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Free-living nematodes in some mangrove sites on the southern Egyptian Red Sea coast with emphasis on their horizontal distribution.

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ABSTRACT

The generic composition, abundance, and distribution of free-living nematodes along three mangrove sites in the southern Egyptian Red Sea coast were studied. Sediment samples were collected in April 2022 at three different tidal levels in each site. Densities of free-living nematodes varied among sites and tidal levels. The mangrove of El-Readaa site had the most abundant and diversified nematofauna and the mid-tidal level attained the highest densities and number of recorded genera. Nematofauna was represented by 25 genera belonging to 13 families, of them; Xvalidae, Desmodoridae and Linhomoeidae were the most abundant. Terschellingia and Daptonema were the common genera in fine sediment areas and their abundance increased from high water level towards the mangrove trees at the low tidal level. Genus Spirinia was common in the coarser sediments and showed an opposite distribution pattern. A strong correlation occurred between nematodes' abundance, feeding types and sediment characteristics. The abundance of the different nematodes was studied in relation to their feeding habits.

INTRODUCTION

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Mangroves are among the most productive ecosystems found in the intertidal tropical and subtropicals regions and are largely restricted to latitudes between 30° north and 30° south (**Kathiresan** *et al.*, **1996; Ronnback, 1999**). Their core distribution is in the tropical areas, which corresponds with rapid human activities and poses a huge threat on them (**Oog, 1995**). The coverage of mangroves has been estimated to be between 10 million hectares (**Bunt. 1992**) and 24 million hectares (**Twilley** *el al.*, **1996**).

There are a number of ecological roles provided by this ecosystem. Maintain shoreline stability and water quality by controlling nutrient and sediment circulations (Twilley *et al.*, 1996; Semesi, 1998). They also provide a shelter for their neighboring communities from natural hazardous (Semesi 1998; Ronnback, 1999). Mangrove root parts offer microhabitats for vast groups of organisms (Robertson and Alongi, 1992) and act as nursery for fish and macro-invertebrate animals (Robertson and Alongi, 1992;

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Semesi 1998, Ronnback, 1999; Laegdsgaard and Johnson, 2001). Mangroves also present a variety of diverse natural products for many uses, such as fuel material and tannins as well as incomes to the local communities through harvesting, hunting, and fish gathering operations (Ronnback, 1999).

Within the mangrove forest, an abundant variety of benthic assemblages are found (**Olafsson, 1995**). Meiofauna is numerically dominating the metazoan group particularly nematodes (**Hodda and Nicholas, 1985**) since they provide an important food source for higher trophic levels (**Olafsson and Moore, 1990**). The densities and composition of nematodes have been affected by complex factors; sediment texture, depth of sediment, common species and seasonality (**Hogarth, 1990**). The size of sediment grains is an important aspect when describing benthic habitats and directly controls the spatial situations, and indirectly affects the physico-chemical conditions of sediment (**Giere, 2009**).

Only two studies focused on the free-living nematodes of the Red Sea mangroves: **Sabeel and Vanreusel (2015)** in Sudan and **Al-Sofyani and Al-Sherbiny (2018)** in Saudi Arabia. This is the first study to describe the free-living nematodes of the Egyptian Red Sea mangroves to elucidate their generic composition and distribution patterns at the different tidal levels.

MATERIALS AND METHODS

Study area and sediments sampling:

In April 2022; three sites along the western coast of the Egyptian Red Sea were selected to represent the mangrove habitats; Site I, El-Readaa (24°19'25" N 35°20'31" E), Site II, Lahmy Bay (24°13'19" N 35°25'13" E) and Site III, Hertwaay (24°8'3.4" N 35°29'0.1" E) (Fig. 1). Site I contains a small stand of mangroves visible from the mainland. Some disturbance due to fishing activities and considerable litter are present. Site II a small stand of mangrove tree is found a couple of kilometers to the south of Hamata village on two sides of the shallow lagoon. Site III surrounds a large tidal inlet; several stands of mangroves are found. They are mostly short with a mean height of 1.5 m and occasional older trees of about 3.5 m. Landward, mangrove trees become increasingly stunted with signs of intense camel grazing such as damaged apices and dense coppiced branches. As a result of the grazing, the canopies are open and sediments are exposed to increase evaporation which has led to surface salinization. Recruitment is low but present. Litter and tar balls are abundant. Around the mangroves at this site, there were particularly well-developed salt marsh formations, consisting largely of *Arthrocnemum macrostachyum* and *Halopeplis perfoliata*.

In each site, three replicate sediment samples were taken from each of three tidal levels; high tidal marks (H.T.M.), mid tidal marks (M.T.M) and low tidal marks (L.T.M.) during spring tide by means of a hand metal cylindrical corer of 10 cm² (3.5 cm diameter, 10 cm high) for examination of nematodes and sediments characteristics. The YSI 650

multi-parameter instrument (YSI, Yellow Springs, OH, USA) was used to measure water temperature and salinity (expressed as total dissolved salts, TDS) in the field.



Fig. 1: A map of the Egyptian Red Sea coast showing the studied sites.

Sample Analysis:

Sediment samples collected for the investigation of nematodes were fixed in the field using 5% neutral formalin and placed in labeled plastic bottles and brought to the laboratory. They were immediately stained with 0.5 g/l Rose Bengal for 1 h then washed through sieves of 500 µm and 50 µm mesh size. All individuals retained on the 50 µm sieve were collected and counted under a stereomicroscope (Prior S 2000, magnification 100x). All nematodes were picked out and transferred to vials containing glycerol through two series of ethanol-glycerol solutions and mounted on glycerin slides (**De Griesse, 1969**). They were examined underneath a compound microscope (Carl Zeiss 1000x magnification) and identified to the genus level using the pictorial keys by **Tarjan (1980); Platt and Warwick (1983); Plat and Warwick (1988) and Warwick** *et al.*

(1998). Nematodes were categorized to three different feeding types according to the morphology of their buccal cavities (Wieser, 1953).

The other sediment samples were washed with tap water to remove salt, dried at 60° C for 24 h, then placed on top of a series of sieves of mesh sizes 1,000, 500, 250, 125, and 62 µm. The sediments that retained on top of each sieve were weighed then classified to different grain size categories according to the method of **Holme and McIntyre** (**1984**): silt-clay, very fine sand, fine sand, medium sand and coarse sand and gravel. Sediment sorting indices were determined according to **Gray** (**1981**).

