first larval instar were recorded as 1.36 ± 0.13%, and 3.9 ± 2.61% for both control and exposed larvae (P<0.05), respectively. Whereas, the mortality of the second larval instar was 6.65 ± 1.50 % in case of the control, compared to 7.34 ± 2.31% for the exposed larvae (P<0.05). On the other hand, the recorded mortality of the third larval instar was lower (9.63 ±2.70 %) than that recorded for the exposed group (10.35 ±1.31%; P<0.05). The mortality percentage of the 4<sup>th</sup> larval instar was lower (10.82 ± 2.18%) when compared to the exposed larvae (12.26 ± 3.25%) (P<0.05). To sum up, the total larval mortality increased from 30.85 ± 8.70% for the control to 33.85 ± 9.48% for the exposed larvae (P<0.05), respectively (Table 2).

	% mortality	Pupal % mortality (Failed to				
	1 <sup>st</sup>	2 <sup>nd</sup>	3 <sup>rd</sup>	4 <sup>th</sup>	Total % mortality	emerge)
Control	(1.80-5.30) $1.36 \pm 0.13$	(5.68-9.35) $6.65 \pm 1.50$	(5.65-14.43) 9.63 ±2.70	$(8.53-18.16) \\ 10.82 \pm 2.18$	$\begin{array}{c} (21.66\text{-} 47.24) \\ 30.85 \pm 8.70 \end{array}$	(10.58- 15.95) 13.79 ±0.93
Exposed	(2.30- 6.22) 3.9 ± 2.61*	(5.85-10.42) 7.34 ± 2.31*	(7.11-15.33) 10.35 ±1.31*	(8.65-18.35) 12.26 ± 3.25*	(23.91- 50.32) 33.85 ± 9.48*	(40.43- 60.21) 57.14 ± 0.9***

**Table 2.** Effect of mobile radiation on the percentage mortality (%) of the larval and the pupal stages of Cx. (Cx.) *univitattus* 

The symbol (\*) refers to a statistically significant difference at P < 0.05; n= 50.

The pupal mortality was significantly affected by radiation exposure (control: 13.79  $\pm 0.93$  %, and exposed: 57.14  $\pm 0.9$  %, *p*<0.001).

Similar to the present findings are those of **Ramirez** *et al.* (1983) who confirmed a higher mortality % of eggs, larvae, and pupae of *D. melanogaster* flies, placed for 48h under both pulsed (100 Hz, 1.76 mT). In this respect, **Agrawal** *et al.* (2021) postulated that, the third larval instar of *D. melanogaster* exposed to chronic and acute electromagnetic radiation showed a significant decline in the number of flies (p = 0.007) compared to the control.

It was noticed that, adult emergency and sex ratio were significantly affected by exposure of mobile radiation (P<0.001) as 4:3 and 1:16 (E: F) for adult emergency, and 1:1.8 and 1:2.7 ( $\bigcirc$ :  $\circlearrowleft$ ) for both the control and the exposed independently (Table 3).

**Table 3.** Effect of mobile radiation on adult emergency and sex ratio of *Cx. (Cx.) univitattus* 

	Adult emergency: Emergency failure (E:F)	sex ratio (♀: ♂)
Control	(1.0: 0.5 - 5.0: 4.0) 4.0:3.0	(1.0: 1.35- 1: 1.9) 1.0 :1.8
Exposed	(1.0: 8.7- 1: 20) 1.0 : 16.0***	(1.0: 4.2- 1: 8.3) 1.0 :2.7*

Similar findings are those recorded in the study of Agrawal *et al.* (2021). The authors reported that under chronic electromagnetic radiation, a low number of *D. melanogaster* flies eclosed was detected compared to the control. Additionally, adult flies of *D. melanogaster* exposed to chronic electromagnetic radiation emerged one day later, compared to the control (Agrawal *et al.*, 2021). However, in contrast to the obtained results, Dimitrijević *et al.* (2014) stated that ELF magnetic field does not affect the sex ratio of *D. subobscura*.