For the determination of total organic matter in the sediment samples, 1 g of fine sediment for each one with sizes ranged between 125- 250 μ m were taken and dried at 60 °C overnight in the laboratory. Total organic matter (TOM) was assessed as the percentage weight loss in the samples after combustion at 550 °C for 24 Hours (Jorgensen, 1977).

Data analysis:

To test for the spatial heterogeneity of sediment and abundance of nematode genera between sites and different tidal levels, one-way analysis of variance (ANOVA) with 95% confidence limits were performed. Pearson's Correlation Analysis was used to determine the relationships between the nematode abundance with the sediment characteristics. Analyses were carried out using the statistical software SPSS 18.0 (2002). To assess the structural diversity at each site, nematode diversity indices (Species richness (SR), evenness (Pielou index, J') and the Shannon-Wiener diversity index (H') were calculated using PRIMER v6.0 software (Clarke and Gorley, 2006). All diversity indices were determined at nematode genus level.

RESULTS

1. Physico-chemical parameters and sediment characteristics

In the spatial study, water temperatures, TDS and sediment characteristics displayed little variations among sites. Sediments of site II and III exhibited higher silt/clay content percentages, total organic matter percentages (TOMs) and lower median grain size than site I. Sediments for all sites were classified as "well sorted" textures (Table 1). For horizontal study, the studied physic-chemical parameters and sediment characteristics increased from the high tidal marks (H.T.M.) areas to low tidal marks areas (L.T.M.) in terms of silt/clay percentages and TOMs% while water temperatures, TDS, Mz, sand percentages decreased seawards. The sediment sorting index for the areas of L.T.M was from moderately well sorted type while the other ones were from well sorted types (Table 2).

Site	Temperature °C	TD S	Silt/clay %	Sand %	TOMs %	Mz(µm)	Sediment sorting
Site I	24.6	42.8	48.5	51.5	4.4	275	Well sorted
SiteII	24.5	42.5	51	49	5.8	220	Well sorted
Site III	24.5	42.5	50.8	49.2	5.5	210	Well sorted

Table 1: Water temperature, salinity, silt and clay content %, sediment median grain size (Mz), total organic matter (TOMs) and sorting index at the investigated sites.

Table 2: Water temperature, salinity, silt content %, sediment median grain size (Mz), total organic matter (TOMs) and sorting index at the investigated tidal levels.

Tidal level	Temperature °C	TDS	Silt and clay %	Sand %	TOMs%	Mz (µm)	Sediment sorting
H.M.T.	25.2	43.5	42	58	2.3	325	Well sorted
M.T.M.	24.5	42.7	53	47	5.1	190	Well sorted
L.T.M.	24	42.4	57	43	8.8	147	Moderately well sorted

2. Nematofauna in the spatial study

A total of 13 families represented the nematofauna. Of them, 4 families accounted for 76.6% in terms of abundance namely, Xyalidae (22.4%), Desmodoridae (21.9%), Linhomoeidae (20.5%) and Oncholaimidae (11.9%). In terms of number. of genera, Comesomidae was the most diversified family with 4 genera followed by Dosmodoridae, Leptolaimidae, Oncholamidae and Xyialidae with 3 genera each (Table 3).

Nematoda abundance varied among sites, one way ANOVA showed significant variation among sites (F =89.27, P < 0.001). The highest density value of nematodes was recorded in site I (316±100 individual/10 cm²). Nematoda was represented by 25 genera. Of them 15 were recorded in all sites. *Terschellingia* and *Daptonema* exhibited the highest densities (Table 4). The abundance of the former ranged between 35±21.7 and 55±23.7 individual/10 cm²) with a relative abundance of 18.5% while the latter showed a very close relative abundance value of 18.2% (Fig. 2). *Sprinia* ranked third; its densities ranged between 2.7±1.3 and 55.3±18.7 individual/10 cm² and a relative abundance of about 17% (Table 3 and Fig. 2). Site I had the highest values of number of genera, Shannon-Wiener indices and Species Richness while site III had the lowest apart from the Evenness value which was its highest (Table 5). Deposit feeding was the most common type in all sites (50%) of total nematodes and ranged between 41.1-60.5%. Epistrate feeding was ranked second followed by predator/omnivore feeding (Figs. 3 and 4).

Family	Count	Relative abundance	No. of genera	%
Cyatholaimidae	2.7	0.39	1	4
Monhysteridae	3.3	0.48	1	4
Tripyloididae	4	0.58	1	4
Haliplectidae	4.73	0.68	1	4
Ethmolaimidae	6.7	0.96	1	4
Microlaimidae	13.7	1.97	1	4
Chromadoridae	23.7	3.41	1	4
Leptolaimidae	48.3	6.96	3	12
Comesomidae:	55.1	7.93	4	16
Oncholaimidae	82.5	11.88	3	12
Linhomoeidae	142.3	20.49	2	8
Desmodoridae	152	21.89	3	12
Xyalidae	155.4	22.38	3	12
Total	694.43		25	

Table 3: Total average count of families (expressed as No. of individual/10 cm²), Relative abundance and No. of recorded genera in the studied sites

Table 4: Average genus count in the studied sites expressed as No. of individual/ 10 cm^2 .

		Site I	Site II	Site III
Family	Genera	(Mean±Standard deviation)	(Mean±Standard deviation)	(Mean±Standard deviation)
Chromadoridae	Ptycholaimellus	11.7±5.3	7±2.7	5±1.3
Comesomidae:	Dorylaimopsis	6.7±2.7	-	-
	Laimella	4.3±1.7	-	1.7±0.3
	Paracomesoma.	5.7±1.3	5±0.7	6.7±2.3
	Sabatieria	16.7±7.3	4±1.7	4.3±0.7
Cyatholaimidae	Pomponema	2.7±0.7	-	-
Desmodoridae	Desmodora	8.3±2.7	1.3±0.3	11.7±3.3
	Metachromadora.	11.7±5.7	15±6.7	7.7±2.3
	Spirinia	55.3±18.7	38.3±15.3	2.7±1.3
Ethmolaimidae	Ethmolaimus	1.7±0.7	5±0.3	-
Haliplectidae	Haliplectus	2.7±1.3	0.33±1	1.7±0.3
Leptolaimidae	Camacolaimus	2±0.7	-	-
	Halaphanolaimus	16.7±5.3	1±0.3	13.3±4.7
	Leptolaimus	8±1	2.3±0.7	5±1.3
Linhomoeidae	Metalinhomoeus	5±1.3	2.3±0.7	-
	Terschellingia	55±23.7	45±28.3	35±21.7
Microlaimidae	Microlaimus	7±2.3	2.7±0.3	4±1
Monhysteridae	Diplolaimella	1±0.3	2.3±1	-
Oncholaimidae	Oncholaimellus	5±1.3	21.7±10.7	6.7±0.7
	Oncholaimus	10.7±4.3	17±3.7	5±3.3
	Viscosia	3.7±2.3	12.7±9.3	-
Tripyloididae	Tripyloides	4±1.7	-	-
Xyalidae	Daptonema	51.7±13	36.7±18.7	40.3±15.3
	Stylotheristus	3.3±1.7	2.7±0.3	6.7±1.7
	Theristus	14±3.7	-	-
Total count		316±100	220.3±90	157.5±6



Fig. 2: Relative abundance of the common nematode genera in the studied sites (values are given in the top of each column).



Fig. 3: Percentage of nematodes feeding types in the studied mangroves.





Fig. 4: Percentage of nematodes feeding types in the studied mangroves sites (values are given for each category).

Table 5: Total recorded genera of nematodes (S), total individual count expressed as No. of individual/	10
cm ² (N), Shannon- Wiener (H'), Species Richness (SR) and Evenness (J') at the studied sites.	

	Site I	Site II	Site III
S	25	19	16
Ν	316	220	157
Н'	2.66	2.34	2.35
SR	3.5	2.8	2.4
J '	0.83	0.79	0.85

3. Nematofauna in the horizontal study

The abundance of nematodes varied on the horizontal basis. ANOVA test showed significant variation (F14.46, P<0.01). The highest density was recorded in the mid tidal mark (272±73 individual/10 cm²) while a slight difference was between the densities of the high and low water marks (Table 6). *Terschellingia* attained the highest density (51.7±12.7 individual/10 cm²) followed by *Daptonema* (46.7±23.7 individual/10 cm²) in the mid tidal mark (Table 6). All diversity indices, density and number of genera were higher in the mid tidal levels than the other ones. Nematofauna in the low tidal level showed the lowest values (Table 7). Predator/omnivore feeding type was the least

common with a percentage ranged between 3.9 and 20.5%. Deposit and predator/omnivore feeding types increased seaward while Epistrate feeding showed opposite pattern with a highest percentage in the high tidal mark (Fig. 5).

Table 6: Abundance of nematode genera (expressed as No. of individuals/ 10 cm²) and feeding type in the studied tidal levels where H.T.M=High tide mark, M.T.M=Mid tide mark and L.T.M. =Low tide mark.

Feeding Type	Genera	H.T.M (Mean±Standard deviation)	M.T.M. (Mean±Standard deviation)	L.T.M. (Mean±Standard deviation)
Deposit feeding	Daptonema	30±11.3	46.7±23.7	45.3±19.3
	Halaphanolaimus	13.3±7.3	12.7±5.7	3.3±0.7
	Leptolaimus	4.3±1.7	6.7±2.3	1±0.3
	Sabatieria	3.3±1.3	6±0.7	9±2.3
	Stylotheristus	-	5±1.7	0.7±0.3
	Terschellingia	33.3±7.7	51.7±12.7	39.3±14.3
	Theristus	3.7±1.3	6±0.7	0.3±0.3
Epistrate feeding	Camacolaimus	0.7±0.3	1.3±0.7	-
	Desmodora	9.3±2.7	10±3.3	6±1.3
	Diplolaimella	0.7±0.3	2.3±1.3	0.3±0.3
	Dorylaimopsis	$2.7{\pm}0.7$	5±2.3	_
	Ptycholaimellus	7.3±1.3	12.7±2.3	3.7±0.7
	Ethmolaimus	5.7±1	1±0.3	-
	Laimella	-	3.3±	-
	Metachromadora	15±2.7	10±2.3	9.3±1.7
	Metalinhomoeus	4.3±1.3	2.3±1.7	0.7±0.3
	Microlaimus	7±2.7	4.3±1.3	1.3±0.7
	Paracomesoma .	5±1.3	8.3±2.7	1.7±0.3
	Spirinia	45±12.7	36.7±14.3	32.7±11.7
	Tripyloides	2±0.3	2.7±1.3	0.3±0.3
Predator/omnivore feeding	Oncholaimellus	2.7±1.3	15±8.3	17.3±5.7
	Oncholaimus	3.3±0.7	10±4.3	19.3±10.7
	Viscosia	2.7±1	13.3±2.3	3.3±0.7
	Total count	201±45	272±73	196.7±62



Tidal level

Fig. 5: Percentages of nematode feeding types at the studied tidal levels where H.T.M=High tidal mark, M.T.M=Mid tidal mark and L.T.M. =Low tide marks.

Table 7: Total recorded genera of nematodes (S), total individual count expressed as No. of individual/ 10 cm² (N), Shannon- Wiener (H'), Species Richness (SR) and Evenness (J') of the studied sites.

	H.T.M	M.T.M.	L.T.M.
S	21	23	19
Ν	603	816	587
H'	2.23	2.64	2.2
SR	3.12	3.28	2.82
J '	0.73	0.84	0.75

In the spatial study, Pearson's correlation analysis showed strong significant values between all sediment parameters; silt content %, median grain size (Mz) and total organic matter (Table 8). The abundance and total number of genera of nematodes showed strong negative significant correlations with silt content and moderate ones with TOMs. In contrast, strong positive significant values with sand percentages and Mz were calculated. Epistrate feeding nematodes showed strong positive correlation with sand percentage (r=0.89, P <0.001) while predator/omnivore feeders showed reasonable

positive significant correlation with silt content and TOMs and the negative ones were found with sand percentages and Mz.