Results proved that radiation increased the failure to reach the adult stage and the failure to switch to the extreme phase. The exposure to mobile radiation affected the survival rate significantly during the experimental period. This result coincides with that of Aday (1975). The author postulated that the radiation of 900 MHz is explosively bioactive, causing conspicuous revise in the physiological function of living organisms. GSM 900 MHz radiation was reported to inhibit ants' association between food spots and encountered cues (Adey, 1988). Electromagnetic field (EMF) could incite natural goods on biomolecules and cells (Cammaerts et al., 2012). Panagopoulos et al. (2004) reported a decrease of reproductive capacity of the insect Drosophila melanogaster by 50%- 60%, as an effect of GSM 900 MHz mobile phone radiation. Moreover, Panagopoulos et al. (2007) compared between the natural conditioning of the two systems, GSM (900 MHz) and DCS (1800 MHz). The authors concluded that both types of radiation dropped the reproductive capacity of fruit canvases significantly. Mobile phone is a device that emits the strongest EMF radiation which can suppress the survival of fruit canvases and change the morphology of their reproductive organs (Sudaryadi et al., 2020). Stanojević et al. (2005) reported that the pupae of the house fly Musca domestica, exposed to an EMF (50 Hz) showed braked down transformation. EMF radiation can be a problem for insects resulting from exposure (Balmori 2006, 2009, **2014, 2015**). The EMF affected the experimental period, adult life, adult weight and the fecundity of posterior generations of Callosobruchus chinensis (Coleoptera), Maruca vitrata (Lepidoptera), Nysius plebeius and Nysius hidakai (Hemiptera). The same conclusion was postulated by Maharjan et al. (2019a, 2019b, 2020).

# CONCLUSION

To conclude, all larval instars and pupal life span were significantly delayed by exposure to mobile radiation. In addition, the failure of adult emergency increased significantly as the sex ratio of Cx. (Cx.) univitatus was significantly affected by exposure to mobile radiation, and this failure increased specifically in females.

### REFERENCES

Aday, W.R. (1975). Introduction: Effects of electromagnetic radiation on the nervous system. Annals N.Y. Acad. Sci., 247:1520.

Adey, W.R. (1988). Electromagnetic Field Interactions in the Brain. In: Başar E. (eds) Dynamics of Sensory and Cognitive Processing by the Brain. Springer Ser. in Brain Dyn, vol 1. Springer, Berlin, Heidelberg.

https://doi.org/10.1007/978-3-642-71531-0\_9.

Agrawal, N.; Verma, K.; Baghel, D.; Chauhan, A.; Prasad, D.N.; Sharma, S.K. and Kohli, E. (2021). Effects of extremely low-frequency electromagnetic field on different developmental stages of *Drosophila melanogaster*. Int J Radiat Biol. 97(11):1606-1616.

doi: 10.1080/09553002.2021.1969465. Epub 2021 Aug 31. PMID: 34402374

**Balmori, A.** (2006). The incidence of electromagnetic pollution on the amphibian decline: is this an important piece of the puzzle? Toxicol. Environ. Chem., 88: 287-299. https://doi.org/10.1080/02772240600687200

**Balmori, A.** (2009). Electromagnetic pollution from phone masts. Effects on wildlife.Pathophysiol., 16:191–199.

https://doi.org/10.1016/j.pathophys.2009.01.007

**Balmori, A.** (2014). Electrosmog and species conservation. Sci. Total Environ., 496:314–316. DOI: 10.1016/j.scitotenv.2014.07.061

**Balmori, A.** (2015). Anthropogenic radiofrequency electromagnetic fields as an emerging threat to wildlife orientation. Sci. Total Environ.,518:58–60. DOI:10.1016/j.scitotenv.2015.02.077

**Balmori, A.** (2021). Electromagnetic radiation as an emerging driver factor for the decline of insects. Sci. Total Environ., 767:144913. doi:10.1016/j.scitotenv.2020.144913

Cammaerts, M.C.; De Doncker, P.; Patris, X.; Bellens, F.; Rachidi, Z. and Cammaerts, D. (2012). GSM 900 MHz radiation inhibits ants' association between food sites and encountered cues. Electromagn. Biol. Med., 31:151-65.

**Dimitrijević, D.; Savić, T.; Anđelković, M.; Prolić, Z. and Janać, B.**(2014). Extremely low frequency magnetic field (50 Hz, 0.5 mT) modifies fitness components and locomotor activity of *Drosophila subobscura*. Int. J. Radiat. Biol. 90(5):337-43.doi: 10.3109/09553002.2014.888105. Epub 2014 Mar 19. PMID: 24475738.

**Dutta, S.K. ; Subramaniam, A.; Ghosh, B. and Parshad, R.** (1984). Microwave radiation-induced calcium ion efflux from human neuroblastoma cells in culture. Bioelectromagnet. N.Y., 5:71–78. http://dx.doi.org/10.1002/bem.2250050108

**Eisa, M.; Obeid, H.M.A. and El Sawi, A.S.A**.(1977). Rift Valley fever in the Sudan. Bull. Anim. Health Prod. Afr., 24: 343-347.

**Favre, D.** (2011). Mobile phone induced honey bee worker piping. Apidologie, 42: 270-279.

**Favre, D. and Johansson, O.** (2020). Does enhanced electromagnetic radiation disturb honeybees' behaviour? observations during new year's eve 2019. Internat. J. res. granthaalayah.,8:7-14. https://doi.org/10.29121/granthaalayah.v8.i11.2020.2151

**Feynman, R. ; Leighton, R. and Sands M.** (2013). The Feynman Lectures on Physics. Vol II, Mainly Electromagnetism and Matter, Basic Books, New York.