For the different tidal marks, Pearson's correlation analysis showed strong significant values between all sediment parameters (Table 9). For nematodes, only the total number of genera showed a negative reasonable correlation with TOMs (r=-0.57, P<0.027). Deposit and predator/omnivore feeders showed strong positive significant correlations with silt and clay contents and TOMs. In contrast, epistrate feeders showed strong negative significant values with silt and clay percentages and TOMs while the strong positive significant correlations were calculated for their densities with sand contents and Mz (Table 9).

Table 8: Pearson's correlation coefficients between sediment parameters: silt content percentage %, Sand %, total organic matter (TOMs) and the median grain size (Mz), abundance of Nematoda, No. of recorded genera, predator/omnivores %, deposit feeders % and epistrate feeders % in the spatial study. ** Highly significant, *significant and NS non-significant.

Silt & clay	Sand %	TOMs	Mz
-0.99**			
0.95**	-0.95**		
-0.97**	0.97**	-0.88**	
-0.91**	0.92**	-0.79**	0.98**
-0.92	0.92	-0.79	0.98
0.63*	-0.6*	0.77**	-0.45*
NS	NS	NS	NS
-0.89**	0.89**	-0.75**	0.97**
	Silt & clay -0.99** 0.95** -0.97** -0.91** -0.91 0.63* NS -0.89**	Silt & clay Sand % -0.99** -0.95** 0.95** -0.95** -0.97** 0.97** -0.91** 0.92** -0.92 0.92 0.63* -0.6* NS NS -0.89** 0.89**	Silt & clay Sand % TOMs -0.99** -0.95** - 0.95** -0.95** - -0.97** 0.97** -0.88** -0.91** 0.92** -0.79** -0.92 0.92 -0.79 0.63* -0.6* 0.77** NS NS NS -0.89** 0.89** -0.75**

Table 9: Pearson's correlation coefficients between sediment parameters: silt content percentage %, Sand
%, total organic matter (TOMs) and the median grain size (Mz), abundance of Nematoda, No. of
recorded genera, predator/omnivores %, deposit feeders % and epistrate feeders % in the different
tidal levels. ** Highly significant, * significant and NS non-significant.

	Silt & clay	Sand %	TOMs	Mz
Sand %	-0.9**			
TOMs	-0.94**	-0.94**		
Mz	-0.99**	0.99**	-0.93**	
Abundance	NS	NS	NS	NS
No. of genera	NS	NS	-0.57*	NS
Predator/Omnivores %	0.99**	-0.99**	0.97**	-0.99*
Deposit feeder %	0.99**	-0.96**	0.96**	-0.99*
Epistrate feeder %	-0.99**	0.99**	-0.97**	0.99**

DISCUSSION

Densities of nematodes in the current study were within the range of other previous studies in mangrove regions around the world in general (their counts ranged between 35-280 in Bay of Bengal, India; Ali et al., 1983; 62-411 in South Africa, Dye 1983; 113-451 in Australia, Nicholas et al. 1991; 374-885 in Malaysia, Sasekumar, 1994; 271-656 in Zanzibar, Africa, Olafsson et al. 2000 and 196-811 in Brazil, Netto and Galucci, 2003).and in the Red Sea mangroves in particular (44-223 in Gulf of Aqaba, Jordan, Hulings, 1975; 100-130 in Red Sea, Egypt, Hanafy et al. 2011; 343 in South Sinai, Egypt, Pusceddu et al., 2014; 10-370 in Sudan; Sabeel, 2015 and 72-1100 in Red Sea, Saudi Arabia, Al-Sofiyani and El-Sherbiny, 2018). The last study in the Red Sea mangroves found higher densities of nematodes in several sites along the Saudi Arabian coast of the Red Sea than other studied mangroves. They attributed that to the nature of which are within sites situated in protected or sheltered lagoons and near fish farm effluents. Furthermore, Della Patrona et al. (2016) in New Caledonia, southwest Pacific Ocean observed that the effluent-receiving mangrove usually displays a double count in the density and biomass of nematodes due to the availability of organic matter and other nutrients. Alongi (1987a in Australia); Olafsson (1995 in Eastern Africa) stated that tidal oscillation has a negative impact on meiofaunal communities in terms of abundance and the sheltered ones are likely those areas where tidal currents have only

insignificant effects, which might be considered as possible reason of the higher abundance observed at these sites.

Water temperature, salinity, dissolved oxygen, redox potential, sediment characteristics and predator density are environmental factors that affected the diversity and abundance of meiobenthic organisms (Coull, 1999; Giere, 2009; Braeckman et al., 2011). The present study revealed a significant correlation with the granulometry of sediment and the abundances of free-living nematodes. The significance of granulometric parameters for the determination of the spatial variability of meiofauna was studied in many areas (Alongi, 1987a, b; Ansari et al., 1993; Thilagavathi et al., 2011; Semprucci et al., 2015; Al-Sofivani and El-Sherbiny, 2018). The higher availability of space in the interstices of sand substrates permits organisms to use a several niches which increase their density and diversity (Dve, 1978; Coull, 1999; Mirto et al., 2002). In addition, the sandy substrate provides a bigger scope for specialization in their feeding habits in terms of the occurrence of attached materials as well as many biogenic materials like fragments of mollusk shells (Coull and Bell, 1979; Rodriguez et al., 2003; Gheskiere et al., 2005). Although there are several parameters known to impact the variability of meiofauna, there are also quite a few studies that report the absence or lesser effect of several environmental factors on the distribution of them (Olafsson, 1995; Netto and Gallucci, 2003; Tolhurst et al., 2010; Abdullah and Lee, 2017). Tolhurst et al. (2010) clearly pointed out that microhabitat (separate niches within the same mangrove ecosystem) or other factors such as recruitment, food, predation and competition have a more influence on the distribution of meiofauna than other environmental factors.

In the current study, nematodes were represented by 13 families and 25 genera, most of which were typical of intertidal sediments worldwide. The dominant families observed (Desmodoridae Linhomoeidae and Xyalidae) are considered to be common inhabitants of detritus-rich intertidal mangroves as that found in several mangroves (Hopper et al. 1973; Alongi 1990b; Nicholas et al. 1991; Olafsson 1995; Somerfield et al. 1998; Bhadury et al. 2015; Al-Sofiyani and El-Sherbiny, 2018). Low diversity of nematodes in comparisons with other temperate and tropical mangrove ecosystems, this relatively small number can be explained either by the restricted sampling time, that covered only a specific season or by the oligotrophic nature of the Red Sea (Almahasheer et al., 2017), which is low in biodiversity. It has also been documented that tropical mangroves display basically lower diversity and abundance of meiofauna/ nematodes compared to other temperate mangroves as a result of poor nutritional quality of mangrove-derived detritus (Tietjen and Alongi 1990; Alongi and Christoffersen, 1992), increased physical stress (Alongi, 1987a, b; Armenteros et al., 2006), and high tannin content in mangrove sediments (Alongi, 1987c; Tietjen and Alongi, 1990; Abdullah and Lee, 2017). Although some of the sites were characterized by higher densities, the nematode composition did not reveal any dominance pattern in different studied habitats.

The present study found the absence of any specific free living marine genus in each mangrove habitat. This obviously shows that the composition of nematodes in the Red Sea follows the same distribution pattern and does not appear to be impacted by different habitats. Desmodoridae and Xyalidae were the most abundant and diversified families, which is in accordance with several studies (**Bhadury** *et al.*, **2015** in The Indian west coast; **Nicholas** *et al.*, **1991** in Australia, **Olafsson**, **1995** in Zanzibar, Eastern Africa; **Sabeel and Vanreusel**, **2015** in Sudan, Red Sea and **Al-Sofiyani and El-Sherbiny**, **2018** in Saudi Arabia, Red Sea) they reported similar patterns. In the present study, the dominance of certain nematode genera (*Daptonema, Spirinia* and *Terschellingia*) in mangrove sediments were observed, which is similar to the mangroves in the temperate latitudes of South America, Australia as well as tropical ones (**Hodda and Nicholas**, **1985; Nicholas** *et al.*, **1991; Olafsson**, **1995; Netto and Gallucci**, **2003; Sabeel and Vanreusel**, **2015, Al-Sofiyani and El-Sherbiny**, **2018**).

Sandy substrates are known to favor epistrate and non-selective feeders feeding on benthic diatoms, protozoans, plant roots and bacteria (Hodda and Nicholas, 1986; Jensen, 1987; Moens and Vincx, 1997; Netto and Gallucci, 2003). This is evidenced by the significant positive correlation obtained for some genera such as *Spirinia* with the sand fraction. In the current study *Spirinia* genus was common that are in accordance with several studies (Sasekumar, 1994 in Malaysia; Olafsson, 1995 and Olafsson *et al.*, 2000 in Zanzibar, Africa; Sabeel and Vanreusel, 2015 in Red Sea, Sudan; Al-Sofiyani and El-Sherbiny, 2018 in Red Sea, Saudi Arabia).

Deposit feeders were common in sediments of fine nature, while epistrate feeders are predominant in coarser sediments. This finding is in accordance with Heip et al. (1985) and Ansari et al. (2014). Strong significant correlations were found between sediment texture and the abundance of feeding types, Epistrate feeders dominated the high-water levels with coarse sediment nature and decreases seaward where the sediment texture was finer. Opposite pattern was found for deposit feeders which attained the highest occurrence in areas of low water mark. Similar findings were observed by many authors in other mangroves (Dye, 1983; Alongi, 1987 and 1990; Alongi and Christofferen, 1992; Vanhove et al., 1992; Ansari and Parulekar, 1993; Olafsson, 1995; Chinnadurai and Fernando, 2006; Ansari et al., 2014). In this study, the relative abundance of predator nematodes was high in the low water marks which are characterized by low oxygen levels and fine sediment nature. This finding agreed to that stated by Fonseca et al. (2007), Gallucci et al. (2008) and Vanreusel et al. (2010) who elucidated that predator nematodes are having elongated body (i.e. Oncholaimus, Oncholaimellus and Viscosia) which may increase their mobility, permitting short excursions in the sediments with anoxic to the oxic ones. The sediment grain size is known to be the prime factor that determines the species composition and densities of free living nematodes (Heip *et al.*, 1985; Steyaert *et al.*, 1999; Vanaverbeke *et al.*, 2002; Semprucci *et al.*, 2010; Fonseca *et al.*, 2014), as it significantly impacts the interstitial behavior and burrowing of benthic organisms (Vanaverbeke *et al.* 2011).

In the present study, the nematofauna was more abundant and diversified in the mid tidal level. The reason could be avoiding the unfavorable sediment conditions during low tide. **Steyaert** *et al.* (2001) reported that tides cause substantial alterations in sediment temperature, oxygen saturation level, and water content. These alterations produce horizontal distribution patterns and vertical stratification according to the tidal cycle. For thus, the mid tidal level zone is usually are more stable in all conditions between wave action and exposure and, therefore, shows higher in terms of abundance and diversity (Nicholas and Hodda 1999, Gheskiere *et al.*, 2004, Gingold *et al.*, 2010).

In the current study, the low tidal level zones were inhabited by lower density and diversity of nematodes than the other tidal levels. This pattern of distribution might be due to two factors; first is the presence of huge quantities of dead mangrove trees and fallen litters, which eventually leach colloidal compounds such as tannins that inhibit the growth of meiobenthic organisms (Alongi, 1987c; Tietjen and Alongi, 1990; Zhou *et al.*, 2001; Al-Sofiyani and El-Sherbiny, 2018). The amount of these compounds varied according to several ambient environmental factors such as water temperature, salinity and the residence time of water at these sheltered areas. The second factor is the predations that occur exclusively at this zone and is generated by the feeding actions of vast animals and can cause significant reductions in nematode densities. Similar interpretations were reported by Olafsson and Elmgren (1991); Dittmann (1996); Schratzberger and Warwick (1999); Danovaro *et al.* (2007).

CONCLUSION

The distribution of free-living nematodes in the mangrove ecosystems of the Egyptian western coast of the Red Sea is influenced by vast environmental factors. Sediment texture was directly and indirectly affecting their distributions in terms of generic compositions and types of feedings. The distribution of these organisms at different tidal levels was more complicated due to the wide variations in all conditions that provide specific microhabitats in each zone.