Galal, F. H.; Abu elnasr, A.; Abdallah, I.; Seufi, AE.M. and Zaki, O. (2015). Isolation and Characterization of Internal Bacteria from the Mosquito, *Culex pipiens* from Egypt. Inter. J. Sci. Res., 4:2682-8.

**Galal, F.H. and Seufi A.M.** (2017). Studies on Biological Attributes of *Culex* (*Culex*) univittatus Mosquito (Diptera: Culicidae) Population From Saudi Arabia. Egypt. Acad. J. Biolog. Sci., 9:13–20.

**Galal, F.H. and Seufi A.M.** (2022). The impact of electromagnetic radio waves on some biological aspects of *Culex (Culex) pipiens* Mosquitoes (Diptera: Culicidae). Biosci. res., 19(2):805-810.

**Good Morning America** (2007). Where have the bees gone? Cell phones are linked to the mysterious disappearance of honey bees. <u>http://abcnews.go</u>. com/video/playerIndex?id = 3044826

Goodman, E.M.; Greenebaum, B. and Marron, M.T. (1995). Effects of electromagnetic fields on molecules and cells. Int. Rev. Cytol., 158:279–338. http://dx.doi.org/10.1016/S0074-7696(08)62489-4

Hallmann, C.A.; Sorg, M.; Jongejans, E.; Siepel, H.; Hofland, N.; Schwan, H.; Stenmans, W.; Muller, A.; Sumser, H.; Horren, T.; Goulson D. and de Kroon, H. (2017). More than 75 percent decline over 27 years in total flying insect biomass in protected areas. PloSone, 12: e0185809. doi:10.1371/journal.pone.0185809.

Harbach, R.E. (1999). The identity of *Culex perexiguus* Theobald versus *Cx. univittatus* in southern Europe. Eur. Mosq. Bull., 4 : 7. <u>http://www.independent.co.uk/environment/nature/aremobile-phones-wiping-out-our-bees-444768.html</u> https://www.who.int/pehemf/publications/reports/IA C\_2011\_Progress\_Report.pdf

International Telecommunication Union. ICT Fact and Figures. International Telecommunication Union: ICT Data and Statistics Division, Geneva. (2015).

**Jupp, P. G. (1974).** Laboratory studies on the transmission of West Nile virus by *Culex (Culex univittatus)* Theobald; factors influencing transmission rate. J. Med. Entomol., 11: 455–458.

Kimmel, S. ; Kuhn, J. ; Harst, W. and Stever, H. (2007). Effects of electromagnetic exposition on the behavior of the honeybee (*Apis mellifera*). Environ. Syst. Res., 8:1-8.

**Kumar, N. R., ; Sangwan, S. and Badotra, P.** (2011). Exposure to cell phone radiations produces biochemical changes in worker honey bees. Toxicol. Int., 18: 70–72.

**Kumar, S.** (2018). Colony Collapse Disorder (CCD) in Honey Bees Caused by EMF Radiation. Bioinform., 14: 521–524. doi:10.6026/97320630014521

Kwee, S. and Raskmark, P. (1998). Changes in cell proliferation due to environmental nonionizing radiation: 2. Microwave radiation. *Bioelectrochem*. Bioenerget., 44: 251-255. http://dx.doi.org/10.1016/S0302-4598(97)00095-0

Lin-Liu, S. and Ade, W.R. (1982). Low frequency amplitude modulated microwave fields change calcium efflux rates from synaptosomes. Bioelectromagnet. N.Y., 3: 309- 322. http://dx.doi.org/10.1002/bem.2250030303

Maharjan, R.; Bae, S.; Kim, G.H.; Yoon, Y.; Jang, Y.; Kim, Y. and Yi, H. (2019b). Oviposition preference and development of *Maruca vitrata* (Fabricius) (Lepidoptera: Crambidae) on different radiofrequency fields. Entomolog. Res., 49: 214–222.

Maharjan, R.; Yi, H.; Ahn, J.; Roh, G.H.; Park, C.; Yoon, Y. and Bae, S. (2019a). Effects of radiofrequency on the development and performance of *Callosobruchus chinensis* (Coleoptera: Chrysomelidae: Bruchinae) on three different leguminous seeds. Appl. Entomol. Zool., 54: 255–266.

Maharjan, R.; Yoon, Y.; Jang, Y.; Jeong, M.; Jung, T.W.; Cho, H.S. and Yi, H. (2020). Artificial radiofrequency driven life-table parameters of perilla seed bugs (*Nysius* sp.) (Heteroptera:Lygaeidae). J. Asia Pac. Entomol., 23: 1264–1271.