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REFERENCES

- Abdullah, M. and Lee, S. (2017). Structure of mangrove meiofaunal assemblages associated with local sediment conditions in subtropical eastern Australia. Estuar. Coast. Shelf. Sci., 198: 438–449.
- Almahasheer, H.; Serrano, O.; Duarte, C.M.; Arias-Ortiz, A. and Masque, P. (2017). Low Carbon sink capacity of Red Sea mangroves. Sci. Rep., 7(1): 9700.
- Alongi, D. and Christofferson, P. (1992). Benthic infauna and organism-sediment relations in a shallow, tropical coastal area: influence of out welled mangrove detritus and physical disturbance. Mar. Ecol. Prog. Ser., 81(3): 229–245.
- Alongi, D. (1986). Population structure and trophic composition of the free-living nematodes inhabiting carbonate sands of Davies Reef, Great Barrier Reef, Australia. Aust. J. Mar. Freshwat. Res., 37: 609-619.
- Alongi, D. (1987a). Inter-estuary variation and intertidal zonation of free-living nematode communities in tropical mangrove systems. Mar. Ecol. Prog. Ser., 40: 103–114.
- Alongi, D. (1987b). Intertidal zonation and seasonality of meiobenthos in tropical mangrove estuaries. Mar. Biol. 95(3): 447–458.
- Alongi, D. (1987c). The influence of mangrove-derived tannins on intertidal meiobenthos in tropical estuaries. Oceanolog. 71: 537–540.
- Alongi, D. (1989). The role of soft-bottom benthic communities in tropical mangrove and coral reef ecosystems. Crit. Rev. Aquat. Sci., 1: 243–280.
- Alongi, D. (1990a). The ecology of tropical soft-bottom benthic ecosystems. Oceanogr. Mar. Biol. Annu. Rev., 28:381–496.
- Alongi, D. (1990b). Community dynamics of free-living nematodes in some tropical mangrove and sandflat habitats. Bull. Mar. Sci., 46: 358–373.
- Al-Sofyani, A. and Al-Sherbiny, M. (2018). Meiobenthic assemblage of the grey mangrove (*Avicennia marina*) along the Saudi Arabian coast of the Red Sea with emphasis on free-living nematodes. Oceanolog. and Hydrobiol. St., 47(4): 359-375.
- Ansari, K. and Bhadury, P. (2017). An updated species checklist for free-living marine nematodes from the world's largest mangrove ecosystem, Sundarbans. Zootaxa. 4290(1): 177–191.
- Ansari, K.; Manokaran, S.; Raja, S.; Lyla. P. and Ajmal Khan, S. (2014). Interaction of free-living marine nematodes in the artificial mangrove environment (southeast of India). Environ. Monit. Assess., 186:293-305.
- Ansari, Z.; Sreepada, R.; Matondkar, S. and Parulekar, A. (1993). Meiofauna stratification in relation to microbial food in a tropical mangrove mudflat. Trop. Ecol. 34: 63–75.
- Ansari. Z. and Parulekar, A. (1994). Meiobenthos in the sediment of seagrass meadows of Lakshadweep atolls, Arabian Sea. Vie Milieu., 44: 185–190.

- Armenteros, M.; Martın, I.; Williams, J.P.; Creagh, B. and Gonzalez- Sanson, G. (2006). Spatial and temporal variations of meiofaunal communities from the western sector of the Gulf of Batabano Cuba. I. Mangrove Systems. Estuar. and Coas., 29: 124–132.
- Bhadury, P.; Mondal, N.; Ansari, K.; Philip, P. and Pitale, R. (2015). Checklist of free-living marine nematodes from intertidal sites along the central west coast of India. Check List 11(2): 1605.
- Braeckman, U.; Van Colen, C.; Soetaert, K.; Vincx, M. and Vanaverbeke, J. (2011). Contrasting macrobenthic activities differentially affect nematode density and diversity in a shallow subtidal marine sediment. Mar. Ecol. Prog. Ser., 422: 179–191.
- Chinnadurai, G. and Fernando, O. (2006). Meiobenthos of Cochin mangroves (Southwest coast of India) with special emphasis on free-living marine nematode assemblages. Russ. J. Nematol., 64(2): 127–137.
- Chinnadurai, G. and Fernando, O. (2007). Meiofauna of mangroves of the southeast coast of India with special reference to the free-living marine nematodes assemblage. Estuar. Coast. Shelf Sci. 72: 329–336.
- Clarke, K. and Gorley, R. (2006). Primer v6: User Manual/Tutorial. Primer-E Ltd, Plymouth.
- Coull, B. and Bell, S. (1979). Perspectives of marine meiofaunal ecology. In R.J. Livingston (Eds.), Ecological Processes in Coastal and Marine Systems. Mar. Sci., Vol 10: 189–216. Springer, Boston, MA.
- **Coull, B.** (1985). Long-term variability of estuarine meiobenthos: an 11-year study. Mar. Ecol. Prog. Ser., 24: 205–218.
- **Coull, B.** (1999). Role of meiofauna in estuarine soft-bottom habitats. Austral Ecol., 24(4): 327–343.
- **Danovaro, R. and Gambi, C.** (2002). Biodiversity and trophic structure of nematode assemblages in seagrass system: evidence for a coupling with changes in food availability. Mar. Biol., 141: 667–677.
- Danovaro, R.; Gambi, C. and Mirto, S. (2002). Meiofaunal production and energy transfer efficiency in a seagrass *Posidonia oceanica* bed in the western Mediterranean. Mar. Ecol. Prog. Ser., 234: 95–104.
- Danovaro, R.; Scopa, M.; Gambi, C. and Fraschetti, S. (2007): Trophic importance of subtidal metazoan meiofauna: evidence from in situ exclusion experiments on soft and rocky substrates. Mar. Biol., 152: 339 – 350.
- Della Patrona, L.; Marchand, C.; Hubas, C.; Molnar, N. and Deborde, J. (2016). Meiofauna distribution in a mangrove forest exposed to shrimp farm effluents (New Caledonia). Mar. Environ. Res., 119: 100–113.
- **Dittmann, S.** (1996): Effects of macrobenthic burrows on infaunal communities in tropical tidal flats. Mar. Ecol. Prog. Ser., 134: 119 130.

- **Dye, A.** (1978). An ecophysiological study of the meiofauna of the Swartkops Estuary. 2. The meiofauna: composition, distribution, seasonal fluctuation, and biomass. Afr. Zool., 13: 19–32.
- **El-Serehy, H.; Al-Misned, F. and Al-Rasheid, K.** (2015). Population fluctuation and vertical distribution of meiofauna in the Red Sea interstitial environment. Saudi J. of Biol. Sci., 22: 459-465.
- Fonseca, G.; Muthumbi, A. and Vanreusel, A. (2007): Species richness of the genus Molgolaimus (Nematoda) from local to ocean scale along continental slopes. Mar. Ecol., 28: 446 – 459.
- Fonseca, G.; Hutchings, P. and Gallucci, F. (2011). Meiobenthic communities of seagrass beds (Zostera capricorni) and unvegetated sediments along the coast of New South Wales, Australia. Estuar. Coast. Shelf Sci., 91: 69–77.
- Fonseca, V., Carvalho, G., Nichols, B., Quince, C., Johnson, H. (2014). Metagenetic analysis of patterns of distribution and diversity of marine meiobenthic eukaryotes. Glob. Ecol. Biogeogr., 23: 1293–1302.
- Gallucci, F.; Moens, T.; Vanreusel, A. and Fonseca, G. (2008b): Active colonization of disturbed sediments by deep-sea nematodes: evidence for the patch mosaic model. Mar. Ecol. Prog. Ser., 367: 173 – 183.
- Gee, J. and Somerfield, P. (1997). Do mangrove diversity and leaf litter decay promote meiofaunal diversity? J. Exp. Mar. Biol. Ecol., 218(1): 13–33.
- Gee, J. (1989). An ecological and economic review of meiofauna as food for fish. Zool. J. Linn. Soc., 96: 243–261.
- Gheskiere, T.; Hoste, E.; Vanaverbeke, J.; Vincx, M. and Degraer, S. (2004): Horizontal zonation patterns and feeding structure of marine nematode assemblages on a macrotidal, ultra-dissipative sandy beach (De Panne, Belgium). J. Sea Res., 52: 221–226.
- Gheskiere, T.; Vincx, M.; Urban-Malinga, B.; Rossano, C. and Scapini, F. (2005). Nematodes from wave-dominated sandy beaches: diversity, zonation patterns and testing of the isocommunities concept. Estuar. Coast. Shelf. Sci., 62(1– 2):365–375.
- Giere, O. (2009) Meiobenthology: the microscopic motile fauna of aquatic sediments, 2nd edn. Springer-Verlag, Berlin.
- **Gingold, R.; Mundo-Ocampo, M.; Holovachov, O. and Rocha-Olivares, A.** (2010): The role of habitat heterogeneity in structuring the community of intertidal freeliving marine nematodes. Mar. Biol., 157: 1741–1753.
- Hall, M, and Bell, S. (1993). Meiofauna on the seagrass Thalassia testudium population characteristics of harpacticoid copepods and associations with algal epiphytes. Mar. Biol., 116: 137–146.

- Hanafy, M.; Mohammed, D. and Ahmed, A. (2011). Seasonal distribution of the littoral interstitial meiofauna in the northern Red Sea, Egypt. Egypt. J. Aquat. Biol. & Fish., 15(2): 35-51.
- Heip, C.; Vincx, M. and Vranken, G. (1985). The ecology of marine nematodes. Oceanogr. Mar. Biol. Ann. Rev. 23: 399–489.
- Hodda, M. and Nicholas, W. (1986). Temporal changes in littoral meiofauna from the Hunter River Estuary. Aust. J. Mar. Freshwat. Res., 37: 729–741.
- **Hodda, M.** (1990). Variation in estuarine littoral nematode populations over three spatial scales. Estuar. Coast. Shelf. Sci., 30: 325–340.
- Hodda, M. and Nicholas, W.L. (1985). Meiofauna associated with mangroves in the Hunter River estuary and Fullerton Cove, south-eastern Australia. Aust. J. Mar. Freshwat. Res., 36: 41– 50.
- Holme, N. and McIntyre, A. (1984). Methods for the Study of Marine Benthos, Second ed. Blackwell, Oxford, pp. 387.
- Hopper, B. and Meyers, S. (1967). Population studies on benthic nematodes within a subtropical seagrass community. Mar. Biol., 1: 85–96.
- Hopper, B. E.; Fell, J. W. and Cefalu, R. C. (1973). Effect of temperature on life cycles of nematodes associated with the mangrove (Rhizophora mangle) detrital system. Mar. Biol., 23(4): 293–296.
- Hulings, N. (1975). Spatial and quantitative distribution of sand beach meiofauna in the northern Gulf of Aqaba. Rapp. Comm. Int. Mer. Medit., 23: 163–181.
- Jensen, P. (1987). Feeding ecology of free-living aquatic nematodes. Mar. Ecol. Prog. Ser., 35: 187–196.
- Kathiresan, K.; Rajendran, N. and Thangadurai, G. (1996). Growth of mangrove scalding in the intertidal area of Vellar estuary, Southeast coast of India. Indian J. of Mar. Sci., 25: 240–243.
- McLachlan, A.; Erasmus, T. and Furstenberg, J. (1977). Migrations of sandy beach meiofauna. Zool. Afr., 12: 257–277.
- Mirto, S.; La Rosa, T.; Gambi, C.; Danovaro, R. and Mazzola, A. (2002). Nematode community response to fish-farm impact in the western Mediterranean. Environ. Pollut., 116(2): 203–214.
- Moens, T. and Vincx, M. (1997). Observations on the feeding ecology of estuarine nematodes. J. Mar. Biol. Ass. U.K., 77(1): 211–227.
- Ndaro, S. and Ólafsson, E. (1999). Soft-bottom fauna with emphasis on nematode assemblage structure in a tropical intertidal lagoon in Zanzibar, eastern Africa: I. Spatial variability. Hydrobiol., 405: 133–148.
- **Netto, S. and Gallucci, F.** (2003). Meiofauna and macrofauna communities in a mangrove from the Island of Santa Catarina, South Brazil. Hydrobiol., 505: 159–170.