**Mal, P. and Kumar, Y.** (2014). Effect of electromagnetic radiation on brooding, honey production and foraging behavior of European honey bees (*Apis mellifera L.*). Afri. J. Agricult. Res., 9:1078-1085.

Mixson, T.A.; Abramson, C. I.; Nolf, S.L.; Johnson, G.A.; Serrano, E. and Wells, H. (2009).Effect of GSM cellular phone radiation on the behavior of honey bees (*Apis mellifera*). Sci. Bee Cult., 1: 22-27.

**Mohamed, S. A.**; Galal, F.H. and Hafez, E.E. (2007). Characterization of a *Schistocerca gregaria* cDNA Encoding a Novel Member of Mobile Phone Radiation-Induced Polypeptide Related to Chitinase Polypeptide Family. J. Appl. Sci. Res., 3: 733-740.

**Panagopoulos, D.J. ; Chavdoula, E.D.; Karabarbounis, A. and Margaritis, L.H.** (2007). Comparison of bioactivity between GSM 900 MHz and DCS 1800 MHz mobile telephony radiation. Electromagnet. Biol. Medic., 26: 33–44.

**Panagopoulos, D.J. ; Karabarbounis, A.y. and Margaritis, L.H.** (2004). Effect of GSM 900 MHz mobile phone radiation on the reproductive capacity of *Drosophila melanogaster*. Electromagnet. Biol. Medic., 23: 29–43.

Penafiel, L.M.; Litovitz, T.; Krause, D.; Desta, A. and Mullins, J.M. (1997). Role of modulation on the effects of microwaves on ornithine decarboxylase activity in L929 cells. Bioelectromagnet., 18:132-141. http://dx.doi.org/10.1002/(SICI)1521-186X(1997)18:2<132::AID- BEM6>3.0.CO;2-3

**Ramirez, E.; Monteagudo, J.L.; Garcia-Gracia, M. and Delgado, J.M**. (1983). Oviposition and development of *Drosophila* modified by magnetic fields. Bioelectromag., 4:315–326.

**Sainudeen, S.S.** (2011). Electromagnetic Radiation (EMR) clashes with honey bees. Inter. J. Environ. Sci., 1: 897-900.

Sharma, V.P. and Kumar, N.R. (2010). Changes in honeybee behavior and biology under the influence of cellphone radiations. Curr. Sci. 98:1376–1378.

**Stanojević, V. ; Prolić, Z. ; Savić, T. ; Todorović, D. and Janać, B.** (2005). Effects of extremely low frequency (50 Hz) magnetic field on development dynamics of the housefly (Musca domestica L.). Electromagnet. Biol. Med., 24: 99–107.

Sudaryadi, I.; Rahmawati, A.N. and Rizqiyah, M. (2020). Effect of handphone EMF radiation on survival rate and morphological reproductive organ changes of fruit fly (*Drosophila melanogaster* Meigen, 1830). THE 6TH INTERNATIONAL

CONFERENCE ON BIOLOGICAL SCIENCE ICBS 2019: "Biodiversity as a Cornerstone for Embracing Future Humanity. doi:10.1063/5.0015846

**Taye, R.R. ; Deka, M.K. ; Rahman, A. and Bathari, M.** (2017).Effect of electromagnetic radiation of cell phone tower on foraging behaviour of Asiatic honey bee, *Apis cerana F*. Hymenoptera: Apidae). J. Entomol. and Zool. Studies, 5: 1527-1529.

**Taylor, R.M Hurlbut, H.S.; Work, T.H.; Kingston, J.R. and Frothingham, T.E.** (1955).Sindbis virus: a newly recognized arthropodtransmitted virus. Am J Trop Med Hyg., 4:844–62.

**Thill, A.** (2020). Biologische Wirkungen elektromagnetischer Felder auf Insekten Felder auf Insekten. Umwelt Medizin Gesellschaft (Sonderbeilage), 3: 31-28.

**Velizarov, S. ; Raskmark, P. and Kwee, S.** (1999). The effects of radiofrequency fields on cell proliferation are nonthermal. Bioelectrochem. Bioenerget.,48:177-180. http://dx.doi.org/10.1016/S0302- 4598(98)00238-4

Williams, M.C.; Simpson, D.I.; Haddow, A.J. and Knight, E.M.(1964). The isolation of West Nile Virus from man and of Usutu virus from the bird-biting mosquito *Mansonia aurites* (Theobald) in the Entebbe area of Uganda. Ann. Trop. Med. Parasitol., 58:367–374.

**Xenos, Th. D. and Margas, I.N.** (2003). Low power density RF radiation effects on experimental animal embryos and foetuses. In, Stavroulakis, P., ed. Biological Effects of Electromagnetic Fields, Springer, pp. 579- 602.