- Nicholas, W.; Elek, J.; Stewart, A. and Marples, T. (1991). The nematode fauna of a temperate Australian mangrove mudflat; its population density, diversity, and distribution. Hydrobiol. 209: 13–27.
- Nicholas, W. L. and Hodda, M. (1999). Free-living nematodes of a temperate, highenergy sandy beach: faunal composition and variation over space and time. Hydrobiolo., 394: 113 – 127.
- **Olafsson, E. and Moore, C.** (1990). Control of meiobenthic abundance by macroepifauna in a subtidal muddy habitat. Mar. Ecol. Prog. Ser., 65(3): 241–249.
- **Olafsson, E.** (1995). Meiobenthos in mangrove areas in eastern Africa with emphasis on assemblage structure of free-living marine nematodes. Hydrobiol., 312(1): 47–57.
- **Olafsson, E. and Elmgren, R.** (1991): Effects of biological disturbance by benthic amphipods *Monoporeia affinis* on meiobenthic community structure: a laboratory approach. Mar. Ecol. Prog. Ser., 74: 99 107.
- Platt, H. and Warwick, R. (1988). Free-living marine nematodes. Part II. British chromadorids, Brill/Backhuys, Leiden Sanders HL (1968) Marine benthic diversity: a comparative study. Am. Nat., 102: 243–282.
- Pusceddu, A.; Gambi, C.; Corinaldesi, C.; Scopa, M. and Danovaro, R. (2014). Relationships between meiofaunal biodiversity and prokaryotic heterotrophic production in different tropical habitats and oceanic regions. PLoS ONE 9(3):e91056
- Rodriguez, J.; Lastra, M. and Lopez, J. (2003). Meiofauna distribution along a gradient of sandy beaches in northern Spain. Estuar. Coast Shelf Sci., 58: 63–69.
- Sabeel, R. and Vanreusel, A. (2015). Potential impact of mangrove clearance on biomass and biomass size spectra of nematode along the Sudanese Red Sea coast. Mar. Environ. Res., 103: 46-55.
- Sabeel, R. (2015). Variation in distribution of Sudanese mangroves and their ecological significance for benthic fauna. Unpublished doctoral dissertation, Ghent University, Ghent, Belgium.
- Sabeel, R. and Vanreusel, A. (2015). Potential impact of mangrove clearance on biomass and biomass size spectra of nematode along the Sudanese Red Sea coast. Mar. Environ. Res., 103: 46–55.
- Sasekumar, A. (1994). Meiofauna of a mangrove shore on the west coast of peninsular Malaysia. Raffles Bull. Zool., 42(4): 901–915.
- Semprucci, F.; Colantoni, P.; Baldelli, G.; Rocchi, M. and Balsamo, M. (2010). The distribution of meiofauna on back-reef sandy platforms in the Maldives (Indian Ocean). Mar. Ecol., 31(4): 592–607.

- Semprucci, F.; Sbrocca, C.; Rocchi, M. and Balsamo, M. (2015). Temporal changes of the meiofaunal assemblage as a tool for the assessment of the ecological quality status. J. Mar. Biol. Assoc. U.K., 95(2): 247–254.
- Schratzberger, M. and Warwick, R. M. (1999). Impact of predation and sediment disturbance by *Carcinus maenas* (L.) on free-living nematode community structure. J. Exp. Mar. Biol. Ecol., 235: 255 – 271.
- Somerfield, P.; Gee, J. and Aryuthaka, C. (1998). Meiofaunal communities in a Malaysian mangrove forest. J.Mar. Biol. Assoc. U.K., 78(3): 717–732.
- Steyaert, M.; Garner, N.; van Gansbeke, D. and Vincx, M. (1999). Nematode communities from the North Sea: environmental controls on species diversity and vertical distribution within the sediment. J. Mar. Biol. Assoc. U.K., 79(2): 253–264.
- Steyaert, M.; Vanaverbeke, J.; Vanreusel, A.; Barranguet, C.; Lucas, C. and Vincx, M. (2003). The importance of fine-scale, vertical profiles in characterising nematode community structure. Estuar. Coast. Shelf Sci., 58: 353–366.
- **Tarjan, A.** (1980). Illustrated guide to the marine nematodes. IFAS, Univ. of Florida, Gainesville. 135 pp.
- Thilagavathi, B.; Das, B.; Saravanakumar, A. and Raja, K. (2011). Benthic meiofaunal composition and community in the Sethukuda mangrove area and adjacent open sea, east coast of India. Ocean Sci. J., 46(2): 63–72.
- **Tietjen, J. and Alongi, D.** (1990). Population growth and effects of nematodes on nutrient regeneration and bacteria associated with mangrove detritus from northeastern Queensland (Australia). Mar. Ecol. Prog. Ser., 68(1–2): 169–180.
- Vanaverbeke, J. (2011). Contrasting macrobenthic activities differentially affect nematode density and diversity in a shallow subtidal marine sediment. Mar. Ecol.Prog. Ser., 422: 179–191.
- Vanaverbeke, J.; Gheskiere, T.; Steyaert, M. and Vincx, M. (2002). Nematode assemblages from subtidal sandbanks in the Southern Bight of the North Sea: effect of small sedimentological differences. J. Sea Res., 48(3): 197– 207.
- Vanaverbeke, J.; Merckx, B.; Degraer, S. and Vincx, M. (2011). Sediment-related distribution patterns of nematodes and macrofauna: Two sides of the benthic coin? Mar. Environ. Res., 71(1): 31–40.
- Vanhove, S.; Vincx, M.; Vangansbeke, W.; Gijselinck, W. and Schram, D. (1992). The meiobenthos of five mangrove vegetation types in Gazi Bay, Kenya. Hydrobiol., 247: 99–108.
- Vanreusel, A.; De Groote, A.; Gollner, S. and Bright, M. (2010): Ecology and biogeography of free-living nematodes associated with chemosynthetic environments in the deep sea: a review. PLoS One 5: e12449.

- Warwick, R.; Platt, H. and Somerfield, P. (1998). Free living marine nematodes. Part III. British Monhysterida. Synopses of the British fauna no 53. Field Studies Council, Shrewsbury.
- Wieser, W. (1953). Die Beziehung zwischen Mundhohlengestalt, Ernahrungsweise und Vorkommen bei freilebenden marinen Nematoden. Ark. Zool., 4(2): 439–484.
- Zhou, H. (2001). Effects of leaf litter addition on meiofaunalcolonization of azoic sediments in a subtropical mangrove in Hong Kong. J. Exp. Mar. Biol. Ecol., 256(1): 99–121.